Research Paper

Developing a quantitative estimate of muscle age acceleration by a novel phenotypic clock: cross-sectional study in healthy, middle-aged and older adults

Lucia Ventura^{1,2,*}, Antonella Cano^{1,*}, Marco Morrone^{1,*}, Gianluca Martinez¹, Anna Boi¹, Maria Grazia Catte¹, Beniamina Mercante¹, Nicola Loi¹, Oksana Yurchyshyn³, Giovanni Fiorito⁴, Giuliana Solinas¹, Sara Cruciani¹, Donatella Coradduzza¹, Margherita Maioli¹, Angelo Zinellu¹, Ciriaco Carru¹, Franca Deriu^{1,5}, Andrea Manca¹

Correspondence to: Franca Deriu; email: deriuf@uniss.it

Keywords: ageing, biological age, EWGSOP, frailty, sarcopenia

Received: December 10, 2024 **Accepted:** June 2, 2025 **Published:** June 9, 2025

Copyright: © 2025 Ventura et al. This is an open access article distributed under the terms of the <u>Creative Commons</u> <u>Attribution License</u> (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Sarcopenia is a progressive disease characterized by reductions in muscle mass strength and physical performance. Among the initiatives launched to increase awareness, the European Working Group on Sarcopenia in Older People (EWGSOP) is considered the most influential. This cross-sectional study was planned to develop, in healthy middle-aged and older adults, a novel predictor of sarcopenia based on the motor-functional and anthropometric tests derived from EWGSOP2, which were the primary outcome measures.

Participants were tested for body composition, physical performance, blood biomarkers, and risk scores for major healthy issues. Muscle Age Acceleration (MAA) was modelled with Elastic Net regression to extract EWGSOP test mostly contributing to the musculoskeletal ageing trajectory.

Two-hundred-fifteen participants were tested (118 women, 97 men; mean age; 66.0 ± 7.3 years). Muscle Age was correlated with chronological age (r = 0.645; p < 0.001). Parsimonious modelling extracted TUG ($\beta = 2.93$; 2.48 - to -3.51), ASMM ($\beta = -2.23$; -2.99 to -1.67) and Handgrip ($\beta = -1.12$; -1.70 to -0.42) for men, and TUG ($\beta = 2.69$; 1.96 to 4.19), Handgrip ($\beta = -1.27$; -1.56 to -0.98), and Six-MWT ($\beta = -1.15$; -1.71 to -0.53) for women. According to MAA, three trajectories were identified: accelerated agers displayed higher risk for sarcopenia (19%), as compared to normal (9%; p < 0.0001) and decelerated (2%; p < 0.0001), paralleled by significant subclinical alterations of haemato-chemical markers in accelerated agers.

MAA could validly identify accelerated agers with higher risk of sarcopenia, whereas PhenoAge detected subclinical haematochemical alterations. Longitudinal studies are needed to appraise the validity of this newly introduced predictor of sarcopenia and verify if accelerated agers are at higher risk for developing sarcopenia.

¹Department of Biomedical Sciences, University of Sassari, Sassari, Italy

²Azienda Ospedaliero-Universitaria di Sassari, Sassari, Italy

³Ternopil National Medical University, Ternopil, Ukraine

⁴IRCCS Ospedale Pediatrico Giannina Gaslini, Genova, Italy

⁵Endocrinology Unit, AOUSS, Sassari, Italy

^{*}Equal contribution

INTRODUCTION

Sarcopenia is a progressive, generalised skeletal muscle disease linked to negative health changes that accumulate across the lifespan [1]. It is common among adults of older age, but occurs also earlier in life, leading to increased likelihood of falls, fractures, disability and mortality. Literally meaning "poverty of flesh" [2], sarcopenia is recognized as a broader disorder where reductions in muscle strength, muscle quantity/quality and physical performance are detected. Multifactorial pathogenesis underpins this condition. From a pathophysiological standpoint, endocrine and metabolic abnormalities interact with the low-grade chronic inflammation (i.e., "inflammageing"), that is observed in advanced agers [3, 4], leading to a reduction of protein-synthesis and regeneration, and a parallel pattern of muscle wasting due to increased apoptosis and protein-lysis [5].

While healthcare professionals are now more aware of this condition and can recognise its negative impact on health, such knowledge has not fully translated into clinical practice. Consequently, sarcopenia is still underdiagnosed and undertreated, leading to a considerable financial burden on healthcare systems for the inherent condition of frailty, increased risk for hospitalisation and cost of care during hospitalisation

for sarcopenic compared to non-sarcopenic patients [6].

