Research Perspective

Nuclear lipid droplets: a novel regulator of nuclear homeostasis and ageing

Konstantinos Palikaras¹, Nektarios Tavernarakis^{2,3}

¹Department of Physiology, Medical School, National and Kapodistrian University of Athens, Athens, Greece ²Institute of Molecular Biology and Biotechnology, Foundation for Research and Technology - Hellas, Crete, Greece

³School of Medicine, University of Crete, Heraklion, Crete, Greece

Correspondence to: Konstantinos Palikaras, Nektarios Tavernarakis; email: palikarask@med.uoa.gr, tavernarakis@imbb.forth.grKeywords: aging, ATGL-1, HLH-30/TFEB, lipid droplet, non-linear optical phenomena, nucleusReceived: November 11, 2024Accepted: December 3, 2024Published: December 9, 2024

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ABSTRACT

Aging is a fundamental driver of numerous life-threatening diseases, significantly compromising cellular structures and functions, including the integrity of the nucleus. A consistent feature of aging across diverse species is the progressive accumulation of lipid droplets (nLDs) within the nuclear compartment, which disrupts nuclear architecture and functionality. Notably, aging is accompanied by a marked increase in nLD accumulation at the nuclear envelope. Interventions known to extend lifespan, such as caloric restriction and reduced insulin signaling, significantly reduce both the rate of accumulation and the size of nLDs. The triglyceride lipase ATGL-1, which localizes to the nuclear envelope, plays a critical role in limiting nLD buildup and maintaining nuclear lipid balance, especially in long-lived mutant worms. These findings establish excessive nuclear lipid deposition as a key hallmark of aging, with profound implications for nuclear processes such as chromatin organization, DNA repair, and gene regulation. In addition, ATGL-1 emerges as a promising therapeutic target for preserving nuclear health, extending organismal healthspan, and combating age-related disorders driven by lipid dysregulation.

Ageing is a multifaceted biological process that manifests as gradual decline of cellular functions, and ultimately leads to deterioration of organismal physiology. This complex phenomenon is influenced by a variety of factors, including genetic, environmental, and metabolic effects [1, 2]. A prominent hallmark of ageing is the accumulation of dysfunctional cellular components, which disrupts tissue homeostasis, contributing to agerelated pathologies [3, 4]. This feature includes the abnormal deposition of lipids in non-adipose tissues, known as ectopic fat, which is highly associated with metabolic syndrome and various age-related conditions, including cardiovascular disease, type 2 diabetes, and neurodegenerative disorders, among others, [5, 6].

Lipid droplets (LDs) are primarily recognized as cytoplasmic organelles and have a pivotal role in energy

storage, lipid metabolism, and cellular homeostasis. Traditionally, LDs presence has been confined to the cytoplasm, where they serve as reservoirs of neutral lipids, supporting energy balance and membrane biosynthesis [7, 8]. For a long time, the association between LDs and the nucleus was under-investigated. However, recent studies have unveiled an unexpected aspect of lipid biology demonstrating that LDs can also exist within the nucleus, forming nuclear lipid droplets (nLDs) [9–15]. Although LDs are typically generated from ER membranes and remain closely associated with the outer nuclear membrane, recent studies provide compelling evidence of LDs residing within nucleus [7, 8, 16, 17]. This nuclear localization is surprising and points to previously unappreciated roles for lipid storage and metabolism within the nucleus, introducing an additional layer of complexity to our understanding of cellular processes, particularly in the context of ageing.

