**Research Perspective** 

### Cdk2: a key regulator of the senescence control function of Myc

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Abstract: Proto-oncogenes such as MYC and RAS promote normal cell growth but fuel tumor development when deregulated. However, over-activated Myc and Ras also trigger intrinsic tumor suppressor mechanisms leading to apoptosis and senescence, respectively. When expressed together MYC and RAS are sufficient for oncogenic transformation of primary rodent cells, but the basis for their cooperativity has remained unresolved. While Ras is known to suppress Myc-induced apoptosis, we recently discovered that Myc is able to repress Ras-induced senescence. Myc and Ras thereby together enable evasion of two main barriers of tumorigenesis. The ability of Myc to suppress senescence was dependent on phosphorylation of Myc at Ser 62 by cyclin-dependent kinase 2 (Cdk2), uncovering a new non-redundant role of this kinase. Further, utilizing Cdk2 as a cofactor, Myc directly controlled key genes involved in senescence. We speculate that this new role of Myc/Cdk2 in senescence has relevance for other Myc functions, such as regulation of stemness, self-renewal, immortalization and differentiation, which may have an impact on tissue regeneration. Importantly, selective pharmacological inhibition of Cdk2 forced Myc/Ras expressing cells into cellular senescence, highlighting this kinase as a potential therapeutic target for treatment of tumors driven by Myc or Ras.

### Apoptosis and cellular senescence are two main barriers to cancer development

Cancer is in part driven by aberrant activation of oncogenes. growth-promoting However, overstimulation of cell growth by oncogenic signals causes severe cellular stress, which induces intrinsic tumor suppressor mechanisms resulting in apoptosis or cellular senescence [1]. Apoptosis is a programmed, energydependent cellular suicide process [2], while cellular senescence is a state of irreversible cell cycle arrest occurring in active, viable cells as a result of telomere erosion, DNA damage, hypoxia, oncogene activation or aging. Telomere erosion leads to replicative senescence as cells exhaust their proliferative capacity, while acute stress caused by oncogene activity can lead to premature, so called oncogene-induced senescence.

Cellular senescence has been demonstrated in different types of premalignant tumor cells *in vivo* during recent years, and has emerged as an important tumor-suppressive mechanism [3,4,5,6,7].

*MYC* and *RAS* are two prototypic oncogenes involved in development of numerous cancers. *MYC* encodes a pleiotropic transcription factor [8,9], while *RAS* codes for a signal-transducing GTPase [10]. However, Myc and Ras affect the failsafe mechanisms mentioned above in quite different ways. While Myc primarily induces apoptosis, Ras usually triggers cellular senescence. Myc-induced apoptosis is brought about though activation of p19Arf [11], which controls turnover of the tumor suppressor protein/transcription factor p53. p53 is a main executor of both apoptosis and senescence by controlling a large number of genes involved in these processes [12]. Myc can also induce apoptosis by suppressing anti-apoptotic members of the Bcl-2 family [9]. Mutations interfering with apoptotic signaling/execution are strongly selected for in Mycdriven tumorigenesis.

Ras on the other hand mainly triggers oncogeneinduced senescence, which also involves upregulation of p53 via Arf [13,14]. In this case, anti-proliferative p53 target genes dominate, including p21Cip1, an inhibitor of cyclin-dependent kinase 2 (Cdk2) and Cdk1. Independent of p53, Ras also upregulates another Cdk inhibitor, p16Ink4, which targets Cdk4/6. The Cdks are the engines that drive the cell cycle by phosphorylating various substrates involved in this process. For instance, Cdk4/6 and Cdk2 target and inactivate the retinoblastoma tumor suppressor protein (pRb), an important "gatekeeper" that controls G1-S phase transition [15]. The Arf/p53/p21 and p16/pRb pathways therefore cooperatively regulate induction of senescence, and usually both of these pathways need to be intact for maintaining the senescent state [6,7].