Among the number of initiatives launched to advance knowledge on sarcopenia and prompt preventive/ therapeutic approaches, the European Working Group on Sarcopenia in Older People (EWGSOP) has emerged as the most influential in raising awareness and moving the field forward. EWGSOP consensus firstly introduced a broad clinical definition for sarcopenia not limited to muscle loss [7]. This was eventually developed more recently (EWGSOP2) [1] to move muscle weakness and reduced performance to the forefront as primary indicators of sarcopenia. EWGSOP2's recommendations also developed an algorithm for case-finding, diagnosis, and severity determination for a consistent identification of people with sarcopenia or its risk, and simple, specific cut-off points for measures that identify and characterise sarcopenia. These important implementations are summarised in Figure 1.

While EWSGOP2's algorithm for sarcopenia screening has undeniably increased awareness of this condition, its categorical nature does not allow to automatically obtain an outcome that quantifies the degree of sarcopenia. Conversely, a scalar, quantitative measure would help identify individuals who do not qualify as

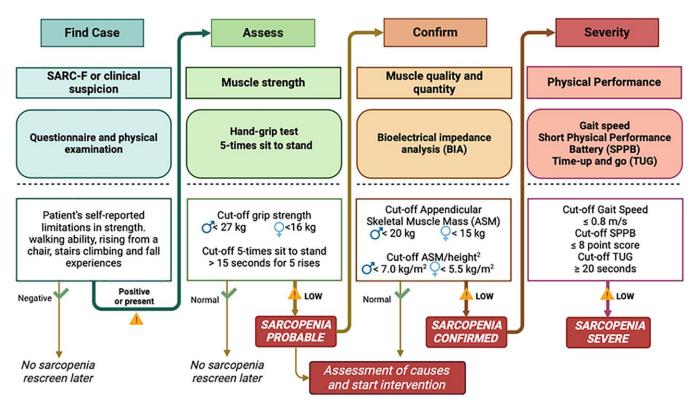


Figure 1. EWGSOP2 algorithm for case-finding, assessment, confirmation and level of severity for sarcopenia (modified from A.J. Cruz-Jentoft et al. *Age and ageing* 2019; 48:16–31) [1].

Table 1. Main demographic, anthropometric and psycho-cognitive characteristics of the participants.

	Total sample (<i>n</i> = 215)	Women (n = 118)	Men (n = 97)	Men vs. Women One-way ANOVA
Chronological aga (vaara)	66.01 ± 7.3	65.7 ± 6.7	66.4 ± 7.8	p = 0.487
Chronological age (years)	(65.1-66.9)	(64.5-66.9)	(64.8-67.9)	p = 0.467
Waight (Ira)	66.6 ± 12.2	59.1 ± 7.5	76.2 ± 10.0	m < 0.0001
Weight (kg)	(65.0-68.2)	(57.8–60.4)	(74.3 - 78.2)	p < 0.0001
Height (cm)	161.1 ± 10.6	154.1 ± 7.3	170.0 ± 6.9	m < 0.0001
Height (cm)	(159.7-162.5)	(152.8-155.4)	(168.7 - 171.4)	p < 0.0001
Dody Mass Inday (lyg/m²)	25.4 ± 2.8	24.7 ± 2.7	26.3 ± 2.6	m < 0.0001
Body Mass Index (kg/m²)	(25.0-25.8)	(24.3-25.2)	(25.8-26.8)	p < 0.0001
ADL (recus)	6.0(0.0)	6.0 (0.04)	6.0 (6.04)	0.721
ADL (score)	(6.0-8.0)	(6.0-8.0)	(6.0-8.0)	p = 0.731
LADI (saara)	8.0 (0.0)	8.0 (0.0)	8.0 (0.0)	n = 0.070
I-ADL (score)	(8.0-8.0)	(8.0-8.0)	(8.0-8.0)	p = 0.070
MaCA (adjusted seems)	24.98 (0.24)	24.62 (0.33)	24.98 (0.37)	n = 0.504
MoCA (adjusted score)	(24.6-25.6)	(24.0-25.5)	(24.5-26.0)	p = 0.594
Charleson Comarkidita Indon (a)	2.78 (1.34)	2.80 (1.35)	2.75 (1.34)	0 927
Charlsson Comorbidity Index (n)	(2.56-3.01)	(2.50-3.10)	(2.40-3.11)	p = 0.837
IDAO (1 larry 2 madamata, 2 kink)	2 (0.06)	2 (0.04)	2 (0.09)	0.267
IPAQ (1-low; 2-moderate; 3-high)	(1.8-2.2)	(1.7-2.1)	(1.9–2.3)	p = 0.367