Over the last decade, several methods have been developed to monitor LDs number, morphology, and distribution within cells and across tissues. However, many of these methods face limitations that prevent the real-time, in vivo tracking of LDs throughout the lifespan of an organism [18–21]. In previous studies, we have established the use of non-linear imaging modalities to assess LDs formation and deposition during ageing in vivo [21–23]. Using this label free and non-invasive methodology, we revealed that LDs progressively accumulate in the nuclear envelope as C. elegans ages [24]. Recent findings suggest that the nuclear envelope is a metabolically active site where lipids are synthesized and stored, adding a new dimension to our understanding of lipid metabolism and its relationship with nuclear integrity [12, 15]. Notably, the build-up of nuclear lipid droplets (nLDs) is closely associated with age-related alterations in the nuclear lamina protein LMN-1, the C. elegans homolog of mammalian lamin A/C. These findings underscore an intricate relationship between nuclear envelope components and LDs functionality. These interactions could be crucial for the maintenance of nuclear integrity, as changes in nuclear morphology are wellestablished hallmarks of ageing across species [25, 26]. Further supporting this notion, studies in human cells have shown that mutations in the LMNA gene not only lead to abnormal nuclear morphology but also result in altered expression of metabolism-related genes, features similarly observed in cells derived from metabolic syndrome patients, which also exhibit impaired nuclear distribution of lamin A/C [27, 28]. The accumulation of nLDs, therefore, is not simply a passive consequence of ageing but might actively contribute to nuclear morphology disruption and impaired cellular function. Notably, disruptions in nuclear lipid homeostasis have been linked to metabolic disorders such as fatty liver disease and obesity, suggesting that similar mechanisms could be conserved across species and relevant not only to ageing but also to disease pathology [5, 8, 25]. However, further investigation is needed to assess nLDs distribution within nuclear compartments particularly in cells derived from progeria and metabolic syndrome patients, where such mechanisms might have a critical role.

While the physical association of LDs with the nuclear compartments is highly recognized, our understanding of their functional roles in nuclear homeostasis remains elusive. Genetic studies conducted in mammalian cells, flies and nematodes have been shown that LDs contribute to nuclear integrity by storing histones, binding transcription factors, directly interacting with

other proteins that regulate the transport between the cytoplasm and nucleoplasm [7, 8, 16]. Hence, the uncontrolled accumulation of LDs in the nuclear envelope might be detrimental for several nuclear processes, including nuclear transport, DNA repair, chromatin remodelling and ultimately, gene expression. A recent study in C. elegans revealed that nLDs are coated by LMN-1 and/or heterochromatin, indicating that the accumulation of nLDs could play a role in the disposal of peripheral heterochromatin [14]. These results indicate that nLDs might influence chromatin organization, given their formation at the nuclear envelope and their ability to penetrate the nuclear lamina, entering the nucleoplasm. This is particularly relevant since the loss of heterochromatin and the derepression of silenced genes are considered major contributors to premature ageing. Chromatin loosening is a common feature of ageing, leading to deregulated gene expression and compromised genome integrity [29, 30]. Lamin-Associated Domains (LADs), which are closely linked to heterochromatin, could be destabilized by the age-dependent accrual of nLDs at the nuclear lamina (Figure 1). In turn, these events could contribute to heterochromatin loss during ageing, suggesting that nLD accumulation may play a significant role in age-related chromatin remodelling. Moreover, the excessive nLD accumulation could interfere with chromosome territories, leading to nuclear damage and cellular dysfunction. These concepts are further supported by the well-documented loss of intestinal nuclei during C. elegans ageing, which might, in fact, be driven by increased nLD accumulation [14, 31, 32]. However, further studies are needed to validate these concepts and uncover the precise mechanisms involved.

One of the most compelling aspects of our study is the effect of longevity-promoting interventions on nLD accumulation. Dietary restriction and reduced insulin signalling were found to significantly reduce both the size and number of nLDs in aged nematodes. These findings underscore the role of metabolic regulation in extending lifespan, hinting at the importance of nLDs homeostasis in maintaining cellular integrity during longevity-promoting interventions ageing. Both correlated with lower levels of nLD accumulation, preservation of nuclear morphology, and sustained expression of the nuclear structural protein LMN-1 [24, 33, 34]. Furthermore, our study uncovered the transcription factor HLH-30, the nematode homolog of the mammalian transcription factor EB (TFEB), to play an essential role in preserving nuclear lipid balance. Intriguingly, HLH-30 operates independently of autophagy, establishing a distinct pathway for nLDs regulation. Both HLH-30 and its mammalian counterpart TFEB are activated and translocate to the

