Associations between *TUBB-WWOX* SNPs, their haplotypes, genegene, and gene-environment interactions and dyslipidemia

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ABSTRACT

In this study, we investigated associations between single nucleotide polymorphisms (SNPs) in the tubulin beta class I (TUBB) and WW domain-containing oxidoreductase (WWOX) genes, gene-gene interactions, and geneenvironment interactions and dyslipidemia in the Chinese Maonan ethnic group. Four SNPs (rs3132584, rs3130685, rs2222896, and rs2548861) were genotyped in unrelated subjects with normal lipid levels (864) or dyslipidemia (1129). While 5.0% of Maonan subjects carried the rs3132584TT genotype, none of the Chinese Han in Beijing subjects did. Allele and genotype frequencies differed between the normal and dyslipidemia groups for three SNPs (rs3132584, rs3130685, and rs2222896). rs2222896G allele carriers in the normal group had higher low-density lipoprotein cholesterol and lower high-density lipoprotein cholesterol levels. The rs3132584GG, rs3130685CC+TT, and rs2222896GG genotypes as well as the rs2222896G-rs2548861G and rs2222896G-rs2548861T haplotypes were associated with an elevated risk of dyslipidemia; the rs2222896Ars2548861T and rs2222896A-rs2548861G haplotypes were associated with a reduced risk of dyslipidemia. Among the thirteen TUBB-WWOX interaction types identified, rs3132584T-rs3130685T-rs2222896G-rs2548861T increased the risk of dyslipidemia 1.371-fold. Fourteen two- to four-locus optimal interactive models for SNP-SNP, haplotype-haplotype, gene-gene, and gene-environment interactions exhibited synergistic or contrasting effects on dyslipidemia. Finally, the interaction between rs3132584 and rs2222896 increased the risk of dyslipidemia 2.548-fold and predicted hypertension.

INTRODUCTION

The average global prevalence of dyslipidemia is about 20% [1–3], and it is higher in patients with premature coronary arteriosclerotic diseases (CAD) [4] and chronic kidney disease [5]. Dyslipidemia plays crucial roles in the pathogenesis of hypertension, CAD, stroke, and chronic

kidney failure that differ based on etiology, and identification of novel and safe treatments that decrease blood lipid levels, especially the level of low-density lipoprotein cholesterol (LDL-C), would be beneficial [6]. It is well known that lifestyle, environmental, and genetic factors all contribute to dyslipidemia. In the past decade, technological advances in biological sequencing technology have enabled genome-wide association studies (GWASs) that have helped thoroughly characterize risk factors for dyslipidemia.

The tubulin beta class I gene (TUBB, gene ID: 203068) located on chromosome 6 in humans encodes a beta tubulin protein that is an essential component of microtubules. Microtubules perform many cellular functions. including chromosome segregation, maintenance of cell shape, transport and motility, and organelle distribution [7]. As a substrate of peptidylarginine deiminases, TUBB participates in citrullination, which is related to growth, infiltration, and drug resistance in some tumor cells. Thus, mutations in TUBB are involved in complex diseases [8] and carcinoma [9]. The WW domain-containing oxidoreductase gene (WWOX, gene ID: 51741) on human chromosome 16 encodes a member of the short-chain dehydrogenases/reductases protein family that is involved in a variety of important cellular processes, including induction of apoptosis, cell development, and steroid metabolism [10, 11]. Mutations in WWOX are not only associated with multiple types of cancer [11], but also with lipid metabolism [12]. Both TUBB and WWOX are ubiquitously expressed in many tissues, including fat (reads per kilobase per million mapped reads (RPKM) 97.7 and 1.6, respectively), heart (RPKM 67.2 and 0.5) and liver (RPKM 58.1 and 0.9). Because of their functions and tissue distribution, single nucleotide polymorphisms in the TUBB and WWOX genes can have similar effects on lipid profiles. Recently, GWASs revealed that SNPs at locations rs3132584 and rs3130685 in TUBB are associated with LDL-C profile [13], while SNPs at rs2222896 and rs2548861 in WWOX are associated with LDL-C and high-density lipoprotein cholesterol (HDL-C) profiles [14], respectively. However, associations between TUBB and WWOX and lipid traits in the Chinese population have not been examined; we hypothesize that modifications of TUBB and WWOX affect blood lipid levels in this population as well.

The incidence of dyslipidemia in Chinese adults is as high as 34% [15]. Previous studies revealed that several lipid-related genes are associated with different lipid traits in the Maonan and Han populations [16–18]. The Maonan ethnicity is one of 55 recognized minorities in China, and seventy percent (64,500) of Maonan people live in Huanjiang Maonan Autonomous County of Guangxi Zhuang Autonomous Region according to a wire report from Xinhua News Agency (Beijing, May 20th, 2020). The Maonan people maintain a unique lifestyle and wedding culture characterized by a fat-rich diet, recreational use of alcohol and tobacco, and intraethnic marriages. As a result, the Maonan population has a singular genetic background and specific environmental risk factors that are particularly informative in genetic studies of dyslipidemia. In this study, we therefore explored associations between *TUBB-WWOX* SNPs, their haplotypes, and gene-gene ($G \times G$) and gene-environment ($G \times E$) interactions and the prevalence of dyslipidemia in the Maonan population.

RESULTS

Participant characteristics

As shown in Table 1, no statistically significant differences in mean age, gender ratio, height, cigarette smoking, and proportion over 65 years old were observed between the normal and dyslipidemia groups (P > 0.05). However, mean weight, waist circumference, body mass index (BMI), alcohol consumption, systolic blood pressure (SBP), diastolic blood pressure (DBP), pulse pressure (PP), fasting blood-glucose (FBS), hypertension morbidity, and proportion with BMI greater than 24 kg/m² and with FBS of at least 7.0 mM were higher in the dyslipidemia group than in the normal group (P < 0.01). In addition, total cholesterol (TC), triglyceride (TG), LDL-C, and apolipoprotein (Apo) B levels were also significantly higher in the dyslipidemia group than in the normal group (P < 0.001). In contrast, HDL-C and ApoA1 levels and ApoA1/ApoB ratio were significantly lower in the dyslipidemia group than in the normal group (*P* < 0.001).

Genotype and allele frequencies

Information for four target SNPs (rs3132584, rs3130685, rs2222896, and rs2548861) in the TUBB and WWOX genes among the Chinese Han Beijing (CHB) population in NCBI dbSNP Build 132 is shown in Table 2; genotypic and allelic frequencies of these four SNPs in the Maonan population are presented in Table 3. The frequencies of point mutations at all four SNPs were similar between the CHB and Maonan populations. Mutations from G to T and from C to T were observed at the TUBB rs3132584 and rs3130685 SNPs, respectively, while mutations from A to G and from T to G were observed at the WWOX rs2222896 and rs2548861 SNPs, respectively. However, genotypic and allelic frequencies of the four SNPs differed between the ethnic groups; in particular, the frequency of the rs3132584TT genotype was 5.0% in Maonan population but was 0% in the CHB population. In this study, allelic and genotypic distributions of the four SNPs in both groups were consistent with the Hardy Weinberg Equilibrium (HWE, P > 0.05 for all). Additionally, allele and genotype frequencies of the rs3132584, rs3130685, and rs2222896 SNPs differed between the normal and dyslipidemia groups ($P \le 0.01$ for all).

Parameter	Normal	Dyslipidemia	$t(\chi^2)$	Р
Number	864	1129	_	_
Male/female	431/433	545/584	0.509	0.476
Age, years ¹	55.31±13.08	56.04±12.88	-1.240	0.215
Height, cm	156.34 ± 8.01	156.75 ± 8.32	-1.123	0.262
Weight, kg	54.48±110.29	57.06±11.07	-5.365	0.000
Waistline, cm	76.06±9.31	80.33±9.47	-8.199	0.000
Body mass index (BMI), kg/m ²	22.21±3.31	23.11±3.39	-5.971	0.000
Cigarette smoking, n (%) ²				
0 cigarette/day	622(71.99)	818(72.45)		
≤ 20 cigarettes/day	192(22.22)	239(21.17)		
> 20 cigarettes/day	50(5.79)	72(6.38)	0.544	0.762
Alcohol consumption, n (%)				
0 g/day	607(70.25)	841(74.49)		
$\leq 25 \text{ g/day}$	50(5.79)	84(7.44)		
> 25 g/day	207(23.96)	204(18.07)	11.633	0.003
Systolic blood pressure, mmHg	127.80±20.45	135.34±22.30	-7.844	0.000
Diastolic blood pressure, mmHg	79.83±12.29	84.53±12.91	-8.276	0.000
Pulse pressure, mmHg	48.91±15.00	51.31±16.39	-3.404	0.001
Fasting blood-glucose (FBS), mM	6.03±0.94	6.32±1.23	-5.915	0.000
Total cholesterol, mM	4.33±0.50	4.89±0.72	-20.024	0.000
Triglyceride, mM	1.00(1.07)	1.56(2.02)	-28.603	0.000
HDL-C, mM	1.30±0.18	1.15 ± 0.17	19.195	0.000
LDL-C, mM	2.90±0.15	3.43±0.31	-49.871	0.000
Apolipoprotein (Apo)A1, g/L	1.32±0.18	1.28±0.16	5.931	0.000
ApoB, g/L	0.94±0.14	1.00 ± 0.10	-10.755	0.000
ApoA1/ApoB	1.45±0.32	1.29±0.24	11.697	0.000
Hypertension, n (%)	288(33.33)	571(43.96)	59.334	0.000
Age > 65 years	194(22.45)	255(22.59)	0.0050	0.944
$BMI > 24 \text{ kg/m}^2$	255(22.59)	475(42.07)	55.200	0.000
FBS ≥ 7.0 mM, n (%)	164(18.98)	495(43.84)	52.194	0.000

HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol. ¹Normally distributed quantitative data were assessed in a *t*-test and are described as means \pm SD. ²Qualitative data were assessed using the chi-square test. ³Non-normally distributed quantitative data were assessed using the Wilcoxon-Mann-Whitney test and are described as medians (interquartile range). *P* < 0.05 indicated a statistically significant difference.

Gene	Chromosome	SNP	Allele ₁	Allele ₂	P _{HWE}	Genotype detail of CHB	Source	Phenotype
TUBB	6	rs3132584	G(0.8171)	*T(0.1829)	0.4386	GG(0.6341) GT(0.3591) —	1000 Genome	↑LDL(G)
	6	rs3130685	*C(0.4556)	T(0.5444)	1.0000	CC(0.2000) CT(0.5111) TT(0.2889) 1000 Genome	\downarrow LDL(T)
WWOX	16	rs2222896	*A(0.3659)	G(0.6341)	0.7518	AA(0.1220) AG(0.3902) GG(0.487	3) 1000 Genome	↑LDL(G)
	16	rs2548861	*T(0.4512)	G(0.5488)	0.4028	TT(0.1707) TG(0.5610) GG(0.268	3) 1000 Genome	\downarrow HDL(T)

SNP, single nucleotide polymorphism; HWE, Hardy-Weinberg equilibrium; CHB, Chinese Han Beijing. Allele1 and the allele2 indicate reference and alternate alleles, respectively; *allele indicates the minor allele.

