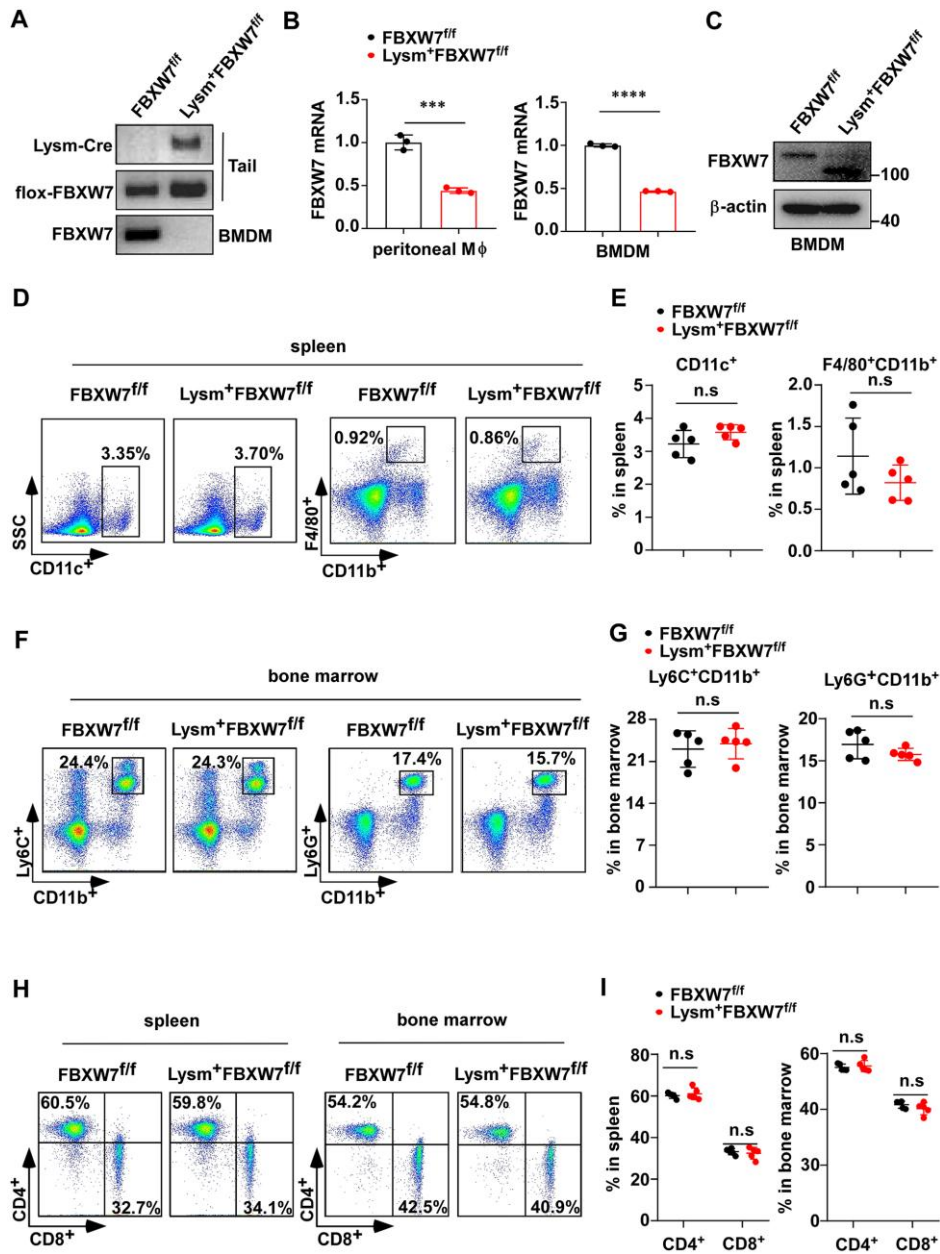