nucleus in response to low insulin signalling and caloric restriction [35, 36]. HLH-30 subsequently regulates the expression of genes involved in autophagy, lysosomal function, and lipid metabolism, including the gene for the lipase ATGL-1 [24, 35, 36]. Our findings also indicate that ATGL-1 is essential for regulating nLD size and abundance in long-lived nematodes, as its overexpression alone is sufficient to maintain nLDs. Notably, ATGL-1 is localized in both the cytoplasm and nuclear envelope, suggesting it plays a dual role in moderating lipid accumulation across cellular compartments [24]. The mammalian homolog of ATGL-1, PNPLA2, exhibits similar dual localization, being found in both cytosolic LDs and the nucleoplasm in a range of human cell lines, including A-549, U-251MG, HEK293, U2OS, and SiHA cells (as documented in the Human Protein Atlas, http://www.proteinatlas.org/). The regulatory relationship between HLH-30 and ATGL-1 provides a mechanistic between longevity-promoting link pathways and nLDs metabolism, offering insights into how these interventions maintain nuclear integrity through lipid homeostasis. Taken together, these findings suggest that metabolic pathways supporting longevity may promote cellular health by directly influencing nLD dynamics. This influence, in turn, likely plays a critical role in sustaining nuclear integrity, positioning nLD regulation as a promising target for interventions aimed at promoting healthy ageing.

The identification of age-dependent nLDs accumulation as a hallmark of ageing marks a significant advancement in our understanding of the interplay between lipid metabolism and nuclear function. Accumulation of nLDs disrupts nuclear integrity, potentially impairing essential nuclear processes such as chromatin organization, DNA repair, and gene expression. Beyond advancing our knowledge of the ageing process, this discovery suggests promising targets for interventions aimed at extending healthspan and preventing age-related diseases. As research into nLDs regulation progresses, it is conceivable that novel therapeutic strategies will emerge, leveraging lipid metabolism to combat the effects of ageing and enhance human health. Given the established links between lipid deregulation, metabolic disorders, and ageing, these findings hold considerable promise for developing therapies aimed at safeguarding nuclear function and mitigating the adverse effects of ageing on cellular and organismal physiology.



Figure 1. Age-dependent nLDs accumulation affects nuclear homeostasis. Over time, lipid droplets (LDs) accumulate progressively in the cytoplasm and within the nuclear lamina, closely associated with age-related changes in the nuclear lamina protein LMN-1 (the nematode homolog of lamin A/C), which supports nuclear morphology. This nLD accumulation may disrupt Lamin-Associated Domains (LADs), chromatin regions anchored to the nuclear periphery, leading to chromatin remodelling, heterochromatin destabilization, and compromised nuclear integrity, which are well-established hallmarks of ageing. Longevity-promoting interventions activate the lipase ATGL-1, regulated by HLH-30/TFEB, linking lipid metabolism to pathways that promote cellular longevity. These changes in nLDs could impact key nuclear processes, including gene expression and DNA repair, thereby contributing to cellular ageing. Created in BioRender (Palikaras, K., 2024, <u>https://BioRender.com/t49f941</u>).

CONFLICTS OF INTEREST

The authors declare no conflicts of interest related to this study.