Gene SNP	Genotype (allele)	Total (<i>n</i> =1993)	Normal (<i>n</i> = 864)	Dyslipidemia (n = 1229)	χ^2	Р
TUBB						
rs3132854	GG	1223(61)	467(54)	756(67)		
	GT	676(34)	337(39)	339(30)		
	TT	94(5)	60(7)	34(3)	40.978	0.000
	G	3122(78)	1271(74)	1851(82)		
	Т	864(22)	457(26)	407(18)	40.896	0.000
	$P_{ m HWE}$	0.95	1.000	0.690		
rs3130685	CC	361(18)	138(16)	223(20)		
	CT	980(49)	415(48)	565(50)		
	TT	652(33)	311(36)	341(30)	9.282	0.010
	Т	2284(57)	1037(60)	1247(55)		
	С	1702(43)	691(40)	1011(45)	9.163	0.002
	P_{HWE}	0.850	1.000	0.720		
WWOX						
rs2222896	AA	395(20)	259(30)	136(12)		
	AG	961(48)	444(51)	517(46)		
	GG	637(32)	161(19)	476(42)	167.338	0.000
	G	2235(56)	766(44)	1469(65)		
	А	1751(44)	962(56)	789(35)	170.763	0.000
	P_{HWE}	0.340	0.240	0.840		
rs2548861	TT	287(14)	118(14)	169(15)		
	TG	947(48)	404(47)	543(48)		
	GG	759(38)	342(40)	417(37)	1.670	0.434
	G	2465(62)	1088(63)	1377(61)		
	Т	1521(38)	640(37)	881(39)	1.626	0.202
	P_{HWE}	0.780	1.000	0.750		

Table 3. Genotypic and allelic frequencies of TUBB and WWOX SNPs in normal and dyslipidemia groups [n(%)].

Qualitative data were assessed using the Chi-square test; P < 0.05 indicated a statistically significant difference. HWE, Hardy-Weinberg equilibrium. $P_{HWE} > 0.05$ indicated a statistically significant difference.

Associations between genotypes and alleles, serum lipid levels, and dyslipidemia

Associations between the four TUBB and WWOX SNPs and serum lipid levels are shown in Figure 1. Serum levels of HDL-C, LDL-C, and ApoA1 differed among the three rs2222896 genotypes in the normal group ($P \leq$ 0.001 for all); rs2222896G allele carriers had higher LDL-C and lower HDL-C levels than those without rs2222896G. However, there were no differences in serum lipid levels depending on genotype for the other three SNPs. In the dyslipidemia group, some lipid parameters differed significantly depending on genotype for the rs3132584, rs2222896, and rs2548861 SNPs (P < 0.0125 for all). Furthermore, higher TC levels and lower ApoA1/ApoB ratio were associated with rs2222896G and rs2548861T alleles, higher LDL-C and ApoB levels and lower HDL-C levels were associated with the rs3132584G, rs2222896G, and rs2548861T alleles,

lower ApoA1 levels were associated with the rs2222896G allele, and lower ApoA1/ApoB ratio was associated with the rs2548861T and rs2222896G alleles.

Because environmental factors can affect lipid phenotypes, we adjusted for the risk factors of age, sex, drinking, smoking, FBS, and BMI, to clarify relationships between genotypes and lipid phenotypes (Table 4). Overall, in the normal group, serum HDL-C levels were lower in rs2222896G allele carriers than in rs2222896G non-carriers ($P \le 0.001$), LDL-C was higher, but ApoA1 was lower, in subjects with the rs2222896GG genotype compared to those with the rs2222896AA genotype (P < 0.001), ApoB levels were higher in subjects with the rs3132584GG genotype than in those with the rs3132584TT genotype (P = 0.006), and ApoA1/ApoB ratio was lower in subjects with the rs2222896GG and rs3132584GT genotypes than in those with the rs2222896AA and rs3132584TT genotypes, respectively (P = 0.012 and P = 0.007). In the dyslipidemia group, serum TC and ApoB levels were higher, but ApoA1/ApoB ratio was lower, in subjects with the rs2548861GG genotype than in those with the rs2548861TG genotype ($P \le 0.005$ for all), serum HDL-C and ApoA1 levels and ApoA1/ApoB ratio were lower, but LDL-C levels were higher, in subjects with the rs2222896GG genotype than in those with the rs2222896AA genotype, and serum HDL-C levels were lower in subjects with the rs3132584GT, rs3132584GG, and rs2548861TT genotypes than in those with the rs3132584TT and rs2548861GG genotypes (P = 0.001 for all).

Associations between the four SNPs and dyslipidemia are shown in Table 5. rs3132584, rs3130685, and rs2222896 were associated with dyslipidemia (P < 0.05).

Subjects with the rs3132584TT [codominant: adjusted odds ratio (OR) = 0.35, 95% confidence interval (CI) = 0.23–0.54, *P* < 0.0001] or rs3132584GT+TT (dominant: adjusted OR = 0.58, 95% CI = 0.48-0.70, P < 0.0001) genotypes had a lower risk of dyslipidemia than those with the rs3132584GG genotype. In contrast, subjects with the rs3130685CC (codominant: adjusted OR =1.47, 95% CI = 1.13–1.92, P = 0.0096) or rs3130685 CT+TT (dominant: adjusted OR = 1.30, 95% CI = 1.08-1.57, P = 0.0064) genotypes had a higher risk of dyslipidemia than those with the rs3130685TT genotype. Finally, subjects with the rs2222896AA (codominant: adjusted OR = 0.18, 95% CI = 0.14-0.23, P < 0.0001) or rs2222896 AG+AA (dominant: adjusted OR = 0.31, 95% CI = 0.26–0.39, P < 0.0001) genotypes had a lower risk of dyslipidemia than those with the rs2222896GG genotype.



Figure 1. Associations between *TUBB-WWOX* genotypes and serum lipid levels in normal and dyslipidemia groups. A = Total cholesterol (mM); B = High-density lipoprotein cholesterol (mM); C = Low-density lipoprotein cholesterol (mM); D = Apolipoprotein (Apo) A1 (g/L); E = ApoB (g/L); F = ApoA1/ApoB ratio; and G = Triglyceride (mM). **P*-value < 0.006 indicated a statistically significant difference after Bonferroni correction.

Tinid	CND	Genotype Reference Alternate		Data	4	Р	95% CI of Beta	
Lipid	SNP			Beta	t	P	Lower	Upper
Normal								
HDL-C	rs2222896	AA	AG	-0.131	-3.313	0.001	-0.076	-0.019
	rs2222896	AA	GG	-0.165	-4.232	0.000	-0.112	-0.041
LDL-C	rs2222896	AA	GG	0.137	3.500	0.000	0.024	0.084
ApoA1	rs2222896	AA	GG	-0.160	-4.080	0.000	-0.109	-0.038
Apo B	rs3132584	TT	GG	0.188	2.739	0.006	0.015	0.090
ApoA1/ApoB	rs2222896	AA	GG	-0.099	-2.509	0.012	-0.143	-0.017
	rs3132584	TT	GT	-0.188	-2.708	0.007	-0.210	-0.034
Dyslipidemia								
TC	2548861	GG	TG	-0.156	-4.832	0.000	-0.318	-0.134
HDL-C	rs3132584	TT	GT	-0.272	-3.302	0.001	-0.159	-0.041
		TT	GG	-0.296	-3.598	0.000	-0.164	-0.048
	rs2222896	AA	GG	-0.155	-3.270	0.001	-0.085	-0.021
	2548861	GG	TT	-0.113	-3.471	0.001	-0.084	-0.023
LDL-C	rs2222896	AA	GG	0.136	2.843	0.005	0.027	0.146
ApoA1	rs2222896	AA	GG	-0.167	-3.535	0.000	-0.083	-0.024
Apo B	2548861	GG	TG	-0.089	-2.695	0.007	-0.031	-0.005
ApoA1/ApoB	rs2222896	AA	GG	-0.120	-2.549	0.011	-0.102	-0.013
-	2548861	GG	TG	0.085	2.577	0.010	0.010	0.070

TC, total cholesterol; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; ApoA1, apolipoprotein A1; ApoB, apolipoprotein B; CI, confidence interval. *P* values were calculated by multivariable linear regression. P < 0.0125 indicated a statistically significant difference after Bonferroni correction and adjusting for genotype, age, gender, drinking, smoking, fasting blood-glucose, and body mass index.

Como	Madal	Geno	otype	.2	D	OD (050/ CI)	*D	
Gene	Model	Reference	Alternate	χ^2	Р	OR (95% CI)	*Р	
TUBB								
rs3132584 G > T	Codominant	GG	GT	24.275	0.000	0.62(0.51-0.75)	< 0.0001	
			TT	23.917	0.000	0.35(0.23-0.54)		
	Dominant	GG	GT+TT	34.413	0.000	0.58(0.48-0.70)	< 0.0001	
	Recessive	GG+GT	TT	16.846	0.000	0.42(0.27-0.64)	< 0.0001	
	Log-additive	GT	TT	6.456	0.011	0.61(0.52-0.71)	< 0.0001	
rs3130685C>T	Codominant	TT	CT	4.542	0.033	1.24(1.02 - 1.52)	0.0096	
			CC	8.448	0.004	1.47(1.13-1.92)		
	Dominant	TT	CT+CC	7.458	0.006	1.30(1.08–1.57)	0.0064	
	Recessive	TT+CT	CC	4.714	0.030	1.29(1.03-1.64)	0.029	
	Log-additive	СТ	CC	1.848	0.174	1.22(1.07-1.38)	0.0024	
WWOX	-							
rs2222896A>G	Codominant	GG	AG	71.311	0.000	0.39(0.32-0.49)	< 0.0001	
			AA	164.027	0.000	0.18(0.14-0.23)		
	Dominant	GG	AG+AA	124.580	0.000	0.31(0.26-0.39)	< 0.0001	
	Recessive	GG+AG	AA	99.024	0.000	0.32(0.25-0.40)	< 0.0001	
	Log-additive	AG	AA	42.060	0.000	0.42(0.37-0.48)	< 0.0001	
rs2548861T>G	Codominant	GG	TG	0.985	0.321	1.10(0.91-1.34)	0.4300	
			TT	1.315	0.251	1.17(0.89–1.55)		
	Dominant	GG	TG+TT	1.455	0.228	1.12(0.93-1.34)	0.2300	
	Recessive	GG+TG	TT	0.683	0.409	1.11(0.86–1.43)	0.4100	
	Log-additive	TG	TT	0.216	0.642	1.09(0.96-1.24)	0.2000	

Table 5. Associations between genetic models of the four SNPs and dyslipidemia.