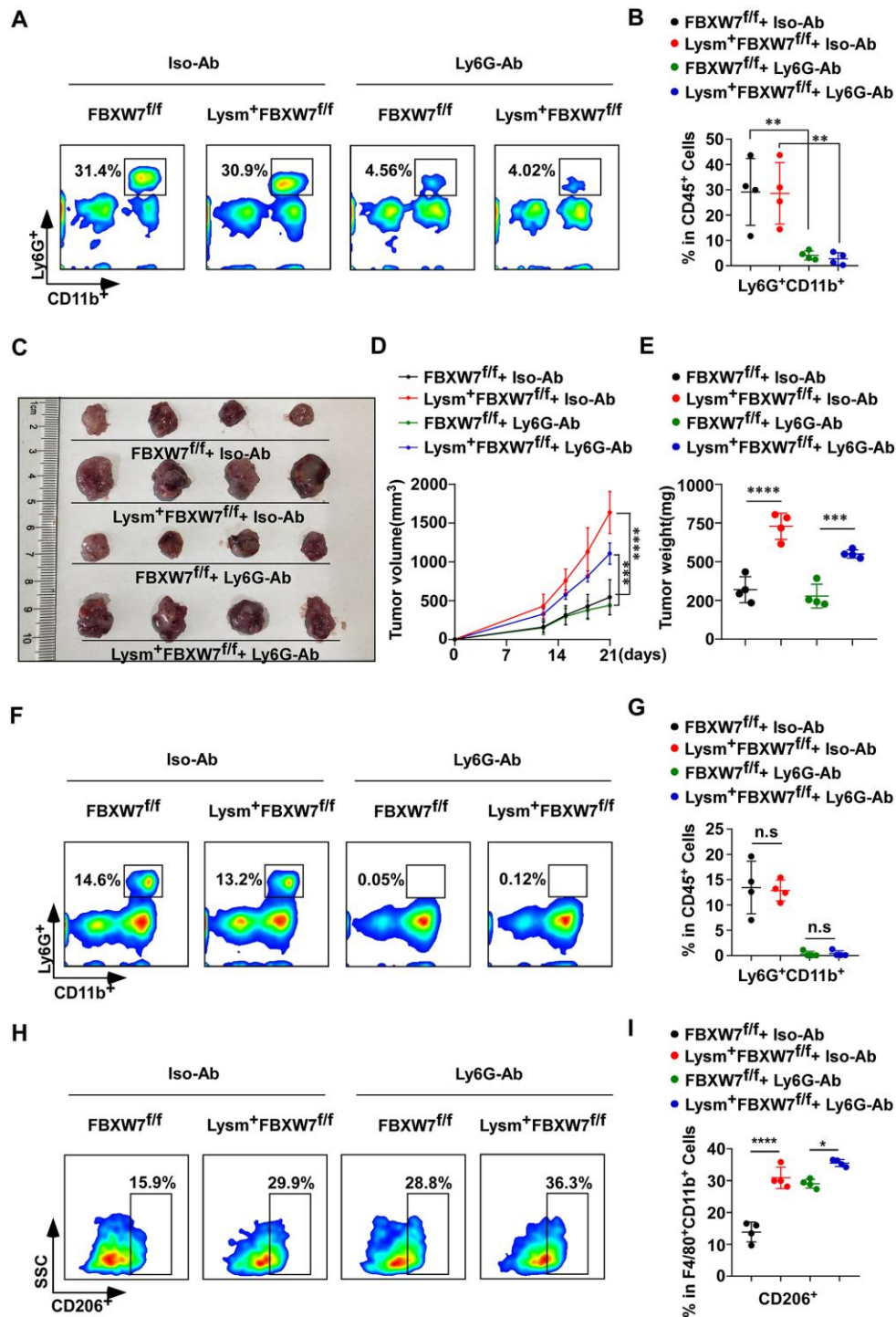