All data are presented as mean ± standard deviation and confidence interval of the mean, except for ADL, I-ADL, MoCA scores and IPAQ, which are expressed as median (standard error) and (confidence interval of the median). Abbreviations: kg: kilograms; cm: centimeters; m²: square meters; ADL: Activities of Daily Living; I-ADL: Instrumental ADL; MoCA: Montreal Cognitive Assessment.

sarcopenic despite displaying subclinical alterations that potentially deserve preventive strategies. Such marker would also allow to determine whether interventions aimed at mitigating sarcopenia are truly effective.

This sex-based study, which was conducted in a cohort of healthy, middle-aged and older adults, aimed at (1) developing a predictor of sarcopenia based on the performance of various motor-functional and anthropometric tests, named Muscle Age Acceleration (MAA), as derived from the EWGSOP2 consensus [1], (2) identifying, by means of MAA, subclinical musculoskeletal impairments separately in women and men, and (3) test MAA association with health-related, biological features, including a biological ageing clock built on a set of blood-measured biomarkers strongly predictive of longevity, known as the Phenotypic Age (PhenoAge) [8].

Justification for this study resides in that EWGSOP2-based MAA would allow to capture even subtle, subclinical musculoskeletal dysfunctions also in apparently non-sarcopenic individuals transitioning from middle-age to senescence, allowing predictive selection of individuals at risk for developing sarcopenia.

RESULTS

Two-hundred-fifteen healthy participants (118 women, 97 men) volunteered for this study. Main demographic, anthropometric, clinical and lifestyle characteristics are summarised in Table 1. Participants were 66.0 ± 7.3 years old (women: 65.7 ± 6.7 ; men: 66.4 ± 7.8). Women and men were of comparable age (p = 0.49) while, as expected, men were significantly heavier and taller (p < 0.0001). The two sexes did not differ in terms of median physical activity levels, as estimated by IPAQ (women: 3; men: 3) and comorbidities, as assessed by the median Charlsson Comorbidity Index score (women: 3 ± 0.15 ; men: 3 ± 0.18).

Muscle age and muscle age acceleration (MAA)

Derived from the penalised elastic net model, Muscle Age was significantly correlated with chronological age (r = 0.645; p < 0.001). Figure 2 displays the sex-based contributors based on the results of the motor-functional tests recommended by EWGSOP2. The contributions of each test in accelerating (red bars) or decelerating (green bars) the ageing trajectory are reported as coefficients indicating the standardised weight of each test in the construction of the biological/Muscle Age.

Coefficients equal to zero indicate no contribution to the ageing measure. Being standardised, the weights can be read as the increase in Muscle Age for each increase by one standard deviation of the corresponding test result. Positive coefficients/weights indicate motor tests whose results are higher in an individual with higher Muscle Age and vice versa. Parsimonious modelling to retain only those features mostly contributing to the estimation of MAA extracted TUG (β = 2.93; 2.48- to -3.51), ASMM (β = -2.23; -2.99 to -1.67) and Handgrip (β = -1.12; -1.70 to -0.42) for men, and TUG (β = 2.69;

1.96 to 4.19), Handgrip (β = -1.27; -1.56 to -0.98), and Six-MWT (β = -1.15; -1.71 to -0.53) for women. RMSE and R² of these parsimonious models were RMSE = 5.56 ± 0.90, R² = 0.42 ± 0.17 for men, and RMSE = 5.43±0.65, R² = 0.35 ± 0.16 for women.

Muscle Age was then linearly regressed on chronological age and the residuals retained as age accelerations in years, i.e., MAA. This allowed to account for the inherent age-related decline of motor-functional performance that is positively correlated with age.