OR, odds ratio; CI, confidence interval. P and *P < 0.0125 indicated a statistically significant difference.

Associations between haplotype frequencies, serum lipid levels, and dyslipidemia

Linkage disequilibrium (LD) was observed between the *WWOX* rs2222896 and rs2548861 SNPs in the normal and dyslipidemia groups (D' = 0.870 or $R^2 = 0.350$ and

D' = 0.810 or $R^2 = 0.220$, respectively; Figure 2). Frequencies of the common haplotypes [lowest frequency threshold (LFT) > 0.03] of the two *WWOX* SNPs are shown in Table 6. The frequencies of the rs2222896A-rs2548861T, rs2222896G-rs2548861G, rs2222896A-rs2548861G, and rs2222896G-rs2548861T



Figure 2. Linkage disequilibrium (LD) analysis for the four *TUBB* and *WWOX* SNPs in normal and dyslipidemia groups. LD status is indicated by D' and R^2 values.

Table 6. Haplotype frequencies of the two WWOX SNPs in the normal and dyslipidemia groups [n(frequency)].

Label	Haplotype	Normal (<i>n</i> =1728)	J 1		Р	OR (95% CI)	*Р
H1	rs2222896A-rs2548861T	604(0.349)	451(0.199)	112.869	2.30×10^{-26}	0.464(0.420-0.535)	< 0.0001
H2	rs2222896G–rs2548861G	730(0.422)	1039(0.460)	5.632	0.017	1.165(1.026–1.322)	0.003
H3	rs2222896A-rs2548861G	358(0.207)	338(0.149)	22.445	2.16×10^{-6}	0.673(0.571-0.793)	0.007
H4	rs2222896G-rs2548861T	36(0.020)	430(0.190)	272.729	2.88×10^{-61}	11.055(7.818-15.633)	< 0.0001

H, haplotype; OR, odds ratio; CI, confidence interval. P and *P < 0.05 indicated a statistically significant difference. n = total number with that haplotype.

haplotypes differed between the two groups (P < 0.05 for all). The rs2222896G-rs2548861G and rs2222896G-rs2548861T haplotypes were associated with increased risk of dyslipidemia, whereas the rs2222896A-rs2548861T and rs2222896A-rs2548861G haplotypes

were protective factors against dyslipidemia (P < 0.007 for all).

Associations between the four haplotypes and serum lipid levels are shown in Figure 3. rs2222896G-rs2548861G



Figure 3. Associations between WWOX haplotypes, serum lipid levels, and dyslipidemia. *P-value < 0.006 indicated a statistically significant difference after Bonferroni correction.

haplotype carriers in the normal group had lower HDL-C levels than those without rs2222896G-rs2548861G haplotype (P = 0.001), rs2222896A-rs2548861G haplotype carriers had higher ApoA1 and HDL-C levels and ApoA1/ApoB ratio, but lower TG levels, than those without rs2222896A-rs2548861G haplotype ($P \le 0.002$), and rs2222896G-rs2548861T haplotype carriers had higher TC levels than those without rs2222896Grs2548861T haplotypes (P < 0.001). In the dyslipidemia group, rs2222896A-rs2548861T haplotype carriers had lower TC and ApoB levels, but higher HDL-C and ApoA levels and ApoA1/ApoB ratio than those without corresponding haplotype (P < 0.001), rs2222896Ars2548861G haplotype carriers had lower LDL-C and higher ApoA1 and TG levels than those without corresponding haplotype ($P \le 0.004$), and rs2222896Grs2548861T haplotype carriers had lower ApoA1 and HDL-C levels and ApoA1/ApoB ratio than those without corresponding haplotype (P < 0.001).

As shown in Table 7, after adjusting for sex, age, BMI, FBS, smoking, and drinking in the normal group, the rs2222896G-rs2548861T haplotype was associated with increased levels of serum TC and LDL-C (P < 0.001), the rs2222896A-rs2548861G haplotype was associated with increased serum HDL-C and ApoA1 levels and ApoA1/ApoB ratio, but decreased serum TG levels ($P \leq$ 0.006), and the rs2222896G-rs2548861G haplotype was associated with decreased serum HDL-C levels (P <0.001). In the dyslipidemia group, the rs2222896Ars2548861T haplotype was associated with decreased serum TC and ApoB levels and increased serum HDL-C and ApoA1 levels and ApoA1/ApoB ratio (P < 0.001), the rs2222896A-rs2548861G haplotype was associated with decreased serum LDL-C levels and increased serum ApoA1 ($P \le 0.006$) and TG levels (P < 0.001), and the rs2222896G-rs2548861T haplotype was associated with decreased serum ApoA1 and HDL-C levels and ApoA1/ApoB ratio (P < 0.001).

Interactions between the four haplotypes and environmental factors on dyslipidemia are shown in Figure 4. Dyslipidemia morbidity was increased in those with the rs2222896G-rs2548861G and rs2222896G-rs2548861T haplotypes compared to those without corresponding haplotypes (OR = 1.44, 95% CI = 1.18-1.76, P < 0.001 and OR = 15.08, 95% CI = 9.24–24.60, P < 0.001, respectively); environmental factors such as BMI > 24 kg/m² and FBS \ge 7.0 mM also increased dyslipidemia morbidity. In contrast, dyslipidemia morbidity was decreased in those with the rs2222896A-rs2548861T and rs2222896A-rs2548861G haplotypes compared to those without corresponding haplotypes (OR = 0.47, 95% CI = 0.39-0.56, P < 0.001and OR = 0.70, 95% CI = 0.56–0.86, P < 0.001, respectively). Interactions between haplotype and environmental factors had different effects on the incidence of dyslipidemia than those observed for each individual locus. Interactions between the rs2222896G-rs2548861G and rs2222896G-rs2548861T haplotypes and smoking, high BMI, or FBS increased the risk of dyslipidemia, as did the interaction between the rs2222896G-rs2548861T haplotype and sex or age (P < 0.05 for all). Furthermore, interactions between the rs2222896A-rs2548861T haplotype and sex, age, BMI, drinking, or smoking, and between the rs2222896G-rs2548861T haplotype and sex, age, BMI, drinking, or smoking, and between the rs2222896G-rs2548861T haplotype and BMI or FBS decreased the effects of the environmental factors on dyslipidemia (P < 0.05 for all).

Associations between $\mathbf{G} \times \mathbf{G}$ interactions and dyslipidemia

As shown in Table 8, thirteen *TUBB-WWOX* G × G interactions were observed in the normal and dyslipidemia groups (LEF for gene interactions > 0.03). Frequencies of nine of these G × G interactions, namely G-C-A-T, G-C-A-G, G-C-G-T, T-T-A-G, T-T-G-T, T-T-G-G, T-C-G-G, T-T-A-T, and T-C-G-T, differed significantly between the two groups (P < 0.05). Among them, only the T-T-G-T interaction was associated with increased incidence of dyslipidemia (OR = 1.371, 95% CI = 1.112–1.689, P = 0.003), while the other eight interactions, particularly T-T-G-G, were associated with reduced incidence of dyslipidemia (OR = 0.795, 95% CI = 0.640–0.988, P = 0.038).

Models of the effects of different interactions on dyslipidemia

Using generalized multifactor dimensionality reduction (GMDR) analysis and adjusting by covariates, we identified the fourteen best interactive models of SNP-SNP, haplotype-haplotype, $G \times G$, and $G \times E$ effects on dyslipidemia (Table 9). The two- to four-locus interactive models for SNP-SNP and SNP-environment were rs2222896-rs3130685, rs2222896-rs3130685-rs2548861, rs2222896-rs3130685-rs2548861-rs3132584, rs2222896rs3130685-age > 65 years, rs2222896-rs3132584-FBS ≥ 7.0 mM, and rs2222896-rs3132584-drinking (crossvalidation (CV) constancy of 10 of 10, balanced accuracy test (Bal. Acc.) \geq 64.98%, and permutation test P < 0.001for all). The two- to three-locus interactive models for haplotype-haplotype and haplotype-environment were H1-H3 (interaction of rs2222896A-rs2548861T and rs2222896A-rs2548861G haplotypes), H2-H3-H4 (interaction of rs2222896G-rs2548861G, rs2222896Ars2548861G, and rs2222896G-rs2548861T haplotypes), H1-H3-BMI > 24 kg/m², and H1-H3-FBS \geq 7.0 mM (CV constancy of 10 of 10, Bal. Acc. \geq 62.05%, and permutation test P < 0.001 for all). Additionally, the twoto three-locus models of $G \times G$ and $G \times E$ interactions

Tinid	Hanlatuna	Subjects	Data	4	Р	95%CI	of Beta
Lipid	Haplotype	Subjects	Beta	t	P	Lower	Upper
Normal							
TC	rs2222896G-rs2548861T	carriers/non-carriers	0.128	3.765	0.000	0.212	0.673
TG	rs2222896A-rs2548861G	carriers/non-carriers	-0.167	-4.912	0.000	-0.085	-0.036
HDL-C	rs2222896A-rs2548861G	carriers/non-carriers	0.151	4.414	0.000	0.034	0.089
	rs2222896G-rs2548861G	carriers/non-carriers	-0.124	-3.610	0.000	-0.074	-0.022
LDL-C	rs2222896G-rs2548861T	carriers/non-carriers	0.136	4.012	0.000	0.074	0.217
ApoA1	rs2222896A-rs2548861G	carriers/non-carriers	0.109	3.163	0.002	0.017	0.071
ApoA1/ApoB	rs2222896A-rs2548861G	carriers/non-carriers	0.095	2.772	0.006	0.020	0.116
Dyslipidemia							
TC	rs2222896A-rs2548861T	carriers/non-carriers	-0.138	-4.632	0.000	-0.290	-0.118
	rs2222896G-rs2548861G	carriers/non-carriers	-0.083	-2.796	0.005	-0.233	-0.041
TG	rs2222896A-rs2548861G	carriers/non-carriers	0.126	4.207	0.000	0.093	0.256
HDL-C	rs2222896A-rs2548861T	carriers/non-carriers	0.136	4.530	0.000	0.026	0.067
	rs2222896G-rs2548861T	carriers/non-carriers	-0.193	-6.556	0.000	-0.099	-0.053
LDL-C	rs2222896A-rs2548861G	carriers/non-carriers	-0.120	-4.006	0.000	-0.135	-0.046
ApoA1	rs2222896A-rs2548861T	carriers/non-carriers	0.121	4.053	0.000	0.020	0.058
	rs2222896A-rs2548861G	carriers/non-carriers	0.083	2.773	0.006	0.009	0.054
	rs2222896G-rs2548861T	carriers/non-carriers	-0.117	-3.931	0.000	-0.064	-0.022
ApoB	rs2222896A-rs2548861T	carriers/non-carriers	-0.117	-3.913	0.000	-0.036	-0.012
ApoA1/ApoB	rs2222896A-rs2548861T	carriers/non-carriers	0.142	4.769	0.000	0.040	0.097
	rs2222896G-rs2548861T	carriers/non-carriers	-0.106	-3.570	0.000	-0.091	-0.026

Table 7. Association between WWOX haplotypes and serum lipid levels in normal and dyslipidemia groups.