SUPPLEMENTARY FIGURES

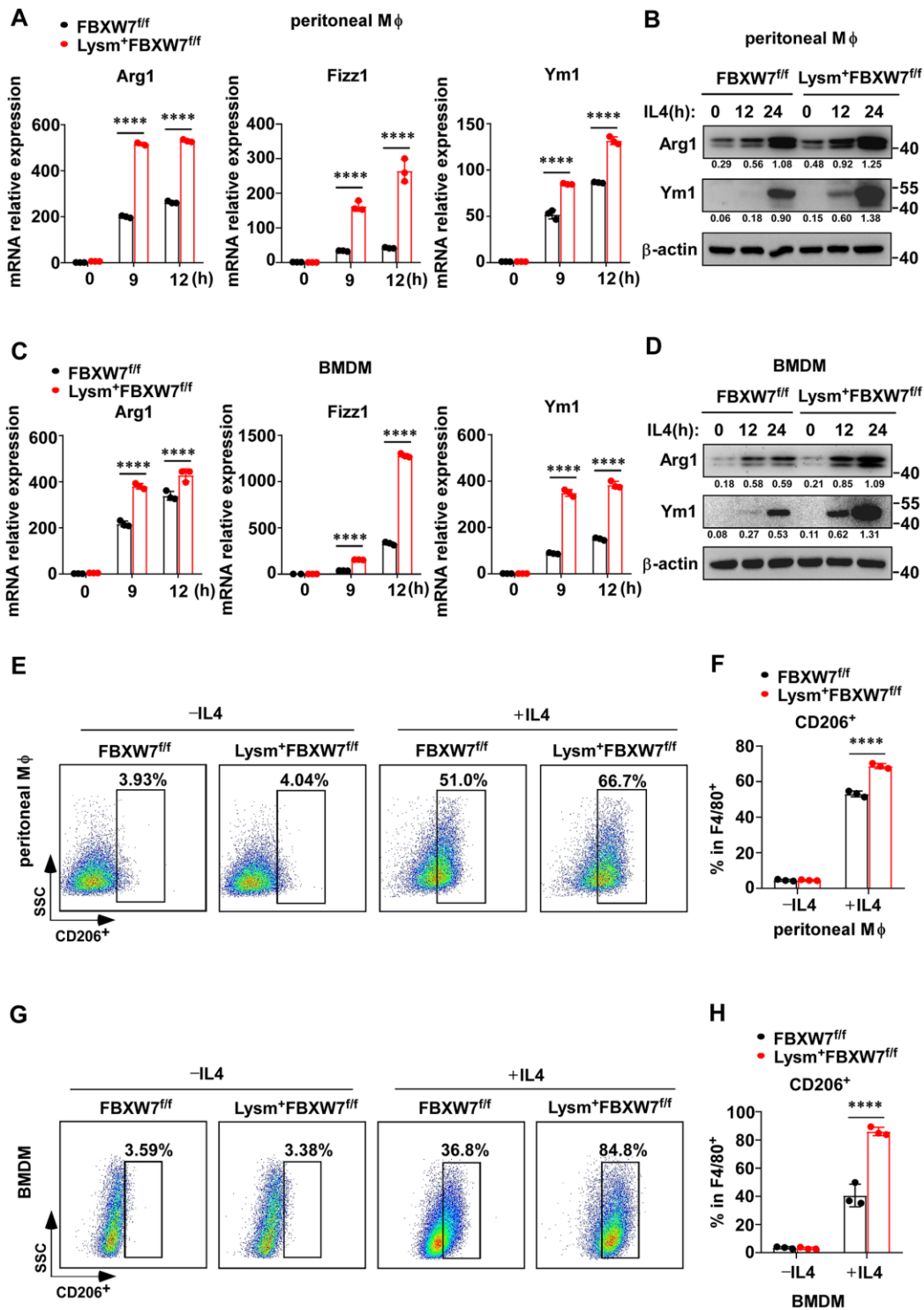


**Supplementary Figure 1. FBXW7 knockout in myeloid cells does not affect the development of myeloid cells and lymphocytes.** (A) PCR analysis “Lysm-Cre”, “flox” sequences of FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice, PCR analysis the excision of exons 5 and 6 of FBXW7 in bone marrow-derived macrophages (BMDMs) from FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice. (B) qRT-PCR analysis of *FBXW7* expression in peritoneal macrophages and BMDMs derived from FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice. (C) Western blot analysis of FBXW7 expression in BMDMs derived from FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice. (D, E) Flow cytometry analysis (D) and statistical analysis (E) of CD11c<sup>+</sup> dendritic cells and F4/80<sup>+</sup>CD11b<sup>+</sup> macrophages in the spleen of FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice (n=5 per group). (F, G) Flow cytometry analysis (F) and statistical analysis (G) of Ly6C<sup>+</sup>CD11b<sup>+</sup> monocytes and Ly6G<sup>+</sup>CD11b<sup>+</sup> granulocytes in the bone marrow of FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice (n=5 per group). (H, I) Flow cytometry analysis of CD4<sup>+</sup> and CD8<sup>+</sup> T cells in the spleen and bone marrow from FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice. (I) Statistical differences in the proportions of CD4<sup>+</sup> and CD8<sup>+</sup> T cells in the spleen and bone marrow of FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice by flow cytometry (n=5 per group). Data are shown as the mean ± SD and are representative of three independent experiments. n = 3 or 5. \*\*\*P < 0.001; \*\*\*\*P < 0.0001; n.s, no significance (Student’s t test (B, E, G, I)).