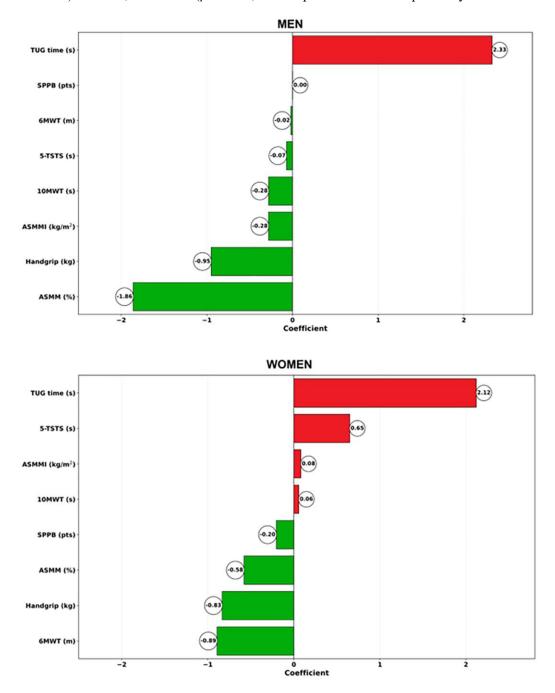


Figure 2. Contributors from the penalized elastic net regression model run with coefficients indicating the standardized weight in women (upper panel) and men (lower panel).

Classifying participants according to MAA

Based on the above results, participants were classified into three categories of agers according to the percentile-based distribution of the data: 'decelerated' (up to 25th percentile: n = 53; 23W:30M) for individuals with MAA scores lower than -3.8 years for women and -3.3 years for men; 'normal' (26th to 75th percentile: n = 109; 69W:40M) for individuals with MAA between -3.8 and +2.5 years for women and -3.3 to +4.2 for men; 'accelerated' (from 76th percentile: n = 53; 26W:27M) for individuals with MAA higher than 2.5 years for women and 4.2 years for men.

Comparing MAA trajectories

Comparisons among the three MAA categories carried out by general linear model ANOVA on the whole cohort are detailed in Table 2. MAA could identify participants at higher risk for sarcopenia, as assessed by Ishii's formula [9], with accelerated agers displaying 19% probability of sarcopenia, which proved significantly higher than normal (9%; p < 0.0001) and decelerated agers (2%; p < 0.0001). Sex-based comparisons showed higher probability of sarcopenia both in accelerated (14%) compared to normal (2%) and decelerated male agers (all p-values ≤ 0.03). The same applied to women (accelerated: 25%; normal: 9%; decelerated: 4%; all p-values ≤ 0.001) (Figure 3).

No significant main effects of MAA, sex or interaction term (MAA_status*sex) emerged for the scores of MOCA, ADL, IADL, and health status (total and subscores). However, MOCA proved significantly correlated with MAA in the whole group (r = -0.14; p = 0.04; n = 212) and, more specifically, in the accelerated subgroup (r = -0.39; p = 0.03).

Table 3 summarises the main effects of MAA, sex and MAA*sex interaction for the comparisons run on the haematochemical samples. Bonferroni-corrected pairwise comparisons revealed a significant difference in WBC count only for men when comparing decelerated $(5.62/\mu L)$ vs. accelerated agers $(6.63/\mu L; +18\%; p =$ 0.02). Regardless of sex, HGB mass was significantly lower in accelerated (13.5 g/dL) compared to decelerated (14.2 g/dL; p < 0.0001) and normal agers (14.0 g/dL; p < 0.0001). HCT was significantly lower in accelerated agers (42.7%) compared to normal agers (44.4%; p = 0.04). Only among women MCV was found significantly smaller (p = 0.02) for accelerated (82.1 μm³) compared to decelerated agers (90.3 μm³). The same applied to MCH for which women with accelerated ageing showed lower MCH (25.9 µm³) than decelerated ones (29.1 pg; p = 0.02). %RDW was found significantly higher in accelerated women than decelerated ones (14.32% and 13.50%, respectively; p = 0.04).

Monocyte count was significantly higher in accelerated (3.87/µL) compared to decelerated agers (3.33/µL; p=0.02), with no differences by sex. Conversely, only among men eosinophil count was significantly higher in accelerated (0.26/µL) compared to normal (0.18/µL, p=0.03) and decelerated agers (0.17/µL; p=0.02). Likewise, neutrophil count was significantly lower in decelerated men (3.17/µL) compared to normal (3.79/µL; p=0.04) but not accelerated agers. The same applied to creatinine, which was found significantly higher in decelerated men (1.03 mg/dL) compared to accelerated (0.91 mg/dL; p=0.03). Regardless of sex, plasma PSH was significantly higher in decelerated (5.09 µmol/g) compared to accelerated agers (4.54 µmol/g; p=0.03).