TC, total cholesterol; TG, triglyceride; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; ApoA1, apolipoprotein A1; ApoB, apolipoprotein B; CI, confidence interval. *P* values were calculated by multivariable linear regression. P < 0.0125 indicated a statistically significant difference after Bonferroni correction and adjusting for genotype, age, gender, drinking, smoking, fasting blood-glucose, and body mass index.

were I1-I2 (G-C-A-T and G-C-A-G), I3-I12 (G-C-G-T and T-T-A-T), I4-I8-I11 (G-C-G-G, T-T-G-T, and T-C-G-G), I4-I5-I10 (G-C-G-G, G-T-A-T, and G-T-A-G), and I1-I11-BMI > 24 kg/m² (CV constancy of 10 of 10, Bal. Acc. \geq 56.87%, and permutation test *P* < 0.001 for all).

Furthermore, associations between the fourteen interactions and dyslipidemia were further verified through logistic regression analysis as shown in Figure 5 and Table 10. The following interactions between genotypes, haplotypes, and factors increased the risk of dyslipidemia: rs3130685TT-rs2548861TG+TT (OR = 3.099, 95% CI = 2.397-4.005, P < 0.001) and rs3130685CT+CC-rs2548861GG (OR = 3.136, 95% CI = 2.489–3.952, P < 0.001) compared to rs3130685TTrs2548861GG; rs3130685TT-rs2222896AG+GG (OR = 2.532, 95% CI = 1.982 - 3.263, P < 0.001) and rs3132584GT+GG-rs2222896AG+GG (OR = 2.548, 95% CI = 2.035 - 3.189, P < 0.001) compared to rs3130685TTrs2222896AA; rs2222896AG+GG-nondrinking compared to rs2222896AA-nondrinking (OR = 1.792, 95% CI = 1.490–2.155, P < 0.001); H1-H3 compared to H1–H3 non-carriers (OR = 3.315, 95% CI = 2.681-4.098, P < 0.001); I4 non-carriers-I10 carriers (OR = 2.295, 95% CI

= 1.783–2.954, *P* < 0.001) and I4 carriers-I10 non-carriers (OR = 2.877, 95% CI = 2.253 - 3.675, P < 0.001)compared to I4-I10 non-carriers; and I4 non-carriers-I8 carriers (OR = 5.292, 95% CI = 3.694-7.582, P < 0.001) and I4 carriers-I8 non-carriers (OR = 1.465, 95%) CI = 1.219 - 1.760, P < 0.001) compared to I4-I8 non-carriers. In contrast, interactions between the following genotypes, phenotypes, and factors decreased of dyslipidemia: the risk rs3132584GT+GGrs2222896AA compared to rs3132584TT-rs2222896AA (OR = 0.441, 95% CI = 0.345 - 0.563, P < 0.001);rs2222896AA-drinking compared to rs2222896AAnondrinking (OR = 0.298, 95% CI = 0.196-0.455, P < 0.001); and I1-I1 carriers compared to I1-I11 non-carriers (OR = 0.114, 95% CI = 0.097 - 0.164, P < 0.001).

Risk of hypertension based on different interactive models of dyslipidemia

The effects of traditional and potential genetic risks associated with dyslipidemia on hypertension are shown in Tables 11, 12. In this study, dyslipidemia, female sex, age > 65 years, BMI > 24 kg/m², and FBS \ge 7.0 mM were clear risk factors for hypertension (*P* < 0.05).



Figure 4. Associations between *WWOX* haplotypes, environmental factors, and their interactions on dyslipidemia. **P*-value < 0.05 and ***P*-value < 0.001.

Label	Ge	ne-gene	interact	ion	No	ormal	Dysli	pidemia	χ^2	Р	OR (95% CI)
	G1	G2	G3	G4							
I 1	G	С	А	Т	412	(0.238)	260	(0.115)	128.782	0.000	0.328(0.217-0.398)
I 2	G	С	А	G	489	(0.283)	449	(0.199)	55.068	0.000	0.506(0.423-0.606)
I 3	G	С	G	Т	375	(0.217)	388	(0.172)	16.853	0.000	0.683(0.569-0.819)
I 4	G	С	G	G	479	(0.277)	612	(0.271)	0.300	0.584	0.951(0.796–1.137)
I 5	G	Т	А	Т	357	(0.207)	388	(0.172)	1.800	0.180	0.884(0.373-1.059)
I 6	G	Т	G	G	523	(0.303)	634	(0.281)	3.847	0.050	0.835(0.697-1.000)
I 7	Т	Т	А	G	185	(0.107)	196	(0.087)	5.180	0.023	0.771(0.616-0.965)
I 8	Т	Т	G	Т	185	(0.107)	307	(0.136)	8.761	0.003	1.371(1.112–1.689)
I 9	Т	Т	G	G	199	(0.115)	217	(0.096)	4.926	0.038	0.795(0.640-0.988)
I 10	G	Т	А	G	470	(0.272)	576	(0.255)	2.241	0.134	0.873(0.731-1.043)
I 11	Т	С	G	G	257	(0.149)	108	(0.048)	121.925	0.000	0.250(0.195-0.320)
I 12	Т	Т	А	Т	234	(0.135)	180	(0.080)	36.219	0.000	0.511(0.410-0.636)
I 13	Т	С	G	Т	211	(0.122)	102	(0.045)	81.633	0.000	0.307(0.238-0.397)

Table 8. Frequencies of gene-gene interactions in the normal and dyslipidemia groups [n (frequency)].

G1, *TUBB* rs3132584 G>T; G2, *TUBB* rs3130685 C>T; G3, *WWOX* rs2222896 A>G; G4, *WWOX* rs2548861 T>G; I, interaction. Gene interaction frequencies < 0.03 in the two groups were excluded.

Interactive model	Training Bal. Acc.	Testing Bal. Acc.	CV consistency	Sign test P	Permutation test P
SNP-SNP					
A-B	0.7189	0.7208	10/10	0.001	< 0.0001
A-B-C	0.8061	0.8043	10/10	0.001	< 0.0001
A-B-C-D	0.8434	0.8290	10/10	0.001	< 0.0001
SNP-environment					
A-B-F	0.7189	0.7208	10/10	0.001	< 0.0001
A-D-H	0.6492	0.6509	10/10	0.001	< 0.0001
A-D-J	0.6483	0.6498	10/10	0.001	< 0.0001
Haplotype–haplotype					
H1-H3	0.6176	0.6176	10/10	< 0.0001	< 0.0001
Haplotype-environment					
H1-H3-G	0.6469	0.6469	10/10	< 0.0001	< 0.0001
Н1-Н3-Н	0.6381	0.6369	10/10	< 0.0001	< 0.0001
Gene-gene					
I1-I2	0.6233	0.6233	10/10	< 0.0001	< 0.0001
I3-I12	0.5687	0.5687	10/10	< 0.0001	< 0.0001
I4-I8-I11	0.6950	0.6950	10/10	< 0.0001	< 0.0001
I4-I5-I10	0.6488	0.6488	10/10	< 0.0001	< 0.0001
Gene-environment					
I1-I11-G	0.6302	0.6268	10/10	< 0.0001	< 0.0001

A = rs2222896, B = rs3130685, C = rs2548861, D = rs3132584, E = female, F = age > 65 years, G = body mass index > 24 kg/m², H = fasting blood-glucose \geq 7.0 mM, I = smoking, J = drinking; Bal. Acc., balanced accuracy; CV, cross-validation.

Moreover, interactions between the rs3132584 and rs2222896 SNPs increased the risk of hypertension 1.523-fold (P < 0.006) after adjustment for BMI, sex, age, and FBS.

DISCUSSION

Dyslipidemia, a multifactorial non-communicable chronic disease, contributes to pathogenesis of arteriosclerotic diseases and is characterized by high TC. TG, and LDL-C and low HDL-C levels [6]. LDL-C and other ApoB-rich lipoproteins accumulate in the arterial wall, playing a key role in atherosclerotic plaque formation and subsequent related cardiovascular events [19]. Studies have shown that the risk of developing carotid plaque increases by 62% for every 1 mM increase in serum LDL-C concentration [20]. Lipid levels in the plasma are influenced by many factors, and genetic variation in lipid-related genes leads to diverse phenotypes in different countries and ethnicities. Previous GWASs demonstrated associations between the TUBB and WWOX genes and LDL-C and HDL-C levels, and differences in the expression of these two genes in different populations contributes to diversity in lipid traits. According to recently updated data from 1000Genomes (http://www.ncbi.nlm.nih.gov/snp/), alternate allele frequencies in American, European, East Asian, and African populations, respectively, are as follows: TUBB rs3132584T, 0.1800, 0.1769, 0.1796, and 0.4803; TUBB rs3130685T, 0.5320, 0.4632, 0.5367, and 0.5479; WWOX rs2222896G, 0.5750, 0.6252, 0.6131, and 0.8313; and WWOX rs2548861G, 0.6341, 0.5656, 0.6282, and 0.2829. In this study, the frequencies of these allelic mutations in the Maonan population differed significantly from their frequencies in the CHB

population: frequencies of the rs3132584T, rs3130685T, rs2222896G, and rs2548861G alleles in the Maonan and the CHB populations, respectively, were 0.2168 vs. 0.1829, 0.5730 vs. 0.5444, 0.5607 vs. 0.6341, and 0.6184 vs. 0.5488. Moreover, the frequency of the mutant homozygous TT genotype at rs3132584 was 0.0471 in the Maonan population but 0 in the CHB population, further demonstrating the differences in gene distribution between these two ethnic groups. Furthermore, although genotypic frequencies for rs3132584, rs3130685, and rs2222896 in the Maonan population differed significantly between the normal and dyslipidemia groups, only rs2222896 genotype was correlated with various serum lipid parameters. Normal group subjects with the rs2222896GG genotype had higher LDL-C and lower HDL-C and ApoA1 concentrations than those with other genotypes. This association between rs2222896 and serum LDL-C level was consistent with a previous study [21]. Additionally, Yamada et al. identified two novel loci in TUBB, rs3132584 and rs3130685, that were associated with high LDL-C in an exome-wide association study of early-onset dyslipidemia [13]. Another study of 2911 Japanese subjects revealed that the reference alleles rs3132584A and rs3130685C were associated with high LDL-C (P < 0.0001). In this study, we found that a rs3132584 mutation from G to T was present at a higher frequency. In the dyslipidemia group, rs3132584G was associated with high LDL-C levels; however, we observed little association between rs3130685C and LDL-C concentration. Two previous studies [14, 22] indicated that the rs2548861 SNP in WWOX was associated with HDL-C concentration in the Young Finns Study population ($n = 6,728, P = 6.9 \times 10^{-7}$), but another [23] failed to replicate this result in the Spanish population (n = 801). In our study, an association

Strong synergy SNP-SNP	n:2222896	, SNP-Environment ,13132584 ,12222896 Ditaking	Synergy Gene-Gene	3
Week interactions Strong in	rs2548861 Interactions Week interactions	Strong interactions	Week interactions	Strong interactions
	ge>65years	Haplotype-Environment BHD-24kg/m2	Synergy Gene-Gene Moderate synergy Gene-Gene	9 n1
	szzzz896 iteractions Week interactions	H3 Strong interactions	Week interactions	18 Strong interactions
Redundancy SNP-Environment	Synergy Moderate synergy	Haplotype-Environment	Redundancy Gene-Environ	nment BMI>24kg/m2
n2	132584	H1		n
	222896 nteractions Week interactions	Strong interactions	Week interactions	Strong interaction

Figure 5. Optimal interactions affecting dyslipidemia shown in a dendrogram. Elements that interact strongly with each other appear close together in the leaves of the tree, while elements that interact weakly appear far from each other.