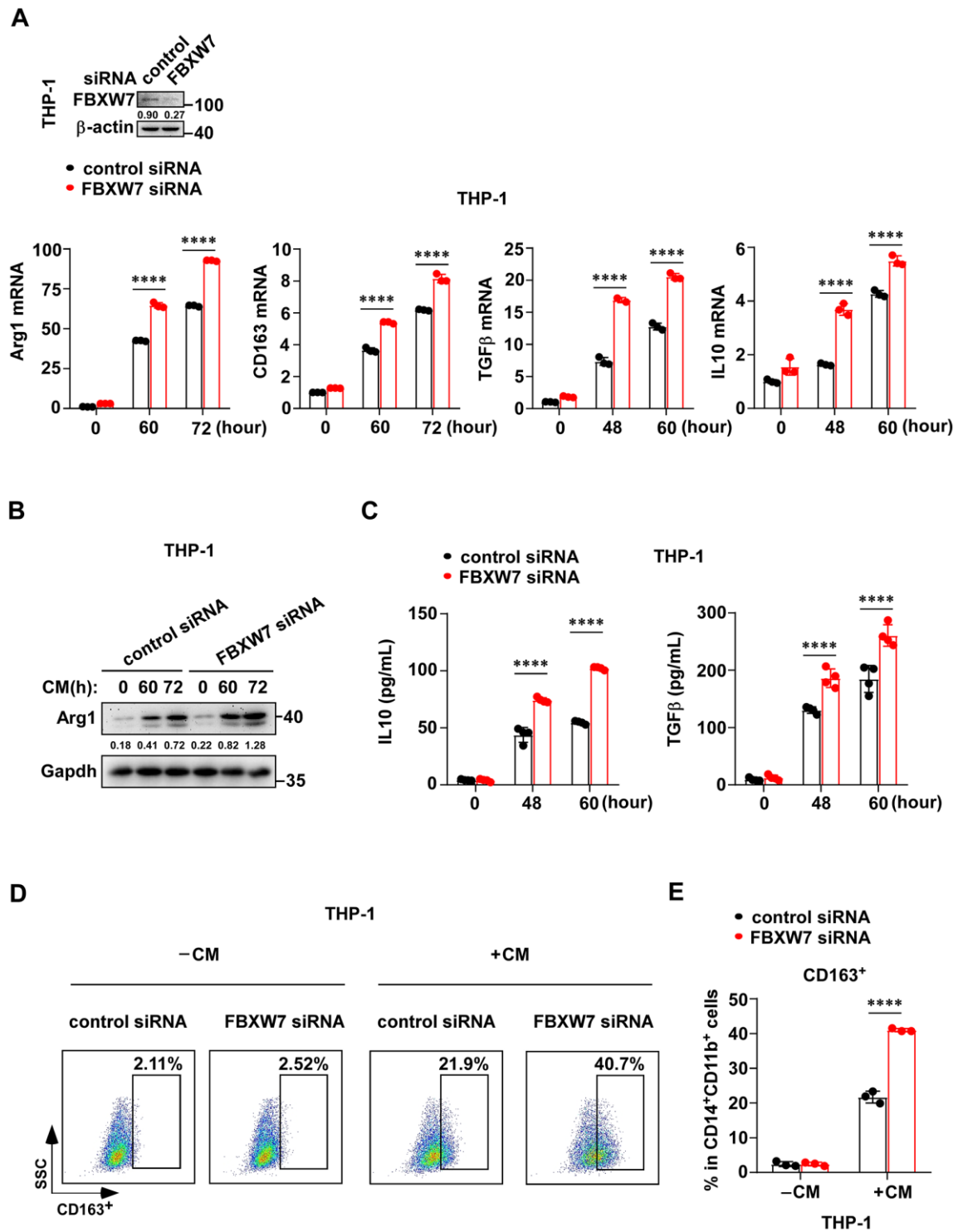


**Supplementary Figure 2. Neutrophils are not responsible for the aggravated tumor progression in Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice.**