No differences by MAA emerged for RBC, mean corpuscular haemoglobin concentration, haemoglobin distribution width, platelets count, mean platelet volume, neutrophil %, lymphocyte %, monocyte %, eosinophil %, basophil %, large unstained cells %, lymphocyte count, basophil count, large unstained cells count, glycemia, plasma concentration of urea, albumin, total proteins, alkaline phosphatase, serum iron, CRP, IL-6, IGF-1, leptine, malondialdehyde, and paraoxonase, and homocysteine (Table 3). Figure 4 shows a visual comparison of decelerated, normal, and accelerated aging trajectories for three exemplificative haematochemical parameters proved that significantly differ by MAA category.

Biological age estimation

The PhenoAge calculator by Levine (2018) [8] allowed to estimate participants' biological age and the related age acceleration. While Phenotypic and Muscle ages proved significantly correlated with one another (r=0.59; p<0.001) as well as with chronological age (r=0.90 and 0.69, respectively; p<0.001), no correlation was found between the derived measures of age acceleration, MAA and biological age acceleration (r=0.01; p=0.87).

Health risks

Of the two markers of age acceleration, only PhenoAA proved significantly correlated, in both women and men, with all cardiovascular risk scores and mortality risk, but not with comorbidities (Table 4). Conversely, MAA was associated with mortality risk only in men.

DISCUSSION

This study was conducted in healthy, middle-aged and older adults aged 50-90 years, to (1) portray their

Table 2. Descriptive and statistic results by European Working Group of Sarcopenia in Older People (EWGSOP2) assessments according to Muscle Age Acceleration (MAA) status and sex (*N* = 215; 53 decelerated, 109 normal, 53 accelerated agers; 118 women, 97 men).