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REFERENCES

- Melzer D, Pilling LC, Ferrucci L. The genetics of human ageing. Nat Rev Genet. 2020; 21:88–101. <u>https://doi.org/10.1038/s41576-019-0183-6</u> PMID:<u>31690828</u>
- Guo J, Huang X, Dou L, Yan M, Shen T, Tang W, Li J. Aging and aging-related diseases: from molecular mechanisms to interventions and treatments. Signal Transduct Target Ther. 2022; 7:391. <u>https://doi.org/10.1038/s41392-022-01251-0</u> PMID:<u>36522308</u>
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. Hallmarks of aging: An expanding universe. Cell. 2023; 186:243–78. <u>https://doi.org/10.1016/j.cell.2022.11.001</u> PMID:<u>36599349</u>
- Aman Y, Schmauck-Medina T, Hansen M, Morimoto RI, Simon AK, Bjedov I, Palikaras K, Simonsen A, Johansen T, Tavernarakis N, Rubinsztein DC, Partridge L, Kroemer G, et al. Autophagy in healthy aging and disease. Nat Aging. 2021; 1:634–50. <u>https://doi.org/10.1038/s43587-021-00098-4</u> PMID:<u>34901876</u>
- Ou MY, Zhang H, Tan PC, Zhou SB, Li QF. Adipose tissue aging: mechanisms and therapeutic implications. Cell Death Dis. 2022; 13:300. <u>https://doi.org/10.1038/s41419-022-04752-6</u> PMID:<u>35379822</u>
- Neeland IJ, Ross R, Després JP, Matsuzawa Y, Yamashita S, Shai I, Seidell J, Magni P, Santos RD, Arsenault B, Cuevas A, Hu FB, Griffin B, et al, and International Atherosclerosis Society, and International Chair on Cardiometabolic Risk Working Group on Visceral Obesity. Visceral and ectopic fat, atherosclerosis, and cardiometabolic disease: a position statement. Lancet Diabetes Endocrinol. 2019; 7:715–25.

https://doi.org/10.1016/S2213-8587(19)30084-1 PMID:<u>31301983</u>

- Mathiowetz AJ, Olzmann JA. Lipid droplets and cellular lipid flux. Nat Cell Biol. 2024; 26:331–45. <u>https://doi.org/10.1038/s41556-024-01364-4</u> PMID:38454048
- Zadoorian A, Du X, Yang H. Lipid droplet biogenesis and functions in health and disease. Nat Rev Endocrinol. 2023; 19:443–59. <u>https://doi.org/10.1038/s41574-023-00845-0</u> PMID:37221402
- Morales A, Greenberg M, Nardi F, Gil V, Hayward SW, Crawford SE, Franco OE. Loss of ephrin B2 receptor (EPHB2) sets lipid rheostat by regulating proteins DGAT1 and ATGL inducing lipid droplet storage in prostate cancer cells. Lab Invest. 2021; 101:921–34. <u>https://doi.org/10.1038/s41374-021-00583-9</u> PMID:<u>33824421</u>
- Sołtysik K, Ohsaki Y, Tatematsu T, Cheng J, Fujimoto T. Nuclear lipid droplets derive from a lipoprotein precursor and regulate phosphatidylcholine synthesis. Nat Commun. 2019; 10:473. <u>https://doi.org/10.1038/s41467-019-08411-x</u> PMID:<u>30692541</u>
- Sołtysik K, Ohsaki Y, Tatematsu T, Cheng J, Maeda A, Morita SY, Fujimoto T. Nuclear lipid droplets form in the inner nuclear membrane in a seipin-independent manner. J Cell Biol. 2021; 220:e202005026. <u>https://doi.org/10.1083/jcb.202005026</u> PMID:33315072
- Romanauska A, Köhler A. Reprogrammed lipid metabolism protects inner nuclear membrane against unsaturated fat. Dev Cell. 2021; 56:2562–78.e3. <u>https://doi.org/10.1016/j.devcel.2021.07.018</u> PMID:34407429
- Taurino M, Costantini S, De Domenico S, Stefanelli F, Ruano G, Delgadillo MO, Sánchez-Serrano JJ, Sanmartín M, Santino A, Rojo E. SEIPIN Proteins Mediate Lipid Droplet Biogenesis to Promote Pollen Transmission and Reduce Seed Dormancy. Plant Physiol. 2018; 176:1531–46. <u>https://doi.org/10.1104/pp.17.01430</u> PMID:29203558
- 14. Mosquera JV, Bacher MC, Priess JR. Nuclear lipid droplets and nuclear damage in Caenorhabditis elegans. PLoS Genet. 2021; 17:e1009602. <u>https://doi.org/10.1371/journal.pgen.1009602</u> PMID:<u>34133414</u>
- Romanauska A, Köhler A. The Inner Nuclear Membrane Is a Metabolically Active Territory that Generates Nuclear Lipid Droplets. Cell. 2018; 174:700–15.e18. <u>https://doi.org/10.1016/j.cell.2018.05.047</u> PMID:29937227