Variable 1	Variable 2	OR (95% CI)	Р	
SNP-SNP interaction				
rs3130685	rs2548861			
TT	GG	1	_	
TT	TG+TT	3.099(2.397-4.005)	0.000	
CT+CC	GG	3.136(2.489-3.952)	0.000	
CT+CC	TG+TT	0.588(0.489-0.706)	0.000	
rs3130685	rs2222896			
TT	AA	1	_	
TT	AG+GG	2.532(1.982-3.236)	0.000	
CT+CC	AA	1.317(0.929–1.867)	0.123	
CT+CC	AG+GG	1.236(1.027-1.486)	0.025	
rs3132584	rs2222896			
TT	AA	1	_	
TT	AG+GG	1.526×10 ⁹ (0.000–NA)	0.998	
GT+GG	AA	0.441(0.345-0.563)	0.000	
GT+GG	AG+GG	2.548(2.035-3.189)	0.000	
SNP-environment interaction				
rs2222896	Drinking			
AA	No	1	_	
AA	Yes	0.298(0.196-0.455)	0.000	
AG+GG	No	1.792(1.490-2.155)	0.000	
AG+GG	Yes	1.344(1.077–1.678)	0.009	
Haplotype-haplotype interacti	on			
H1	H3			
Non-carriers	Non-carriers	3.315(2.681-4.098)	0.000	
Non-carriers	Carriers	0.704(0.563-0.881)	0.002	
Carriers	Non-carriers	0.485(0.403-0.583)	0.000	
Carriers	Carriers	1	_	
Gene-gene interaction				
I1	I11			
Non-carriers	Non-carriers	1	_	
Non-carriers	Carriers	1.271(0.862–1.875)	0.226	
Carriers	Non-carriers	0.808(0.648–1.006)	0.057	
Carriers	Carriers	0.114(0.079–0.164)	0.000	
I4	18			
Non-carriers	Non-carriers	1	_	
Non-carriers	Carriers	5.292(3.694-7.582)	0.000	
Carriers	Non-carriers	1.465(1.219–1.760)	0.000	
Carriers	Carriers	0.394(0.294–0.529)	0.000	
I4	I10			
Non-carriers	Non-carriers	1	_	
Non-carriers	Carriers	2.295(1.783-2.954)	0.000	
Carriers	Non-carriers	2.877(2.253-3.675)	0.000	
Carriers	Carriers	0.483(0.399–0.586)	0.000	

Table 10. Risk of dyslipidemia with different types of interactions.

H1, rs2222896A-rs2548861T haplotype; H3, rs2222896A-rs2548861G haplotype; I1, rs3132584G-rs3130685C-rs2222896A-rs2548861T; I4, rs3132584G-rs3130685C-rs2222896G-rs2548861G; I8, rs3132584T-rs3130685T-rs2222896G-rs2548861T; I10, rs3132584G-rs3130685T-rs2222896A-rs2548861G; I11, rs3132584T-rs3130685C-rs2222896G-rs2548861G; OR, odds ratio; CI, confidence interval. Different types of interactions were analyzed by logistic regression. P < 0.006 indicated a statistically significant difference after Bonferroni correction and adjusting for body mass index and fasting blood-glucose.

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Table 11. Risk of hypertension in the study population [n(frequency)].

Risk factor	Normal (n)	Hypertension (n)	χ^2	Р	OR (95% CI)
Female	1017(0.510)	976(0.490)	3.936	0.047	1.235(1.003-1.522)
Smoking	1442(0.724)	551(0.276)	0.133	0.715	0.956(0.751-1.217)
Drinking	1448(0.727)	545(0.273)	1.303	0.254	1.149(0.905-1.457)
Dyslipidemia	864(0.434)	1129(0.566)	42.037	0.000	1.884(1.556-2.282)
Age > 65 years	1544(0.775)	449(0.225)	55.483	0.000	2.369(1.888-2.927)
$BMI > 24 \text{ kg/m}^2$	1293(0.649)	700(0.351)	25.641	0.000	1.660(1.364-2.020)
$FBS \ge 7.0 \text{ mM}$	1725(0.866)	268(0.134)	29.408	0.000	2.139(1.625-2.816)

BMI, body mass index; FBS, fasting blood-glucose; OR, odds ratio; CI, confidence interval. P < 0.05 indicated a statistically significant difference.

Variable 1	Variable 2	OR (95% CI)	Р	
SNP-SNP interaction				
rs3130685	rs2548861			
TT	GG	1	_	
CT+CC	TG+TT	0.936(0.771-1.137)	0.506	
rs3130685	rs2222896			
TT	AA	1	_	
CT+CC	AG+GG	0.895(0.739-1.085)	0.259	
rs3132584	rs2222896			
TT	AA	1	_	
GT+GG	AG+GG	1.523(1.189-1.950)	0.001	
SNP-environment inter	action	· · · · ·		
rs2222896	Drinking			
AA	No	1	_	
AG+GG	Yes	1.264(0.997-1.603)	0.053	
Haplotype-haplotype in	nteraction			
H1	H3			
Non-carriers	Non-carriers	0.917(0.525-1.601)	0.761	
Carriers	Carriers	1	_	
Gene-gene interaction				
I1	I11			
Non-carriers	Non-carriers	1	_	
Carriers	Carriers	0.448(0.202-0.993)	0.048	
I4	I8			
Non-carriers	Non-carriers	1	_	
Carriers	Carriers	0.819(0.417-1.609)	0.562	
I4	I10	. ,		
Non-carriers	Non-carriers	1	_	
Carriers	Carriers	0.650(0.411-1.026)	0.064	

Table 12. Effects of meaningful interactive models of dyslipidemia on hypertension.

H1, rs2222896A-rs2548861T haplotype; H3, rs2222896A-rs2548861G haplotype; I1, rs3132584G-rs3130685C-rs2222896A-rs2548861T; I4, rs3132584G-rs3130685C-rs2222896G-rs2548861G; I8, rs3132584T-rs3130685T-rs2222896G-rs2548861G; I11, rs3132584T-rs3130685C-rs2222896G-rs2548861G; OR, odds ratio; CI, confidence interval. Different types of interactions were analyzed by logistic regression. P < 0.006 indicated a statistically significant difference after Bonferroni correction and adjusting for body mass index and fasting blood-glucose.

between rs2548861 and HDL-C concentration was observed in the dyslipidemia group (n = 1129, P = 0.001) but not in the normal group (n = 864, P = 0.597). We also found that HDL-C concentration in the dyslipidemia group might be affected by other non-genetic factors. Thus, we speculated that mutations at rs2548861 were not correlated with HDL-C concentration in these subjects. A correlation analysis between genetic models and disease risk was then conducted to understand the relationship between alleles at these four SNP sites and dyslipidemia. We found that codominant, dominant, recessive, and log-additive models of the rs3132584, rs3130685, and rs2222896 SNPs identified significant differences in dyslipidemia. Using the log-additive models, we identified the direction of the associations between each minor allele frequency (MAF) of these three SNPs and dyslipidemia. The rs3132584T and rs2222896A alleles were potential protective factors, while rs3130685C was a risk factor, for dyslipidemia. Various lipid-related SNPs and the frequencies of different alleles in the TUBB and WWOX genes can therefore lead to different clinical lipid phenotypes in different populations.

Genetic mechanisms are influenced by many types of interactions [24–26], including $G \times G$ and $G \times E$ interactions, which can have synergistic or contrasting effects on gene expression [27, 28]. Interactions among genes can include SNP-SNP interactions between individual genes, haplotype-haplotype interactions, and the combined effects of mutations in genes on different chromosomes [29, 30]. Detailed information on the four target SNPs was obtained from the 1000 Genome database. It indicated that there was a large gap between rs3132584 and rs3130685 at chromosomal positions 30720650 and 31238429, respectively, that prevented LD between them. In contrast, rs2222896 and rs2548861 at chromosomal positions 78058601 and 78624496, respectively, were about 500 kb from each other, resulting in a weak LD between them. Of the four common haplotypes observed for rs2222896 and rs2548861 alleles. rs2222896A-rs2548861T and rs2222896A-rs2548861G were associated with reduced risk of dyslipidemia while rs2222896G-rs2548861G and rs2222896G-rs2548861T were associated with elevated risk for dyslipidemia. In addition, multi-site mutations in TUBB and WWOX altered the incidence of dyslipidemia in the Maonan population. Thirteen types of multi-site mutation combinations for the two genes were screened using SHEsisPlus online software (http://shesisplus.bio-x.cn/SHEsis.html). Among these 13 combinations, rs3132584T-rs3130685T-rs2222896G -rs2548861T increased the risk of dyslipidemia 1.371fold; all of the remaining 12 combinations could potentially decrease the risk of dyslipidemia. Although TUBB and WWOX are located on two different

chromosomes, interaction models were successfully screened using the GMDR method (https://sourceforge. net/projects/gmdr/), and the effects of $G \times G$ and $G \times E$ interactions on dyslipidemia were examined further. The fourteen highest-performing models of interactions linked to dyslipidemia were identified; all had a 10 of 10 CV, training and testing balanced accuracy values greater than 0.5, and sign and permutation test *P* values less than 0.05. Furthermore, an intuitive interactive dendrogram and logistic regression analysis indicated that the rs3130685-rs2548861, rs3130685-rs2222896, H1-H3, I4-I8, and I4-110 interactions exhibited strong synergistic effects increasing the risk of dyslipidemia, while the rs3132584-rs2222896 and I1-I11 interactions decreased the risk of dyslipidemia. This study therefore demonstrated that WWOX haplotype and the combined effects of $TUBB \times WWOX$ have larger impacts on serum lipid levels and dyslipidemia than single SNPs in those genes [31]; additional experiments with larger samples sizes are needed to confirm this finding.