		MAA status			Statistics			
		Decelerated	Normal	Accelerated	Main effect Pairwise compa			
Ishii	Whole sample	0.02 (0.02)	0.05 (0.02)	0.19 (0.03)	MAA status $F = 13.79 p < 0.0001$	Decel <accel <math="">p < 0.0001 Normal<accel <math="">p < 0.0001</accel></accel>		
sarcopenia probability	Women	0.04 (0.04)	0.09 (0.02)	0.25 (0.03)	Sex $F = 6.30 p = 0.013$	Decel <accel <math="">p < 0.0001 Normal<accel <math="">p < 0.001</accel></accel>		
	Men	0.01 (0.03)	0.02 (0.03)	0.14 (0.04)	MAA*Sex F = $0.69 p = 0.503$	Decel <accel <math="">p = 0.034 Normal<accel <math="">p = 0.030</accel></accel>		
Handgrip	Whole sample	31.66 (0.70)	28.57 (0.52)	25.39 (0.70)	MAA status $F = 19.90 p < 0.0001$	Decel>Accel $p < 0.0001$ Decel>Normal $p = 0.001$ Normal>Accel $p = 0.001$		
strength (kg)	Women	23.47 (1.01)	20.68 (0.61)	18.66 (0.96)	Sex $F = 413.35 p < 0.0001$	Decel>Accel $p = 0.002$		
(- 8)	Men	39.85 (0.97)	36.46 (0.84)	32.11 (1.04)	MAA*Sex $F = 1.27 p = 0.283$	Decel>Normal $p = 0.027$ Decel>Accel $p < 0.0001$ Normal>Accel $p = 0.004$		
	Whole sample	7.07 (0.22)	7.36 (0.16)	8.68 (0.22)	MAA status $F = 16.05 p < 0.0001$	Decel <accel <math="">p < 0.0001 Normal<accel <math="">p < 0.0001</accel></accel>		
Five-TSTS (s)	Women	6.71 (0.32)	7.27 (0.19)	8.92 (0.30)	Sex $F = 0.36 p = 0.549$	Decel <accel <math="">p < 0.0001 Normal<accel <math="">p < 0.0001</accel></accel>		
	Men	7.42 (0.30)	7.45 (0.26)	8.44 (0.32)	MAA*Sex F = $1.87 p = 0.158$	ns		
SMM (kg)	Whole sample	25.19 (0.35)	23.30 (0.26)	22.23 (0.35)	MAA status $F = 18.39 p < 0.0001$	Decel>Normal $p < 0.0001$ Decel>Accel $p < 0.0001$ Normal>Accel $p = 0.045$		
	Women	18.95 (0.51)	17.32 (0.30)	16.42 (0.48)	Sex $F = 1034.58 p < 0.0001$	Decel>Normal $p = 0.019$ Decel>Accel $p = 0.001$		
	Men	31.42 (0.49)	29.28 (0.42)	28.03 (0.52)	MAA*Sex F = $0.38 p = 0.685$	Decel>Normal $p = 0.003$ Decel>Accel $p < 0.0001$		
	Whole sample	19.92 (0.29)	18.53 (0.22)	17.72 (0.29)	MAA status $F = 14.50 p < 0.0001$	Decel>Normal $p = 0.001$ Decel>Accel $p < 0.0001$		
ASMM (kg)	Women	15.90 (0.42)	14.87 (0.25)	14.18 (0.40)	Sex $F = 573.86 p < 0.0001$	Decel>Accel $p = 0.010$		
	Men	23.93 (0.41)	22.18 (0.35)	21.26 (0.43)	MAA*Sex $F = 0.75 p = 0.473$	Decel>Accel $p < 0.0001$ Decel>Norm $p = 0.004$		
	Whole sample	1.56 (0.03)	1.52 (0.02)	1.39 (0.03)	MAA status $F = 11.52 p < 0.0001$	Decel>Accel $p < 0.0001$ Normal>Accel $p < 0.001$		
<i>n-</i> 10MWT (m/s)	Women	1.60 (0.04)	1.51 (0.02)	1.35 (0.04)	Sex $F = 0.008 p = 0.929$	Decel>Accel $p < 0.0001$ Normal>Accel $p < 0.001$		
	Men	1.51 (0.04)	1.53 (0.03)	1.42 (0.04)	MAA*Sex F = $2.35 p = 0.098$	ns		
a nn	Whole sample	11.98 (0.06)	11.90 (0.05)	11.78 (0.06)	MAA status F = $2.77 p = 0.065$	ns		
SPPB (score)	Women	11.96 (0.09)	11.87 (0.05)	11.81 (0.08)	Sex $F = 0.05 p = 0.827$	ns		
	Men	12.00 (0.09)	11.94 (0.07)	11.74 (0.09)	MAA*Sex F = $0.51 p = 0.604$	ns		
	Whole sample	5.82 (0.12)	6.30 (0.09)	7.26 (0.12)	MAA status $F = 39.67 p < 0.0001$	Decel <accel <math="">p < 0.001 Decel<normal <math="">p = 0.004 Normal<accel <math="">p < 0.001</accel></normal></accel>		
TUG (s)	Women	5.81 (0.17)	6.61 (0.10)	7.56 (0.16)	Sex $F = 10.23 \ p = 0.002$	Decel <normal <math="">p < 0.001 Decel<accel <math="">p < 0.0001 Normal<accel <math="">p < 0.0001</accel></accel></normal>		
	Men	5.84 (0.16)	5.99 (0.14)	6.95 (0.17)	MAA*Sex $F = 2.81 p = 0.062$	Decel <accel <math="">p < 0.0001 Normal<accel <math="">p < 0.0001</accel></accel>		

Six-MWT (m)	Whole sample	561.45 (9.08)	532.00 (6.69)	474.14 (9.10)	MAA status $F = 24.21 \ p < 0.0001$	Decel>Normal $p = 0.029$ Decel>Accel $p < 0.0001$ Normal>Accel $p = 0.001$
	Women	572.95 (13.10)	508.56 (7.84)	456.39 (12.35)	Sex $F = 4.19 p = 0.042$	Decel>Normal $p < 0.001$ Decel>Accel $p < 0.0001$ Normal>Accel $p = 0.001$
	Men	549.94 (12.58)	555.44 (10.85)	491.88 (13.38)	MAA*Sex $F = 4.99 p = 0.008$	Decel>Accel $p = 0.005$ Normal>Accel $p = 0.001$

All data are scores, which are shown as mean ± standard deviation. Abbreviations: Accel: accelerated agers; Decel: decelerated agers; Normal: normal agers; MAA: Muscle Age Acceleration; kg: kilograms; Five-TSTS: Five-time sit-to-stand; s, seconds; ns: no significance; SMM: Skeletal Muscle Mass; ASMM: Appendicular Skeletal Muscle Mass; n10MWT: 10Meter Walked Test-normal Walk; m: meter; SPPB: Short Physical Performance Battery; TUG: Timed Up and Go Test; Six-MWT: Six-minute Walking Test.

pattern of musculoskeletal ageing through a comprehensive estimate of sarcopenia, MAA, (2) test the ability of this tool to identify subclinical yet potentially prodromal musculoskeletal dysfunctions, and (3) verify whether MAA associated with health-related, biological features.