In addition, Myc and Ras can also cause DNA damage, for instance by generating replication stress or reactive oxygen species (ROS) [13,16,17,18]. This induces DNA damage responses (DDR) that in turn can trigger apoptosis or cellular senescence. Depending on the type of lesion, DDR can cause activation of p53 via pathways involving the checkpoint kinases ATM and Chk2 or ATR and Chk1, which in turn induces p53 [19,20].

It is still unclear why oncogenic stress sometimes cause apoptosis and in other situations senescence. A possible explanation why Ras usually triggers senescence is that it activates the PI3/Akt kinase pathway, which dampens apoptotic signaling by inhibiting GSK3 kinase, FoxO and Bcl-2 family proteins [21]. Conversely, the reason why Myc primarily induces apoptosis may be because it directly or indirectly represses the p21 and p16 Cdk inhibitors as well as several anti-apoptotic genes [22,23,24,25,26], thereby possibly favoring apoptosis over senescence.

# Myc suppresses Ras-induced senescence with the help of Cdk2

It has been known since the early 80s that the combined activities of Myc and Ras are sufficient for oncogenic transformation of primary rodent cells [10]. The basis for the cooperativity has, however, remained unclear. One possibility is that they complement each other's capacity to induce mitogenic signals. Another possibility, that we have been exploring, is that the two oncogenes cooperate in suppressing the intrinsic tumor

suppressor mechanisms described above. Ras is known from previous work to be able to suppress Myc-induced apoptosis through the PI3K/Akt pathway [27]. We recently demonstrated that Myc can repress Rasinduced senescence in primary rat embryo fibroblasts [24]. This suggests that at least part of Myc and Ras cooperativity is based on a "cross-pollination" mechanism where Myc abrogates the predominating tumor suppressing activity of Ras and vice versa, thereby together overriding two major barriers to tumor development (Figure 1A). Myc has previously been implicated in suppressing replicative senescence, which is caused by telomere erosion, by activating transcription of hTERT [28]. hTERT encodes a major component of telomerase, which is normally expressed in stem cells and prevents telomere shortening, thereby extending the replicative lifespan of cells. Deletion of this gene inhibits Myc-driven lymphoma development, correlating with increased senescence [29]. Further, there are several reports demonstrating that reduced Myc levels induce cellular senescence in different settings. Lowering the Myc level by heterozygous knockout triggered telomere-independent senescence in human fibroblasts [26]. An elegant report using mouse tumor models with regulatable Myc showed that Myc shut-off caused regression of lymphomas, osteosarcomas and hepatocellular carcinomas, primarily as a result of increased cellular senescence [30]. Moreover, knockdown of Myc caused senescence in BRAF<sup>V600E</sup>or NRAS  $^{\rm Q61R}\mbox{-}driven$  melanoma cells, while overexpression of Myc suppressed BRAF  $^{\rm V600E}\mbox{-}induced$ NRAS<sup>Q61R</sup>-driven senescence in primary melanocytes, [31]. Taken together, these reports suggest that Myc suppresses oncogene-induced as well as replicative senescence.

How does Myc suppress oncogene-induced senescence? Our recent work suggests that Cdk2 plays an important role in this regulation [24]. We could show that Cdk2mediated phosphorylation of Myc at Ser-62 was crucial for bypassing Ras-induced senescence (Figure 1A). Interestingly, Cdk2 bound and phosphorylated Myc at promoter regions of target genes involved in regulation of cellular senescence such as p21, p16, BMI1, CYCLIN D2 and hTERT. This correlated with low expression of p21 and p16 and high expression of BMI1, CYCLIN D2 and hTERT [24]. The former and latter categories of genes are linked to activation and suppression of senescence, respectively. Importantly, inhibition of Cdk2 activity by selective pharmacological compounds or through interferon-y-mediated upregulation of the endogenous Cdk inhibitor p27Kip1 abolished Ser-62 This correlated phosphorylation. with induced expression of p21 and p16, repressed expression of BMI1, hTERT and CYCLIN D2 and induction of senescence [24] (Figure1B). These results suggest that