Many environmental factors, including aging, sex, unhealthy lifestyle (high-fat diet [32, 33], smoking, drinking), high blood glucose, and weight and obesity, are directly or indirectly involved in the pathogenesis of dyslipidemia. Moreover, these environmental factors can interact with lipid-related genes to alter lipid expression profiles [34, 35]. Several studies demonstrate that the incidence of hyperlipidemia is higher in elderly people (1259/1657, 76%, mean age 69 years) than in young people (203/494, 41%, mean age 29 years); this difference was attributed to age-associated declines in ribosome coverage in the vicinity of start codons and increases near stop codons, alterations in expression of genes associated with lipid metabolism, loss of hepatic LDL receptors, and decreases in sex hormone levels [36, 37]. Furthermore, the risk of dyslipidemia was higher in postmenopausal women and elderly men [38, 39]; food pickling processes can increase concentrations of secondary metabolites in lemons and onions, thus endangering health [40]; and heavy (8 or more drinks per week for women or 15 or more drinks per week for men) rather than low or moderate alcohol consumption was also linked to dangerous increases in lipid levels [41]. The Maonan ethnic group is located mainly in the Huanjiang region, which is known as the "hometown of cattle and grain" due to the prevalence of agriculture and beef cattle industries. Beef, duck, half-cooked chicken, animal offal, and pickled sour pork, snails, and vegetables, are common dishes in this region. In addition, consumption of wine made from rice, corn, sweet potatoes, pumpkins, etc., three times per day with meals and smoking cigarettes made from local dried tobacco leaves is common among Maonan men. Many high-risk environmental factors may therefore affect the Maonan population. We used random stratified sampling in our

examination of dyslipidemia risk in this population to eliminate systematic differences in age and gender structure between the two groups. We found no differences in either proportion of subjects > 65 years old or severity of smoking between subgroups, but severity of alcohol consumption, BMI > 24 kg/m², and FBS \ge 7.0 mM were significantly higher in the dyslipidemia group than in the normal group, and these factors interacted with the TUBB and WWOX SNPs to affect the prevalence of dyslipidemia. Pairwise multiple regression interaction models and optimal interaction models from GMDR revealed that the rs2222896G-rs2548861G haplotype interacted with smoking, overweight/obesity, or FBS \geq 7.0 mM, while the rs2222896G-rs2548861T haplotype interacted with old age, female sex, smoking, $FBS \ge 7.0$ mM, or overweight status/obesity to exacerbate the risk of dyslipidemia. In addition, age > 65 years increased the synergistic effect of rs2222896-rs3130685, and FBS \geq 7.0 mM increased the synergistic effects of H1-H3 or rs2222896-rs3132584 on dyslipidemia. In contrast, interactions between the rs2222896A-rs2548861T haplotype and female sex, old age, smoking, and drinking, and between the rs2222896A-rs2548861G haplotype and overweight status/obesity or FBS \geq 7.0 mM, decreased the risk of dyslipidemia. Finally, overweight status/obesity decreased the protective effects of H1-H3 or I1-I11 interactions against dyslipidemia, and drinking weakened the increased risk of dyslipidemia associated with rs2222896, possibly because most subjects were low-moderate drinkers.

Blood pressure is also modified by multiple genetic and environmental factors and their interactions [42–44], and dyslipidemia is a common independent risk for hypertension [45]. We therefore further explored differences in the incidence of hypertension between the two groups and found that the prevalence of hypertension was associated with gender, age, BMI, FBS, smoking, drinking, and dyslipidemia. Interestingly, we also found that the rs3132584 and rs2222896 SNP interaction that affected risk of dyslipidemia also significantly increased the risk of hypertension; this might be a novel explanation of how dyslipidemia contributes to the pathogenesis of hypertension.

Some important strengths and weaknesses of this study should be considered when interpreting the results. Notable strengths include the following: (1) identification of differences in *TUBB* and *WWOX* mutation frequencies between the Maonan ethnic group and the CHB population provides novel data for human genomic studies; (2) detection of *TUBB-WWOX* interactions and $G \times E$ interactions that affected the prevalence of dyslipidemia might provide new therapeutic targets for dyslipidemia; and (3) this study characterized an integrated effect of the *TUBB* rs3132584 and *WWOX* rs2222896 SNPs on dyslipidemia that could also predict the risk of hypertension, identifying a possible novel pathogenic mechanism by which dyslipidemia can lead to hypertension. However, the following limitations should also be considered: (1) this study did not include other variable risk factors, such as daily exercise habits and anti-hyperlipidemia interventions; (2) despite the unique diet (high salt and sour, high fat and alcohol) associated with the Maonan population, intake of these foods was not accurately characterized and their effects on dyslipidemia could not be examined; (3) larger sample sizes are needed to verify the results of this study; and (4) biological function studies are needed to further examine the effects of interactions between these two genes on dyslipidemia.

In summary, this study identified associations between *TUBB*, *WWOX*, environmental factors, serum lipid levels, and dyslipidemia in the Maonan population. Our findings revealed that, compared to single-locus effects, *TUBB-WWOX*-environment interactions can result in synergistic or contrasting effects on incidence of dyslipidemia, thereby increasing or reducing the risk of dyslipidemia.

MATERIALS AND METHODS

Participants

A total of 1993 unrelated subjects were selected from our previous sample library in 2015 through stratified random sampling [46]; all subjects were from three generations of the Maonan ethic group and lived in the Maonan Autonomous County of Huanjiang in Southern China. Subjects were separated into normal lipid and dyslipidemia groups with similar age and gender distributions. The mean age of the 864 normal group subjects was 55.31 ± 13.08 years; 431 (49.88%) were males and 433 (50.12%) were females. The mean age of the 1129 dyslipidemia group subjects was 56.04 ± 12.88 years; 545 (48.27%) were males and 584 (51.73%) were females. All subjects were generally healthy without a history of diabetes, thyroid disease, autoimmune disease, cardiovascular disease, stroke, renal disease, tumor, or mental illness. The participants did not take medicine or eat foods with known lipid-lowering properties. The study was reviewed and approved by the Ethics Committee of the First Affiliated Hospital, Guangxi Medical University (No. Lunshen 2014-KY-Guoji-001; March 7, 2014). Subjects who met inclusion criteria and signed informed consent were enrolled after professional investigators explained the experimental rules.

Laboratory values

Serum lipid levels were measured in peripheral blood (3 mL) collected in the morning after fasting for more than

8 hours using an autoanalyzer (type 7170A; Hitachi Ltd., Tokyo, Japan). Serum TC, TG, HDL-C, and LDL-C levels were determined using commercially available enzymatic assay kits, and serum ApoA1 and ApoB levels were determined by turbidimetric immuno-assay [46–48].

Variable definitions

Dyslipidemia is defined by elevated levels of TC, LDL-C, and TG and decreased HDL-C levels [6]. In this study, normal ranges for serum TC, TG, HDL-C, LDL-C, ApoA1, and ApoB levels and ApoA1/ApoB ratio were 3.10-5.17 mM, 0.56-1.70 mM, 0.90-1.81 mM, 2.70-3.10 mM, 1.00-1.78 g/L, 0.63-1.14 g/L, and 1.00-2.50, respectively. Dyslipidemia was therefore defined by any of the following either alone or in combination: TC > 5.17 mM, TG > 1.7 mM, LDL-C >3.10 mM, and/or HDL-C < 0.9 mM [49]. Blood pressure was measured after the subject had been sitting for at least 5 minutes. An average SBP \geq 140 mmHg and/or $DBP \ge 90 \text{ mmHg}$ for three measurements over two days was defined as hypertension [50]. BMI was determined by dividing weight (kilograms) by height (meters squared); overweight and obese categories in China [51] are defined as BMI > 24 kg/m² and BMI \ge 28 kg/m², respectively.

SNP selection

The basic principles of gene and SNP selection were as follows: (1) *TUBB* and *WWOX* were selected from previous GWASs examining lipid metabolism. (2) SNPs correlated with lipid levels were selected using Haploview (Broad Institute of MIT and Harvard, Cambridge, MA, USA version 4.2). (3) SNP information was obtained from NCBI dbSNP Build 132 (http://www.ncbi.nlm.nih.gov/snp/). (4) Only SNPs with MAF greater than 10% were included. (5) Correlations between the identified *TUBB* rs3132584 and rs3130685 SNPs and *WWOX* rs2222896 and rs2548861 SNPs and blood lipids were confirmed in previous studies [13, 14, 22, 23].

DNA quality control and genotyping

We used a phenol-chloroform method to extract DNA from peripheral blood. DNA was stored at -80° C until 10 μ L (10 ng/ μ L) of each sample was sent to the Department for Next-Generation Sequencing, Sangon Biotech Co., Ltd. (Shanghai, China). The absorbance of the DNA sample was measured at 260 nm and 280 nm using a Shimadzu UV-1601 spectrophotometer. An A260 nm/A280 nm ratio of = 1.8 indicates a pure DNA sample. SNP genotyping was performed on HiSeq XTen sequencers (Illumina, San Diego, CA, USA). The forward and

reverse primers for amplification of the four SNPs via polymerase chain reaction were as follows: 5'-AGATC GGTGCCAAGGTAAGAAT-3' and 5'-CAAACCCGAA GAGCCCTTTTACTA-3' for rs3132584; 5'-TGTCAA ATGCCATTCTTTAAACCTCATGT-3' and 5'-TTGGG ACACTTTACTCCTGAATC-3' for rs3130685; 5'-AGC TAAAGCCTGTAGACCATCAACATA-3' and 5'-TCC TCTTTAACACAGGTGAAATGCA-3' for rs2222896; and 5'-CCAAACCCTAAATAATTGTCTGGATGTT-3' and 5'-TCTGAATTTCTGCCAAGTTAGATG-3' for rs2548861.

Statistical analyses

SPSS software, version 25 (SPSS Inc., Chicago, IL, USA) was used to perform statistical analyses. Normally distributed data are presented as means ± standard deviation (SD) and differences between groups were identified using Student's unpaired *t*-test and one-way of variance (ANOVA). analysis Non-normally distributed data are shown as interguartile ranges and medians and differences between groups were identified using Mann-Whitney nonparametric tests. Differences in qualitative variables, including different ratios and HWE between the two groups, were analyzed using Chi square tests. Associations between genotypes or haplotypes and continuous serum lipid level data were assessed by multivariable linear regression. Differences in serum lipid levels associated with genotypes and haplotypes were considered statistically significant at P < 0.0125, and G \times G or $G \times E$ interactions were considered significant at P < 0.006 (corresponding to P < 0.05 after Bonferroni correction for four or eight independent tests). OR values and 95% CIs were calculated by multiple logistic regression after adjustment for stratified risk factors including age, sex, tobacco and alcohol consumption, BMI, and FBS. HWE, genotypic and haplotypic frequencies, and D' and R^2 values used to describe LD were calculated using SHEsisPlus online software. GMDR online software was used to screen optimal SNP-SNP, haplotype-haplotype, $G \times G$, and $G \times E$ interaction models. All other data visualizations were generated using GraphPad Prism (version 8.0.0).