(A, B) Neutrophils were depleted by anti-Ly6G mAb. Flow cytometry analysis (A) and statistical analysis (B) of the percentage of neutrophils (Ly6G<sup>+</sup>CD11b<sup>+</sup>) in the blood of FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice with or without anti-Ly6G mAb used showed to evaluate the efficiency of depletion. (n = 4 per group). (C–E) FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice inoculated with LLCs after anti-Ly6G mAb or isotype control antibody used. The appearance (C), volume (D), weight (E) of tumors in four groups in 21 days. (F, G) Flow cytometry analysis (F) and statistical analysis (G) of the percentage of neutrophils (Ly6G<sup>+</sup>CD11b<sup>+</sup>) in the tumors of FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice with or without anti-Ly6G mAb used (n = 4 per group). (H, I) Flow cytometry analysis (H) and statistical analysis (I) of the percentage of CD206<sup>+</sup> macrophages in tumors of FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice with or without anti-Ly6G mAb used (n = 4 per group). \*P < 0.05; \*\*P < 0.01; \*\*\*P < 0.001; \*\*\*\*P < 0.0001; n.s, no significance (one-way ANOVA (B, D, E, G, I)).



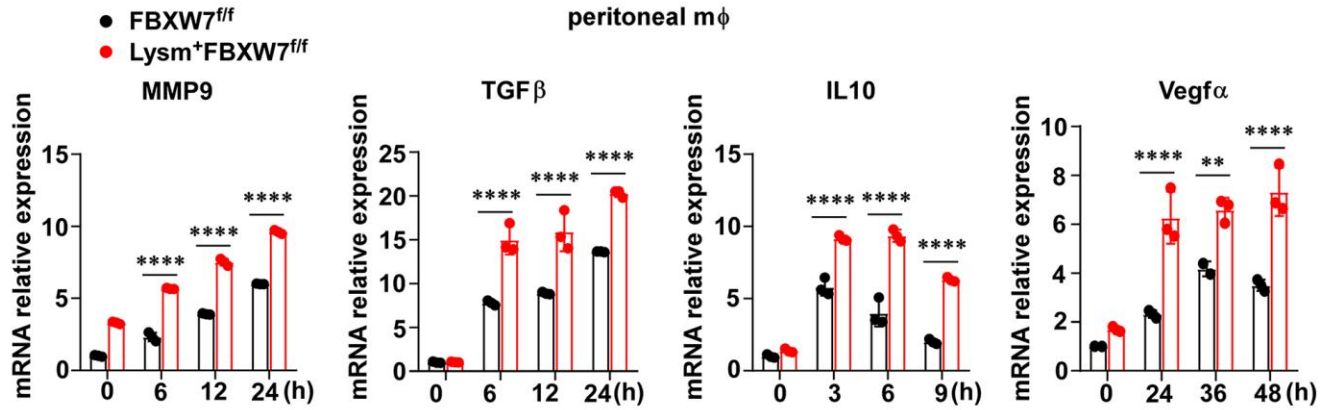
**Supplementary Figure 3. FBXW7 knockout enhances IL-4-induced M2 macrophage polarization.** (A) Peritoneal macrophages extracted from *FBXW7<sup>fl/fl</sup>* and *Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup>* mice were treated with IL-4 (20 ng/ml), and the mRNA expression of *Arg1*, *Fizz1*, and *Ym1* was analyzed by qRT-PCR. (B) The protein expression of Arg1 and Ym1 from wild-type and *FBXW7*-knockout peritoneal macrophages stimulated with IL-4 was detected by immunoblotting. (C, D) The mRNA (C) and protein (D) expression of M2-like TAM-associated genes were analyzed by qRT-PCR and immunoblotting, respectively, in BMDMs stimulated with IL-4. (E, F) Flow cytometry analysis (E) and statistical analysis (F) of the percentage of M2 macrophages (CD206<sup>+</sup>) in wild-type and *FBXW7*-knockdown peritoneal macrophages after IL-4 stimulation for 24 h (n = 3 per group). (G, H) Flow cytometry analysis (G) and statistical analysis (H) of the percentage of M2 macrophages (CD206<sup>+</sup>) in wild-type and *FBXW7*-knockout BMDMs after IL-4 stimulation for 24 h (n = 3 per group). Data are shown as the mean  $\pm$  SD and are representative of three independent experiments. \*\*\**P* < 0.001; \*\*\*\**P* < 0.0001; n.s, no significance (two-way ANOVA (A, C, F, H)).