To these ends, we employed the screening tools recommended by EWGSOP2 consensus on sarcopenia [1], and the phenotypic clock developed by Levine [8], concurrently assessing morphological, motor-functional and biological markers, as increasingly advocated by researchers and practitioners active in ageing research [5].

In line with our hypothesis, using evidence-based, widely supported anthropometric and motor-functional tests allowed to build a marker of musculoskeletal ageing, which could identify early musculoskeletal ageing in a subgroup of apparently non-sarcopenic individuals. According to our estimates, these accelerated agers (approximately 25% of the cohort) would be at higher risk for developing sarcopenia.

Although EWGSOP2 tests individually did not result in detecting overtly sarcopenic individuals, participants who displayed a combination of even slight anthropometric and motor-functional alterations had a 4-to-6fold higher probability of developing sarcopenia, according to Ishii's formula [9]. A quantitative assessment would also potentially provide an objective, scalar physiological proxy of musculoskeletal ageing useful to investigate pathophysiological mechanisms subtending sarcopenia and to evaluate the effect of interventions in reversing/slowing this age-related condition [6]. In this regard, since TUG time and handgrip strength were extracted from the whole set of EWGSOP2 tools as major contributors to MAA both in women and men, substantial improvements in these selected tests following, for instance, reconditioning and strengthening exercise, would likely lead to decelerated musculoskeletal ageing. TUG and handgrip have been previously put forward as foremost predictors of independence and short-term mortality in the elderly, and reduced performances in these tests are taken as red flags [10, 11]. Indeed, the values at these tests are considered relevant markers of physical capability and

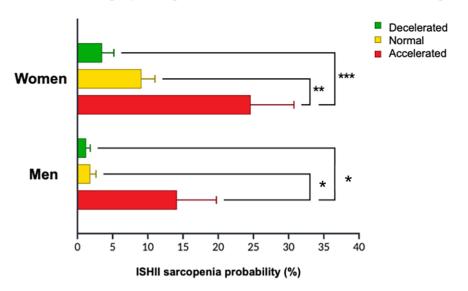


Figure 3. Probability of sarcopenia according to ISHII sarcopenia screening [11] in women and men based on Muscle Age Acceleration (MAA) status.

Table 3. Haemato-chemical sample results analysed by Muscle Age Acceleration (MAA) status and sex (N = 133, 33 decelerated, 67 normal, 33 decelerated agers; 53 men, 80 women).