**Figure 1. Cdk2 controls suppression of cellular senescence by Myc.** (A) Apoptosis and senescence are two intrinsic tumor suppressor mechanism that are triggered by oncogenic signals. Myc and Ras contributes to this by inducing apoptosis and senescence, respectively. However, activated Myc and Ras are together sufficient to transform primary rodent cells for unclear reasons. We recently found that while Ras suppresses Myc-induced apoptosis, Myc is able to suppress Ras-induced senescence, thereby together overcoming two main barriers of tumorigenesis. In order to suppress senescence Myc needs to be phosphorylated by Cdk2. Since Myc stimulates Cdk2 activity, Myc and Cdk2 are involved in an auto-stimulatory loop generating suppression of senescence. (B) Depletion or inhibition of Cdk2 abolishes to ability of Myc to suppress senescence and switches Myc into an activator of senescence, thereby inhibiting tumor development or maintenance.

Myc utilizes Cdk2 as a cofactor to directly control key genes in the p53/p21 and p16/Rb pathways and is thereby able to suppress senescence.

Although MAPK and Cdk1 are also reported to target Ser-62 [32], the anti-senescence function of Cdk2 could not be compensated by these kinases for unclear reasons. One explanation for this unique function of Cdk2 could be that it potentially also target additional proteins associated with Myc-regulated transcription, such as other transcription factors, cofactors or chromatin regulating proteins. Further, one cannot exclude that the Cdk2 function at chromatin synergizes with other unique but non-redundant, nontranscriptional functions of Cdk2, such as phosphorylation of p27.

What is the function of phospho-Ser-62 in suppression of senescence? Phosphorylation of Ser 62 is known to prime for GSK3-mediatied phosphorylation of Thr-58, which regulates the apoptosis function of Myc [33] and also its ubiquitylation and degradation [32]. However, a T58A mutant had no effect on senescence, suggesting that senescence regulation is a new and independent role of Ser-62. We did observe reduced association of Myc with target promoters upon Cdk2 inhibition, indicating that phospho-Ser-62 stabilizes Myc binding to chromatin. This is consistent with the work of Benassi et al [34], which demonstrated that MAPKmediated phosphorylation of Ser-62 increased association of Myc to the  $\gamma$ -GCS gene in response to oxidative stress. Another plausible option is that phospho-Ser 62 provides an interaction surface for

recruitment of a cofactor that participates in regulation of senescence-related Myc target genes.

In a parallel investigation together with the lab of Bruno Amati, we examined the impact of Myc alone, i.e. in the absence of other activated oncogenes, on cellular senescence in murine embryonic fibroblasts (MEFs). Strikingly, Myc activation resulted in senescence induction in Cdk2 knockout but not in wt MEFs, suggesting that Cdk2 suppresses Myc-induced senescence [35] (Figure 1B). Since Cdk2 function in the cell cycle is compensated by other Cdks during development [15], this is a unique, non-redundant role of Cdk2. It has been shown previously that Myc can induce senescence in cells lacking the Werner syndrome protein (WRN), a helicase implicated in DNA repair [36]. In Cdk2-/- cells, Myc-induced senescence was dependent on intact Arf-p53/p21 and p16<sup>INK4a</sup>-pRB pathways, and seemed to involve DDR [35], i.e. essentially the same pathways engaged by Ras [13,19,20]. One possible interpretation of the combined Hydbring and Campaner results is that Cdk2 and Myc constitute a senescence switch; Myc acts as a repressor of senescence when Cdk2 is active, but will provoke induction of senescence when Cdk2 is inactive (Figure 1). However, it is unclear at present whether the role of Cdk2 in suppression of Myc- and Ras-induced senescence is similar or distinct. Interestingly, both Cdk2 and WRN have been implicated in DNA repair [36,37], and could thereby possibly play a role in prevention and termination of the persistent DDR signaling that characterizes oncogene-induced senescence. Since Myc upregulates expression of WRN and hTERT and stimulates the activity of Cdk2 [8,28,36], it is conceivable that these proteins are part of an auto-protective loop that Myc uses to suppress senescence, perhaps at multiple levels. It will be important for future studies to identify Cdk2 substrates (other than Myc) that are essential for this process.