Abbreviations

Apo: apolipoprotein; Bal. Acc.: balanced accuracy; BMI: body mass index; CAD: coronary arteriosclerotic diseases; CHB: Chinese Han Beijing; CI: confidence interval; CV: cross-validation; DBP: diastolic blood pressure; GMDR: generalized multifactor dimensionality reduction; GWAS: genome-wide association study; HDL-C: high-density lipoprotein cholesterol; HWE: Hardy Weinberg Equilibrium; LD: linkage disequilibrium; LDL-C: low-density lipoprotein cholesterol; LFT: lowest frequency threshold; MAF:

minor allele frequency; OR: odds ratio; PP: pulse pressure; FBS: fasting blood-glucose; RPKM: reads per kilobase per million mapped reads; SBP: systolic blood pressure; SNP: single nucleotide polymorphism; TC: total cholesterol; TG: triglyceride; *TUBB*: tubulin beta class I gene; *WWOX*: WW domain-containing oxidoreductase gene.

AUTHOR CONTRIBUTIONS

C.-X.L. conceived and designed the study, collected data, performed statistical analyses, and drafted the manuscript. R.-X.Y. improved the design of study, organized the epidemiology investigation, collected samples, and further revised the manuscript. Z.-H.S. collected data and performed statistical analyses. P.-F.Z., G.-X.D., Y.-Z.G., and B.-L.W. collected samples. All authors read and approved the final manuscript.

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CONFLICTS OF INTEREST

The authors have no financial or other conflicts of interest to report.

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REFERENCES

- Bucholz EM, Rodday AM, Kolor K, Khoury MJ, de Ferranti SD. Prevalence and predictors of cholesterol screening, awareness, and statin treatment among US adults with familial hypercholesterolemia or other forms of severe dyslipidemia (1999-2014). Circulation. 2018; 137:2218–30. https://doi.org/10.1161/CIRCULATIONAHA.117.032321 PMID:29581125
- Noubiap JJ, Bigna JJ, Nansseu JR, Nyaga UF, Balti EV, Echouffo-Tcheugui JB, Kengne AP. Prevalence of dyslipidaemia among adults in Africa: a systematic review and meta-analysis. Lancet Glob Health. 2018; 6:e998–1007. https://doi.org/10.1016/S2214.100Y(18)20275.4

https://doi.org/10.1016/S2214-109X(18)30275-4 PMID:<u>30103999</u>

3. Halcox JP, Banegas JR, Roy C, Dallongeville J, De Backer G, Guallar E, Perk J, Hajage D, Henriksson KM, Borghi C.

Prevalence and treatment of atherogenic dyslipidemia in the primary prevention of cardiovascular disease in Europe: EURIKA, a cross-sectional observational study. BMC Cardiovasc Disord. 2017; 17:160. https://doi.org/10.1186/s12872-017-0591-5 PMID:28623902

 Poorzand H, Tsarouhas K, Hozhabrossadati SA, Khorrampazhouh N, Bondarsahebi Y, Bacopoulou F, Rezaee R, Jafarzadeh Esfehani R, Morovatdar N. Risk factors of premature coronary artery disease in Iran: a systematic review and meta-analysis. Eur J Clin Invest. 2019; 49:e13124. https://doi.org/10.1111/eci.13124 PMID:31038733

 Gai Z, Wang T, Visentin M, Kullak-Ublick GA, Fu X, Wang Z. Lipid accumulation and chronic kidney disease. Nutrients. 2019; 11:722. <u>https://doi.org/10.3390/nu11040722</u> PMID:30925738

 Mach F, Baigent C, Catapano AL, Koskinas KC, Casula M, Badimon L, Chapman MJ, De Backer GG, Delgado V, Ference BA, Graham IM, Halliday A, Landmesser U, et al, and ESC Scientific Document Group. 2019 ESC/EAS guidelines for the management of dyslipidaemias: lipid modification to reduce cardiovascular risk. Eur Heart J. 2020; 41:111–88. https://doi.org/10.1093/eurheartj/ehz455

https://doi.org/10.1093/eurheartj/ehz455 PMID:<u>31504418</u>

- Jiang Z, Cui Y, Wang L, Zhao Y, Yan S, Chang X. Investigating citrullinated proteins in tumour cell lines. World J Surg Oncol. 2013; 11:260. <u>https://doi.org/10.1186/1477-7819-11-260</u> PMID:<u>24099319</u>
- Sferra A, Petrini S, Bellacchio E, Nicita F, Scibelli F, Dentici ML, Alfieri P, Cestra G, Bertini ES, Zanni G. TUBB variants underlying different phenotypes result in altered vesicle trafficking and microtubule dynamics. Int J Mol Sci. 2020; 21:1385. <u>https://doi.org/10.3390/ijms21041385</u>

PMID:<u>32085672</u>

 Yele V, Pindiprolu SK, Sana S, Ramamurty DS, Madasi JR, Vadlamani S. Synthesis and preclinical evaluation of indole triazole conjugates as microtubule targeting agents that are effective against MCF-7 breast cancer cell lines. Anticancer Agents Med Chem. 2020. [Epub ahead of print].

https://doi.org/10.2174/1871520620666200925102940 PMID:<u>32981511</u>

 Johannsen J, Kortüm F, Rosenberger G, Bokelmann K, Schirmer MA, Denecke J, Santer R. A novel missense variant in the SDR domain of the WWOX gene leads to complete loss of WWOX protein with early-onset epileptic encephalopathy and severe developmental delay. Neurogenetics. 2018; 19:151–56. https://doi.org/10.1007/s10048-018-0549-5 PMID:29808465

- Iatan I, Choi HY, Ruel I, Reddy MV, Kil H, Lee J, Odeh MA, Salah Z, Abu-Remaileh M, Weissglas-Volkov D, Nikkola E, Civelek M, Awan Z, et al. The WWOX gene modulates high-density lipoprotein and lipid metabolism. Circ Cardiovasc Genet. 2014; 7:491–504. <u>https://doi.org/10.1161/CIRCGENETICS.113.000248</u> PMID:<u>24871327</u>
- Khawaled S, Nigita G, Distefano R, Oster S, Suh SS, Smith Y, Khalaileh A, Peng Y, Croce CM, Geiger T, Seewaldt VL, Aqeilan RI. Pleiotropic tumor suppressor functions of WWOX antagonize metastasis. Signal Transduct Target Ther. 2020; 5:43. <u>https://doi.org/10.1038/s41392-020-0136-8</u> PMID:<u>32300104</u>
- Yamada Y, Kato K, Oguri M, Horibe H, Fujimaki T, Yasukochi Y, Takeuchi I, Sakuma J. Identification of 12 novel loci that confer susceptibility to early-onset dyslipidemia. Int J Mol Med. 2019; 43:57–82. <u>https://doi.org/10.3892/ijmm.2018.3943</u> PMID:<u>30365130</u>
- Weissglas-Volkov D, Pajukanta P. Genetic causes of high and low serum HDL-cholesterol. J Lipid Res. 2010; 51:2032–57. https://doi.org/10.1194/jlr.R004739 PMID:20421590
- Pan L, Yang Z, Wu Y, Yin RX, Liao Y, Wang J, Gao B, Zhang L, and China National Survey of Chronic Kidney Disease Working Group. The prevalence, awareness, treatment and control of dyslipidemia among adults in China. Atherosclerosis. 2016; 248:2–9. <u>https://doi.org/10.1016/j.atherosclerosis.2016.02.006</u> PMID:26978581
- 16. Liu CX, Yin RX, Shi ZH, Deng GX, Zheng PF, Wei BL, Guan YZ. EHBP1 SNPs, their haplotypes, and geneenvironment interactive effects on serum lipid levels. ACS Omega. 2020; 5:7158–69. <u>https://doi.org/10.1021/acsomega.9b03522</u> PMID:<u>32280856</u>
- Yang S, Yin RX, Miao L, Zhang QH, Zhou YG, Wu J. Association between the LIPG polymorphisms and serum lipid levels in the Maonan and Han populations. J Gene Med. 2019; 21:e3071. https://doi.org/10.1002/jgm.3071 PMID:30657227
- 18. Qiu L, Yin RX, Nie RJ, Hu XJ, Khounphinith E, Zhang FH. The CXCL12 SNPs and their haplotypes are associated with serum lipid traits. Sci Rep. 2019; 9:19524. <u>https://doi.org/10.1038/s41598-019-55725-3</u> PMID:<u>31862910</u>
- 19. Borén J, Chapman MJ, Krauss RM, Packard CJ, Bentzon JF, Binder CJ, Daemen MJ, Demer LL, Hegele RA, Nicholls SJ, Nordestgaard BG, Watts GF, Bruckert E,

et al. Low-density lipoproteins cause atherosclerotic cardiovascular disease: pathophysiological, genetic, and therapeutic insights: a consensus statement from the European Atherosclerosis Society Consensus Panel. Eur Heart J. 2020; 41:2313–30. https://doi.org/10.1093/eurheartj/ehz962 PMID:32052833

- Pan J, Liu J, Wang H, Li W, Du X, Lin Q, Zhang X, Qi D, Tu J, Ning X, Yang Q, Wang J. Association of carotid atherosclerosis with lipid components in asymptomatic low-income Chinese: a population-based crosssectional study. Front Neurol. 2020; 11:276. <u>https://doi.org/10.3389/fneur.2020.00276</u> PMID:<u>32390928</u>
- Kathiresan S, Manning AK, Demissie S, D'Agostino RB, Surti A, Guiducci C, Gianniny L, Burtt NP, Melander O, Orho-Melander M, Arnett DK, Peloso GM, Ordovas JM, Cupples LA. A genome-wide association study for blood lipid phenotypes in the Framingham heart study. BMC Med Genet. 2007 (Suppl 1); 8:S17. <u>https://doi.org/10.1186/1471-2350-8-S1-S17</u> PMID:<u>17903299</u>
- Lee JC, Weissglas-Volkov D, Kyttälä M, Dastani Z, Cantor RM, Sobel EM, Plaisier CL, Engert JC, van Greevenbroek MM, Kane JP, Malloy MJ, Pullinger CR, Huertas-Vazquez A, et al. WW-domain-containing oxidoreductase is associated with low plasma HDL-C levels. Am J Hum Genet. 2008; 83:180–92. <u>https://doi.org/10.1016/j.ajhg.2008.07.002</u> PMID:<u>18674750</u>
- Sáez ME, González-Pérez A, Martínez-Larrad MT, Gayán J, Real LM, Serrano-Ríos M, Ruiz A. WWOX gene is associated with HDL cholesterol and triglyceride levels. BMC Med Genet. 2010; 11:148. <u>https://doi.org/10.1186/1471-2350-11-148</u>
 PMID:20942981
- 24. Rand DM, Mossman JA. Mitonuclear conflict and cooperation govern the integration of genotypes, phenotypes and environments. Philos Trans R Soc Lond B Biol Sci. 2020; 375:20190188. <u>https://doi.org/10.1098/rstb.2019.0188</u> PMID:<u>31787039</u>
- Zan Y, Carlborg Ö. Dynamic genetic architecture of yeast response to environmental perturbation shed light on origin of cryptic genetic variation. PLoS Genet. 2020; 16:e1008801. <u>https://doi.org/10.1371/journal.pgen.1008801</u> PMID:32392218
- 26. Zhang ZH, Zhu YJ, Wang SL, Fan YY, Zhuang JY. Importance of the interaction between heading date genes Hd1 and Ghd7 for controlling yield traits in rice. Int J Mol Sci. 2019; 20:516. <u>https://doi.org/10.3390/ijms20030516</u> PMID:<u>30691093</u>