- Barbosa AD, Siniossoglou S. New kid on the block: lipid droplets in the nucleus. FEBS J. 2020; 287:4838–43. <u>https://doi.org/10.1111/febs.15307</u> PMID:<u>32243071</u>
- Fujimoto T. Nuclear lipid droplets how are they different from their cytoplasmic siblings? J Cell Sci. 2022; 135:jcs259253. <u>https://doi.org/10.1242/jcs.259253</u> PMID:<u>35217856</u>
- Fam TK, Klymchenko AS, Collot M. Recent Advances in Fluorescent Probes for Lipid Droplets. Materials (Basel). 2018; 11:1768. <u>https://doi.org/10.3390/ma11091768</u> PMID:<u>30231571</u>
- Gandhi SA, Parveen S, Alduhailan M, Tripathi R, Junedi N, Saqallah M, Sanders MA, Hoffmann PM, Truex K, Granneman JG, Kelly CV. Methods for making and observing model lipid droplets. Cell Rep Methods. 2024; 4:100774. <u>https://doi.org/10.1016/j.crmeth.2024.100774</u> PMID:<u>38749444</u>
- 20. Kim H, Oh S, Lee S, Lee KS, Park Y. Recent advances in label-free imaging and quantification techniques for the study of lipid droplets in cells. Curr Opin Cell Biol. 2024; 87:102342. <u>https://doi.org/10.1016/j.ceb.2024.102342</u> PMID:<u>38428224</u>
- Débarre D, Supatto W, Pena AM, Fabre A, Tordjmann T, Combettes L, Schanne-Klein MC, Beaurepaire E. Imaging lipid bodies in cells and tissues using thirdharmonic generation microscopy. Nat Methods. 2006; 3:47–53. <u>https://doi.org/10.1038/nmeth813</u> PMID:<u>16369553</u>
- Tserevelakis GJ, Megalou EV, Filippidis G, Petanidou B, Fotakis C, Tavernarakis N. Label-free imaging of lipid depositions in C. elegans using third-harmonic generation microscopy. PLoS One. 2014; 9:e84431. <u>https://doi.org/10.1371/journal.pone.0084431</u> PMID:<u>24392137</u>
- Palikaras K, Mari M, Petanidou B, Pasparaki A, Filippidis G, Tavernarakis N. Ectopic fat deposition contributes to age-associated pathology in Caenorhabditis elegans. J Lipid Res. 2017; 58:72–80. <u>https://doi.org/10.1194/jlr.M069385</u> PMID:<u>27884963</u>
- 24. Palikaras K, Mari M, Ploumi C, Princz A, Filippidis G, Tavernarakis N. Age-dependent nuclear lipid droplet deposition is a cellular hallmark of aging in Caenorhabditis elegans. Aging Cell. 2023; 22:e13788. <u>https://doi.org/10.1111/acel.13788</u> PMID:<u>36718841</u>

- Pathak RU, Soujanya M, Mishra RK. Deterioration of nuclear morphology and architecture: A hallmark of senescence and aging. Ageing Res Rev. 2021; 67:101264. <u>https://doi.org/10.1016/j.arr.2021.101264</u>
 PMID:33540043
- Heckenbach I, Mkrtchyan GV, Ezra MB, Bakula D, Madsen JS, Nielsen MH, Oró D, Osborne B, Covarrubias AJ, Idda ML, Gorospe M, Mortensen L, Verdin E, et al. Nuclear morphology is a deep learning biomarker of cellular senescence. Nat Aging. 2022; 2:742–55.