Regardless of mechanism, selective pharmacologic inhibitors of Cdk2, but not of other Cdks, forces embryonic fibroblasts with deregulated Myc or Myc/Ras into senescence [24,35] (Figure 1B). Further, Cdk2 ablation induced senescence in both pancreatic  $\beta$ -cells and hematopoietic B-cells after Myc activation, the latter correlating with delayed lymphoma development. This underscores that Cdk2 inhibitors should be reassessed as therapeutic agents, especially for Myc- or Ras-driven tumors. Cdk2 may be particularly suited for pharmacologic intervention since its function in the cell cycle is compensated by other Cdks in normal cells [38]. Previously, inhibition of Cdk1 was demonstrated to induce apoptosis in Myctransformed cell but not in cells transformed with other oncogenes, and caused regression of Myc-driven tumors [39]. Therefore inhibition of Cdk1, Cdk2 or combination of these regimens should be considered in future treatment of Myc driven tumors based on molecular diagnosis of genetic and epigenetic status of intrinsic tumor suppressor systems of the tumor cells (Figure 1B).

# How does senescence regulation relate to other functions of Myc?

Cellular senescence is defined as an irreversible exit from the cell cycle - a feature that it shares with terminally differentiated cells. Myc's suppression of senescence therefore in many ways resembles its wellknown function in inhibiting of terminal differentiation. which occurs in most (but not all) studied cell types [8,9]. One might consider inhibition of senescence and differentiation versus immortalization, self-renewal and tissue regeneration as two sides of the same coin (Figure 2). Myc is known since many years to contribute to immortalization of cells, for instance by supporting hTERT expression. Further, an increasing amount of data during recent years suggests that Myc is involved in the regulation of stemness. c-Myc and N-Myc, another Myc-family member, are essential for maintaining pluripotency and self-renewal of embryonic, hematopoietic and neuronal stem cells and progenitors [40,41,42]. This could also have an impact on processes like aging and aging-related diseases. Although the relation between cellular senescence and aging is far from clear, increased cellular senescence within stem cell populations could be one factor contributing to reduced tissue regeneration with age. In support of this, telomerase-deficient mice as well as mice expressing an overactive mutant of p53 have a significantly shortened lifespan [43,44,45]. DNA damage as well as p16Ink4a expression is known to increase with age, correlating with decreased tissue renewal and loss of stem cell function. This has been confirmed by deletion or overexpression of p16 in hematopoietic stem cells. forebrain neuronal progenitors and pancreatic  $\beta$ -cells [43,46,47,48]. The increased expression of p16 in aging tissues may in part be the result of decreased expression of Bmi1 [49], a major repressor of the p16Ink4a locus (Figure 2). One should note that Bmil is positively regulated by Myc [24,26].

This raises the question whether Myc and Cdk2 contribute to renewal and regeneration of adult tissues under normal conditions, and from this point of view could be considered as anti-aging factors? On the other hand, expression of these genes occurs at the expense of increased cancer risk and may therefore not result in increased longevity (Figure 2). It also raises the question whether combating Myc- and Myc/Ras-driven tumors with Cdk2-inhibitors comes with the side-effects of stem cell failure, decreased regeneration capacity in normal tissues and increased aging? We think this is rather unlikely since Cdk2 inhibition in our hands induced senescence only in primary cells over-expressing Myc or Myc/Ras, but had little effect on untransfected cells.

One should also note that other reports do not support the notion that pro-senescence/anti-proliferative factors decrease regeneration capacity or promote aging under normal regulation. The proliferation rate of stem cells is normally kept low to protect them from the potential hazards of increased metabolism and cell division, and too high proliferation rate may lead to stem cell depletion, favoring the view that pro-senescence factors may have a protective role for stem cells [50]. In support of this notion, mice carrying an extra copy of the *p53* or *Arf/Ink4* loci display increased protection against cancer without any change in aging [43], and mice carrying an additional copy of both these loci displayed delayed aging, correlating with decreased aging-related DNA-damage [51], suggesting that Myc/Cdk2 inhibition not necessarily would have a negative impact on tissue regeneration and longevity. Interestingly, exogenous expression of hTERT in mice carrying the extra p53/Arf/p16 loci delayed aging even further [52].