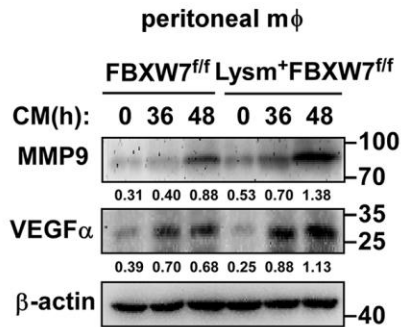
**Supplementary Figure 4. FBXW7 knockdown in THP-1 promotes A549 supernatant-induced M2 macrophage polarization.**

(A) THP-1 cells were differentiated into macrophages in the presence of PMA for 48 hours and transfected with FBXW7 siRNA. The silenced and unsilenced THP-1 stimulated with conditioned medium containing A549 cells cultured supernatant. The protein expression of FBXW7 in THP-1 was examined by immunoblotting and the mRNA expression of *Arg1*, *CD163*, *TGFβ*, and *IL10* was detected by qRT-PCR. (B) The protein expression of Arg1 in two groups was examined by immunoblotting. (C) The protein levels of IL10 and TGFβ in the supernatant of wild-type and FBXW7-knockdown THP-1 that co-cultured with A549 cells for the indicated time were measured by ELISA kits. (D, E) Flow cytometry analysis (D) and statistical analysis (E) of the percentage of M2 macrophages (CD163<sup>+</sup>) in wild-type and FBXW7-knockdown THP-1 after conditioned medium stimulation for 72 h (n = 3 per group). Data are shown as the mean ± SD and are representative of three independent experiments. \*\*\*\*P < 0.0001; (two-way ANOVA (A, C, E)).