https://doi.org/10.1038/s43587-022-00263-3 PMID:<u>37118134</u>

- Desgrouas C, Varlet AA, Dutour A, Galant D, Merono F, Bonello-Palot N, Bourgeois P, Lasbleiz A, Petitjean C, Ancel P, Levy N, Badens C, Gaborit B. Unraveling *LMNA* Mutations in Metabolic Syndrome: Cellular Phenotype and Clinical Pitfalls. Cells. 2020; 9:310. <u>https://doi.org/10.3390/cells9020310</u> PMID:<u>32012908</u>
- Dutour A, Roll P, Gaborit B, Courrier S, Alessi MC, Tregouet DA, Angelis F, Robaglia-Schlupp A, Lesavre N, Cau P, Lévy N, Badens C, Morange PE. High prevalence of laminopathies among patients with metabolic syndrome. Hum Mol Genet. 2011; 20:3779–86. <u>https://doi.org/10.1093/hmg/ddr294</u> PMID:<u>21724554</u>
- 29. Lee JH, Kim EW, Croteau DL, Bohr VA. Heterochromatin: an epigenetic point of view in aging. Exp Mol Med. 2020; 52:1466–74. <u>https://doi.org/10.1038/s12276-020-00497-4</u> PMID:<u>32887933</u>
- 30. Emerson FJ, Lee SS. Chromatin: the old and young of it. Front Mol Biosci. 2023; 10:1270285. <u>https://doi.org/10.3389/fmolb.2023.1270285</u> PMID:<u>37877123</u>
- Poteryaev D, Squirrell JM, Campbell JM, White JG, Spang A. Involvement of the actin cytoskeleton and homotypic membrane fusion in ER dynamics in Caenorhabditis elegans. Mol Biol Cell. 2005; 16:2139–53. <u>https://doi.org/10.1091/mbc.e04-08-0726</u> PMID:<u>15716356</u>
- Gems D, Riddle DL. Defining wild-type life span in Caenorhabditis elegans. J Gerontol A Biol Sci Med Sci. 2000; 55:B215–9. <u>https://doi.org/10.1093/gerona/55.5.b215</u> PMID:10819307
- Haithcock E, Dayani Y, Neufeld E, Zahand AJ, Feinstein N, Mattout A, Gruenbaum Y, Liu J. Age-related changes of nuclear architecture in Caenorhabditis elegans. Proc Natl Acad Sci U S A. 2005; 102:16690–5.

https://doi.org/10.1073/pnas.0506955102 PMID:<u>16269543</u>

- 34. Charar C, Metsuyanim-Cohen S, Bar DZ. Lamin regulates the dietary restriction response via the mTOR pathway in Caenorhabditis elegans. J Cell Sci. 2021; 134:jcs258428. <u>https://doi.org/10.1242/jcs.258428</u> PMID:<u>34383046</u>
- 35. Lapierre LR, De Magalhaes Filho CD, McQuary PR, Chu CC, Visvikis O, Chang JT, Gelino S, Ong B, Davis AE, Irazoqui JE, Dillin A, Hansen M. The TFEB orthologue HLH-30 regulates autophagy and modulates longevity in Caenorhabditis elegans. Nat Commun. 2013; 4:2267. <u>https://doi.org/10.1038/ncomms3267</u> PMID:23925298
- Lin XX, Sen I, Janssens GE, Zhou X, Fonslow BR, Edgar D, Stroustrup N, Swoboda P, Yates JR 3rd, Ruvkun G, Riedel CG. DAF-16/FOXO and HLH-30/TFEB function as combinatorial transcription factors to promote stress resistance and longevity. Nat Commun. 2018; 9:4400.

https://doi.org/10.1038/s41467-018-06624-0 PMID:<u>30353013</u>