In conclusion, there seems to be an intricate balance between anti- and pro-senescence factors in the regulation of stem cell capacity, regeneration of tissues and aging. The impact of Myc and Cdk2 inhibition both on tumor regression through senescence and potential effects on regenerative capacity of normal tissue needs to be addressed in mouse models in the future.



**Figure 2.** Speculative model illustrating regulation of senescence vs stemness and self-renewal by **Myc and Cdk2.** While Myc is a suppressor of senescence together with Cdk2, it stimulates stemness, self-renewal and immortalization, thereby potentially favor tissue regeneration. This is accomplished by activation of *hTERT* and *Bmi1*, and repression of *p21* and *p16*, key genes also implicated in regulating aging. The trade off for this capacity is increased risk for cancer development.

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### CONFLICT OF INTERESTS STATEMENT

The authors of this manuscript have no conflict of interests to declare.

#### REFERENCES

**1.** Lowe SW, Cepero E, Evan G. Intrinsic tumour suppression. Nature. 2004; 432: 307-315.

**2.** Cotter TG. Apoptosis and cancer: the genesis of a research field. Nat Rev Cancer. 2009; 9: 501-507.

**3.** Collado M, Gil J, Efeyan A, Guerra C, Schuhmacher AJ, et al. Tumour biology: senescence in premalignant tumours. Nature. 2005; 436: 642.

**4.** Braig M, Lee S, Loddenkemper C, Rudolph C, Peters AH, et al. Oncogene-induced senescence as an initial barrier in lymphoma development. Nature. 2005; 436: 660-665.

**5.** Michaloglou C, Vredeveld LC, Soengas MS, Denoyelle C, Kuilman T, et al. BRAFE600-associated senescence-like cell cycle arrest of human naevi. Nature. 2005; 436: 720-724.

**6.** Campisi J and d'Adda di Fagagna F. Cellular senescence: when bad things happen to good cells. Nat Rev Mol Cell Biol. 2007; 8: 729-740.

**7.** Collado M, Serrano M Senescence in tumours: evidence from mice and humans. Nat Rev Cancer. 2010; 10: 51-57.

**8.** Grandori C, Cowley SM, James LP, Eisenman RN. The Myc/Max/Mad network and the transcriptional control of cell behavior. Annu Rev Cell Dev Biol. 2000; 16: 653-699.

**9.** Larsson LG, Henriksson MA The Yin and Yang functions of the Myc oncoprotein in cancer development and as targets for therapy. Exp Cell Res. 2010; 316: 1429 - 1437.

**10.** Karnoub AE, Weinberg RA. Ras oncogenes: split personalities. Nat Rev Mol Cell Biol. 2008; 9: 517-531.

**11.** Zindy F, Eischen CM, Randle DH, Kamijo T, Cleveland JL, et al. Myc signaling via the ARF tumor suppressor regulates p53dependent apoptosis and immortalization. Genes Dev. 1998; 12: 2424-2433.

**12.** Vousden KH, Lane DP. p53 in health and disease. Nat Rev Mol Cell Biol. 2007; 8: 275-283.

13. Serrano M, Lin AW, McCurrach ME, Beach D, Lowe SW. Oncogenic ras provokes premature cell senescence associated with accumulation of p53 and p16INK4a. Cell. 1997; 88: 593-602.
14. Schmitt CA, Fridman JS, Yang M, Lee S, Baranov E, et al. A senescence program controlled by p53 and p16INK4a contributes to the outcome of cancer therapy. Cell. 2002; 109: 335-346.

**15.** Sherr CJ and Roberts JM. Living with or without cyclins and cyclin-dependent kinases. Genes Dev. 2004; 18: 2699-2711.

**16.** Vafa O, Wade M, Kern S, Beeche M, Pandita TK, et al. c-Myc can induce DNA damage, increase reactive oxygen species, and mitigate p53 function: a mechanism for oncogene-induced genetic instability. Mol Cell. 2002; 9: 1031-1044.