- Zheng PF, Yin RX, Liu CX, Deng GX, Guan YZ, Wei BL. SYNE1-QK1 SNPs, G × G and G × E interactions on the risk of hyperlipidaemia. J Cell Mol Med. 2020; 24:5772–85. https://doi.org/10.1111/jcmm.15239 PMID:32281752
- 28. Barghi N, Hermisson J, Schlötterer C. Polygenic adaptation: a unifying framework to understand positive selection. Nat Rev Genet. 2020; 21:769–81. <u>https://doi.org/10.1038/s41576-020-0250-z</u> PMID:32601318
- 29. Miao L, Yin RX, Pan SL, Yang S, Yang DZ, Lin WX. BCL3-PVRL2-TOMM40 SNPs, gene-gene and geneenvironment interactions on dyslipidemia. Sci Rep. 2018; 8:6189. <u>https://doi.org/10.1038/s41598-018-24432-w</u> PMID:29670124
- Deng GX, Yin RX, Guan YZ, Liu CX, Zheng PF, Wei BL, Wu JZ, Miao L. Association of the NCAN-TM6SF2-CILP2-PBX4-SUGP1-MAU2 SNPs and gene-gene and geneenvironment interactions with serum lipid levels. Aging (Albany NY). 2020; 12:11893–913. <u>https://doi.org/10.18632/aging.103361</u> PMID:32568739
- 31. Guo T, Yin RX, Pan L, Yang S, Miao L, Huang F. Integrative variants, haplotypes and diplotypes of the CAPN3 and FRMD5 genes and several environmental exposures associate with serum lipid variables. Sci Rep. 2017; 7:45119.

https://doi.org/10.1038/srep45119 PMID:28332615

- 32. Zhang Q, Fan XY, Guo WL, Cao YJ, Lin YC, Cheng WJ, Chen LJ, Rao PF, Ni L, Lv XC. The protective mechanisms of macroalgae laminaria japonica consumption against lipid metabolism disorders in high-fat diet-induced hyperlipidemic rats. Food Funct. 2020; 11:3256–70. https://doi.org/10.1039/d0fo00065e PMID:32219261
- Hu Q, Zhang H, Gutiérrez Cortés N, Wu D, Wang P, Zhang J, Mattison JA, Smith E, Bettcher LF, Wang M, Lakatta EG, Sheu SS, Wang W. Increased Drp1 acetylation by lipid overload induces cardiomyocyte death and heart dysfunction. Circ Res. 2020; 126:456–70.

https://doi.org/10.1161/CIRCRESAHA.119.315252 PMID:<u>31896304</u>

34. Jha CK, Mir R, Elfaki I, Javid J, Babakr AT, Banu S, Chahal SMS. Evaluation of the Association of Omentin 1 rs2274907 A>T and rs2274908 G>A Gene Polymorphisms with Coronary Artery Disease in Indian Population: A Case Control Study. J Pers Med. 2019; 9:30.

https://doi.org/10.3390/jpm9020030 PMID:<u>31174318</u>

- 35. Wang XB, Cui NH, Zhang S, Guo SR, Liu ZJ, Ming L. PARP-1 variant Rs1136410 confers protection against coronary artery disease in a Chinese Han population: a two-stage case-control study involving 5643 subjects. Front Physiol. 2017; 8:916. <u>https://doi.org/10.3389/fphys.2017.00916</u> PMID:29184509
- 36. Rosada A, Kassner U, Weidemann F, König M, Buchmann N, Steinhagen-Thiessen E, Spira D. Hyperlipidemias in elderly patients: results from the Berlin Aging Study II (BASEII), a cross-sectional study. Lipids Health Dis. 2020; 19:92. <u>https://doi.org/10.1186/s12944-020-01277-9</u> PMID:32410691
- Anisimova AS, Meerson MB, Gerashchenko MV, Kulakovskiy IV, Dmitriev SE, Gladyshev VN. Multifaceted deregulation of gene expression and protein synthesis with age. Proc Natl Acad Sci USA. 2020; 117:15581–90. <u>https://doi.org/10.1073/pnas.2001788117</u> PMID:<u>32576685</u>
- Cho L, Davis M, Elgendy I, Epps K, Lindley KJ, Mehta PK, Michos ED, Minissian M, Pepine C, Vaccarino V, Volgman AS, and ACC CVD Womens Committee Members. Summary of updated recommendations for primary prevention of cardiovascular disease in women: JACC state-of-the-art review. J Am Coll Cardiol. 2020; 75:2602–18. <u>https://doi.org/10.1016/j.jacc.2020.03.060</u> PMID:32439010
- Barrientos G, Llanos P, Basualto-Alarcón C, Estrada M. Androgen-regulated cardiac metabolism in aging men. Front Endocrinol (Lausanne). 2020; 11:316. <u>https://doi.org/10.3389/fendo.2020.00316</u> PMID:<u>32499759</u>
- Farag MA, Tawfike AF, Donia MS, Ehrlich A, Wessjohann LA. Influence of pickling process on Allium cepa and Citrus limon metabolome as determined via mass spectrometry-based metabolomics. Molecules. 2019; 24:928. https://doi.org/10.3390/molecules24050928

PMID:30866428

- 41. Wood AM, Kaptoge S, Butterworth AS, Willeit P, Warnakula S, Bolton T, Paige E, Paul DS, Sweeting M, Burgess S, Bell S, Astle W, Stevens D, et al, and Emerging Risk Factors Collaboration/EPIC-CVD/UK Biobank Alcohol Study Group. Risk thresholds for alcohol consumption: combined analysis of individualparticipant data for 599 912 current drinkers in 83 prospective studies. Lancet. 2018; 391:1513–23. https://doi.org/10.1016/S0140-6736(18)30134-X PMID:29676281
- 42. Levy D, Larson MG, Benjamin EJ, Newton-Cheh C, Wang TJ, Hwang SJ, Vasan RS, Mitchell GF.

Framingham heart study 100K project: genome-wide associations for blood pressure and arterial stiffness. BMC Med Genet. 2007 (Suppl 1); 8:S3. https://doi.org/10.1186/1471-2350-8-S1-S3 PMID:<u>17903302</u>

- 43. Yu Z, Coresh J, Qi G, Grams M, Boerwinkle E, Snieder H, Teumer A, Pattaro C, Köttgen A, Chatterjee N, Tin A. A bidirectional mendelian randomization study supports causal effects of kidney function on blood pressure. Kidney Int. 2020; 98:708–16. <u>https://doi.org/10.1016/j.kint.2020.04.044</u> PMID:<u>32454124</u>
- 44. Huang Y, Ollikainen M, Muniandy M, Zhang T, van Dongen J, Hao G, van der Most PJ, Pan Y, Pervjakova N, Sun YV, Hui Q, Lahti J, Fraszczyk E, et al. Identification, heritability, and relation with gene expression of novel DNA methylation loci for blood pressure. Hypertension. 2020; 76:195–205. https://doi.org/10.1161/HYPERTENSIONAHA.120.14973 PMID:32520614
- 45. Arnett DK, Graf GA. Metabolomics, lipid pathways, and blood pressure change. Arterioscler Thromb Vasc Biol. 2020; 40:1801–03. <u>https://doi.org/10.1161/ATVBAHA.120.314816</u> PMID:32697680
- 46. Guo T, Yin RX, Lin WX, Wang W, Huang F, Pan SL. Association of the variants and haplotypes in the DOCK7, PCSK9 and GALNT2 genes and the risk of hyperlipidaemia. J Cell Mol Med. 2016; 20:243–65. <u>https://doi.org/10.1111/jcmm.12713</u> PMID:<u>26493351</u>
- 47. Miao L, Yin RX, Pan SL, Yang S, Yang DZ, Lin WX. Weighted gene co-expression network analysis identifies specific modules and hub genes related to hyperlipidemia. Cell Physiol Biochem. 2018; 48:1151–63.

https://doi.org/10.1159/000491982 PMID:<u>30045016</u>

 Yin RX, Wu DF, Miao L, Aung LH, Cao XL, Yan TT, Long XJ, Liu WY, Zhang L, Li M. Several genetic polymorphisms interact with overweight/obesity to influence serum lipid levels. Cardiovasc Diabetol. 2012; 11:123. https://doi.org/10.1186/1475-2840-11-123

PMID:<u>23039238</u>

49. Cao XL, Yin RX, Huang F, Wu JZ, Chen WX. Chromosome 9p21 and ABCA1 genetic variants and their interactions on coronary heart disease and ischemic stroke in a Chinese han population. Int J Mol Sci. 2016; 17:586.

https://doi.org/10.3390/ijms17040586 PMID:27096864

 Gabb GM, Mangoni AA, Anderson CS, Cowley D, Dowden JS, Golledge J, Hankey GJ, Howes FS, Leckie L, Perkovic V, Schlaich M, Zwar NA, Medley TL, Arnolda L. Guideline for the diagnosis and management of hypertension in adults - 2016. Med J Aust. 2016; 205:85–89.

https://doi.org/10.5694/mja16.00526 PMID:27456450

51. Fan Y, Li W, Liu H, Wang L, Zhang S, Li W, Liu H, Leng J, Shen Y, Tuomilehto J, Yu Z, Yang X, Liu M, Hu G. Effects of obesity and a history of gestational diabetes on the risk of postpartum diabetes and hyperglycemia in Chinese women: obesity, GDM and diabetes risk. Diabetes Res Clin Pract. 2019; 156:107828. <u>https://doi.org/10.1016/j.diabres.2019.107828</u> PMID:<u>31472162</u>