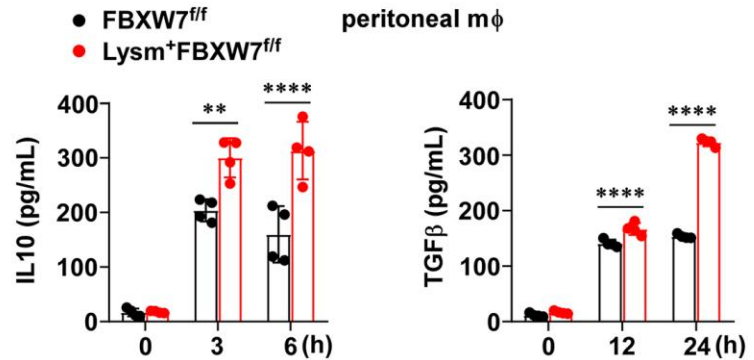
**A**



**B**

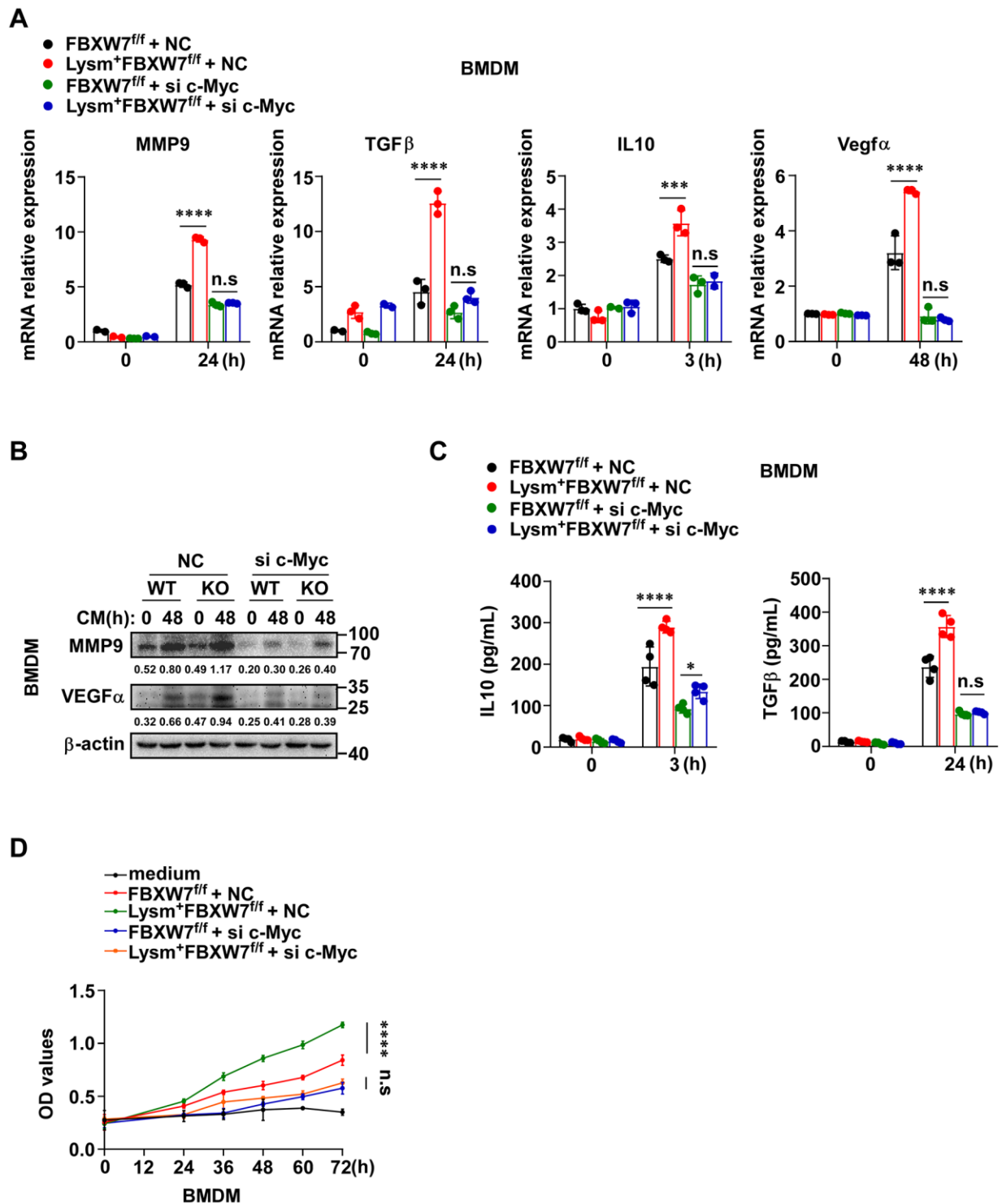


**C**



**Supplementary Figure 5. FBXW7 knockout promotes the expression of pro-tumoral factors in peritoneal macrophages.**

(A) Peritoneal macrophages from *FBXW7<sup>fl/fl</sup>* and *Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup>* mice were stimulated with the conditioned medium, and the mRNA expression of *MMP9*, *IL-10*, *TGFβ*, and *VEGFα* was examined by qRT-PCR. (B) The protein expression of *MMP9* and *VEGFα* in peritoneal macrophages incubated with the conditioned medium were detected by immunoblotting. (C) The protein levels of *IL10* and *TGFβ* in the supernatant of peritoneal macrophages that co-cultured with LLCs for the indicated time was measured by ELISA kits. Data are shown as the mean ± SD and are representative of three independent experiments. \*\**P* < 0.01; \*\*\*\**P* < 0.0001 (two-way ANOVA (A, C)).



**Supplementary Figure 6. The effect of FBXW7 on M2 macrophage expression of pro-tumoral factors is dependent on c-Myc.** (A) qRT-PCR analysis of *MMP9*, *IL-10*, *TGFβ*, and *VEGFα* mRNA expression in BMDMs from FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice transfected with or without c-Myc siRNA and stimulated with conditioned medium. (B) Immunoblotting analysis of MMP9 and VEGFα expression in primary macrophages from FBXW7<sup>fl/fl</sup> and Lysm<sup>+</sup>FBXW7<sup>fl/fl</sup> mice transfected with or without c-Myc siRNA and stimulated with conditioned medium. (C) The protein levels of IL10 and TGFβ in the supernatant of four groups were measured by ELISA kits. (D) LLCs were cultured in serum-free RPMI-1640, supernatant from IL-4-induced wild-type or FBXW7-knockout macrophages, supernatant from IL-4-induced wild-type or FBXW7-knockout macrophages transfected with c-Myc siRNA. The proliferation of LLCs in five groups was measured by the MTT assay. Data are shown as the mean ± SD and are representative of three independent experiments. \**P* < 0.05; \*\*\**P* < 0.001; \*\*\*\**P* < 0.0001; n.s, no significance (two-way ANOVA (A, C, D)).