**17.** Ray S, Atkuri KR, Deb-Basu D, Adler AS, Chang HY, et al. MYC can induce DNA breaks in vivo and in vitro independent of reactive oxygen species. Cancer Res. 2006; 66: 6598-6605.

**18.** Dominguez-Sola D, Ying CY, Grandori C, Ruggiero L, Chen B, et al. Non-transcriptional control of DNA replication by c-Myc. Nature. 2007; 448: 445-451.

**19.** Bartkova J, Rezaei N, Liontos M, Karakaidos P, Kletsas D, et al. Oncogene-induced senescence is part of the tumorigenesis barrier imposed by DNA damage checkpoints. Nature. 2006; 444: 633-637.

**20.** Di Micco R, Fumagalli M, Cicalese A, Piccinin S, Gasparini P, et al. Oncogene-induced senescence is a DNA damage response triggered by DNA hyper-replication. Nature. 2006; 444: 638-642.

**21.** Ramjaun AR and Downward J. Ras and phosphoinositide 3-kinase: partners in development and tumorigenesis. Cell Cycle. 2007; 6: 2902-2905.

**22.** Seoane J, Le HV, Massague J. Myc suppression of the p21(Cip1) Cdk inhibitor influences the outcome of the p53 response to DNA damage. Nature. 2002; 419: 729-734.

**23.** Herold S, Wanzel M, Beuger V, Frohme C, Beul D, et al. Negative regulation of the mammalian UV response by Myc through association with Miz-1. Mol Cell. 2002; 10: 509-521.

**24.** Hydbring P, Bahram F, Su Y, Tronnersjo S, Hogstrand K, et al. Phosphorylation by Cdk2 is required for Myc to repress Rasinduced senescence in cotransformation. Proc Natl Acad Sci U S A. 2010; 107: 58-63.

**25.** Wu S, Cetinkaya C, Munoz-Alonso MJ, von der Lehr N, Bahram F, et al. Myc represses differentiation-induced p21CIP1 expression via Miz-1-dependent interaction with the p21 core promoter. Oncogene. 2003; 22: 351-360.

**26.** Guney I, Wu S, Sedivy JM. Reduced c-Myc signaling triggers telomere-independent senescence by regulating Bmi-1 and p16(INK4a). Proc Natl Acad Sci U S A. 2006; 103: 3645-3650.

**27.** Kauffmann-Zeh A, Rodriguez-Viciana P, Ulrich E, Gilbert C, Coffer P, et al. Suppression of c-Myc-induced apoptosis by Ras signalling through PI(3)K and PKB. Nature. 1997; 385: 544-548.

**28.** Xu D, Popov N, Hou M, Wang Q, Bjorkholm M, et al. Switch from Myc/Max to Mad1/Max binding and decrease in histone acetylation at the telomerase reverse transcriptase promoter during differentiation of HL60 cells. Proc Natl Acad Sci U S A. 2001; 98: 3826-3831.

**29.** Feldser DM and Greider CW. Short telomeres limit tumor progression in vivo by inducing senescence. Cancer Cell. 2007; 11: 461-469.

**30.** Wu CH, van Riggelen J, Yetil A, Fan AC, Bachireddy P, et al. Cellular senescence is an important mechanism of tumor regression upon c-Myc inactivation. Proc Natl Acad Sci U S A. 2007; 104: 13028-13033.

**31.** Zhuang D, Mannava S, Grachtchouk V, Tang WH, Patil S, et al. C-MYC overexpression is required for continuous suppression of oncogene-induced senescence in melanoma cells. Oncogene. 2008; 27: 6623-6634.

**32.** Vervoorts J, Luscher-Firzlaff J, Luscher B. The ins and outs of MYC regulation by posttranslational mechanisms. J Biol Chem. 2006; 281: 34725-34729.

**33.** Hemann MT, Bric A, Teruya-Feldstein J, Herbst A, Nilsson JA, et al. Evasion of the p53 tumour surveillance network by tumour-derived MYC mutants. Nature. 2005; 436: 807-811.

**34.** Benassi B, Fanciulli M, Fiorentino F, Porrello A, Chiorino G, et al. c-Myc phosphorylation is required for cellular response to oxidative stress. Mol Cell. 2006; 21: 509-519.

**35.** Campaner S, Doni M, Hydbring P, Verrecchia A, Bianchi L, et al. Cdk2 suppresses cellular senescence induced by the c-myc oncogene. Nat Cell Biol. 2010; 12: 54-59; sup pp 51-14.

**36.** Grandori C, Wu KJ, Fernandez P, Ngouenet C, Grim J, et al. Werner syndrome protein limits MYC-induced cellular senescence. Genes Dev. 2003; 17: 1569-1574.

**37.** Deans AJ, Khanna KK, McNees CJ, Mercurio C, Heierhorst J, et al. Cyclin-dependent kinase 2 functions in normal DNA repair and is a therapeutic target in BRCA1-deficient cancers. Cancer Res. 2006; 66: 8219-8226.

**38.** Ortega S, Prieto I, Odajima J, Martin A, Dubus P, et al. Cyclindependent kinase 2 is essential for meiosis but not for mitotic cell division in mice. Nat Genet. 2003; 35: 25-31.

**39.** Goga A, Yang D, Tward AD, Morgan DO, Bishop JM. Inhibition of CDK1 as a potential therapy for tumors over-expressing MYC. Nat Med. 2007; 13: 820-827.

**40.** Cartwright P, McLean C, Sheppard A, Rivett D, Jones K, et al. LIF/STAT3 controls ES cell self-renewal and pluripotency by a Myc-dependent mechanism. Development. 2005; 132: 885-896.

**41.** Knoepfler PS, Cheng PF, Eisenman RN. N-myc is essential during neurogenesis for the rapid expansion of progenitor cell populations and the inhibition of neuronal differentiation. Genes Dev. 2002; 16: 2699-2712.

**42.** Laurenti E, Varnum-Finney B, Wilson A, Ferrero I, Blanco-Bose WE, et al. Hematopoietic stem cell function and survival depend on c-Myc and N-Myc activity. Cell Stem Cell. 2008; 3: 611-624.

**43.** Collado M, Blasco MA, Serrano M. Cellular senescence in cancer and aging. Cell. 2007; 130: 223-233.

**44.** Tyner SD, Venkatachalam S, Choi J, Jones S, Ghebranious N, et al. p53 mutant mice that display early ageing-associated phenotypes. Nature. 2002; 415: 45-53.

**45.** Maier B, Gluba W, Bernier B, Turner T, Mohammad K, et al. Modulation of mammalian life span by the short isoform of p53. Genes Dev. 2004; 18: 306-319.

**46.** Janzen V, Forkert R, Fleming HE, Saito Y, Waring MT, et al. Stem-cell ageing modified by the cyclin-dependent kinase inhibitor p16INK4a. Nature. 2006; 443: 421-426.

**47.** Molofsky AV, Slutsky SG, Joseph NM, He S, Pardal R, et al. Increasing p16INK4a expression decreases forebrain progenitors and neurogenesis during ageing. Nature. 2006; 443: 448-452.

**48.** Krishnamurthy J, Ramsey MR, Ligon KL, Torrice C, Koh A, et al. p16INK4a induces an age-dependent decline in islet regenerative potential. Nature. 2006; 443: 453-457.

**49.** Ressler S, Bartkova J, Niederegger H, Bartek J, Scharffetter-Kochanek K, et al. p16INK4A is a robust in vivo biomarker of cellular aging in human skin. Aging Cell. 2006; 5: 379-389.

**50.** Cheng T, Rodrigues N, Shen H, Yang Y, Dombkowski D, et al. Hematopoietic stem cell quiescence maintained by p21cip1/waf1. Science. 2000; 287: 1804-1808.

**51.** Matheu A, Maraver A, Klatt P, Flores I, Garcia-Cao I, et al. Delayed ageing through damage protection by the Arf/p53 pathway. Nature. 2007; 448: 375-379.

**52.** Tomas-Loba A, Flores I, Fernandez-Marcos PJ, Cayuela ML, Maraver A, et al. Telomerase reverse transcriptase delays aging in cancer-resistant mice. Cell. 2008 135: 609-622.