			MAA status			Statistics
		Decelerated	Normal	Accelerated	Main effect	Pairwise comparisons
	Whole sample	5.46 (0.22)	6.00 (0.18)	6.10 (0.23)	MAA status $F = 2.55 p = 0.082$	ns
White Blood Cell × 10³/μL)	Women	5.29 (0.32)	5.56 (0.18)	5.58 (0.32)	Sex $F = 9.74 p = 0.002$	ns
(10 (Men	5.62 (0.31)	6.44 (0.30)	6.63 (0.32)	MAA*Sex F = $0.74 p = 0.480$	Decel <accel <math="">p = 0.024</accel>
	Whole sample	5.05 (0.10)	5.10 (0.08)	5.06 (0.10)	MAA status $F = 0.09 p = 0.915$	ns
Red Blood Cells × 10³/μL)	Women	4.70 (0.14)	4.91 (0.08)	5.05 (0.14)	Sex $F = 12.17 p = 0.001$	ns
	Men	5.41 (0.14)	5.29 (0.13)	5.08 (0.14)	MAA*Sex $F = 2.99 p = 0.054$	ns
	Whole sample	14.16 (0.20)	14.19 (0.16)	13.49 (0.21)	MAA status $F = 4.16 p = 0.018$	Decel <accel <math="">p < 0.0001 Normal<accel <math="">p < 0.0001</accel></accel>
Iemoglobin g/dL)	Women	13.56 (0.29)	13.59 (0.17)	12.76 (0.29)	Sex $F = 34.10 p < 0.0001$	Normal>Accel $p = 0.015$
	Men	14.76 (0.28)	14.80 (0.27)	14.21 (0.29)	MAA*Sex $F = 0.13 p = 0.879$	ns
	Whole sample	44.47 (0.54)	44.35 (0.43)	42.66 (0.55)	MAA status $F = 3.68 p = 0.028$	Normal>Accel $p = 0.048$
laematocrit test %)	Women	42.13 (0.77)	42.60 (0.44)	40.67 (0.77)	Sex $F = 48.11 p < 0.0001$	Normal>Accel $p = 0.032$
	Men	46.80 (0.75)	46.11 (0.73)	44.66 (0.77)	MAA*Sex $F = 0.36 p = 0.699$	Decel>Accel $p = 0.048$
Aean	Whole sample	88.27 (1.78)	87.87 (1.41)	85.48 (1.80)	MAA status $F = 0.74 p = 0.478$	ns
vican Corpuscular Volume (μm³)	Women	90.33 (2.55)	87.73 (1.46)	82.08 (2.55)	Sex $F = 0.26 p = 0.611$	Decel>Accel $p = 0.024$
(p)	Men	86.21 (2.47)	88.01 (2.40)	88.88 (2.55)	MAA*Sex $F = 2.36 p = 0.098$	ns
Mean	Whole sample	28.23 (0.66)	28.07 (0.52)	27.04 (0.67)	MAA status $F = 1.00 p = 0.371$	ns
orpuscular iemoglobin	Women	29.11 (0.94)	27.87 (0.54)	25.85 (0.94)	Sex $F = 0.23 p = 0.632$	Decel>Accel $p = 0.016$
10 × (Hb/RBC))	Men	27.35 (0.92)	28.28 (0.89)	28.23 (0.94)	MAA*Sex $F = 2.43 p = 0.092$	ns
Aean Corpuscular	Whole sample	31.85 (0.19)	31.98 (0.15)	31.60 (0.19)	MAA status $F = 1.22 p = 0.300$	ns
Corpuscular Iemoglobin Concentration	Women	32.14 (0.27)	31.88 (0.15)	31.41 (0.27)	Sex $F = 0.00 p = 0.965$	ns
g/dL)	Men	31.55 (0.26)	32.07 (0.25)	31.79 (0.27)	MAA*Sex $F = 1.98 p = 0.142$	ns
Red Blood Cell	Whole sample	13.80 (0.19)	13.72 (0.15)	14.06 (0.19)	MAA status $F = 0.99 p = 0.376$	ns
Red Blood Cell Distribution Vidth (%)	Women	13.50 (0.28)	13.72 (0.16)	14.32 (0.28)	Sex $F = 0.02 p = 0.892$	Decel <accel <math="">p = 0.038</accel>
Idili (70)	Men	14.09 (0.27)	13.72 (0.26)	13.81 (0.28)	MAA*Sex $F = 2.81 p = 0.133$	ns
Tomoglob!	Whole sample	2.50 (0.06)	2.49 (0.05)	2.51 (0.07)	MAA status $F = 0.04 p = 0.957$	ns
Iemoglobin listribution vidth (mg/dL)	Women	2.37 (0.09)	2.43 (0.05)	2.25 (0.09)	Sex $F = 3.00 p = 0.086$	ns
racii (ilig/uL)	Men	2.62 (0.09)	2.55 (0.09)	2.51 (0.09)	MAA*Sex $F = 1.06 p = 0.350$	ns
	Whole sample	216.07 (8.10)	232.83 (6.41)	226.06 (8.22)	MAA status $F = 1.32 p = 0.272$	ns
Platelets x 10 ⁹ /L)	Women	236.31 (11.63)	245.39 (6.65)	232.31 (11.63)	Sex $F = 8.75 p = 0.004$	ns
	Men	195.82 (11.28)	220.28 (10.96)	219.81 (11.63)	MAA*Sex $F = 0.74 p = 0.480$	ns
A Dl-4 1 4	Whole sample	8.18 (0.16)	8.07 (0.12)	8.18 (0.16)	MAA status $F = 0.24 p = 0.790$	ns
Mean Platelet Volume	Women	8.24 (0.23)	7.91 (0.13)	8.12 (0.23)	Sex $F = 0.37 p = 0.543$	ns
fL)	Men	8.12 (0.22)	8.23 (0.21)	8.24 (0.23)	MAA*Sex $F = 0.61 p = 0.546$	ns
	Whole sample	53.76 (1.43)	55.99 (1.13)	55.00 (1.45)	MAA status $F = 0.75 p = 0.472$	ns
Neutrophils %)	Women	51.82 (2.05)	53.20 (1.17)	52.91 (2.05)	Sex $F = 8.58 p = 0.004$	ns
,	Men	55.69 (1.99)	58.79 (1.949)	57.10 (2.05)	MAA*Sex F = $0.13 p = 0.875$	ns

	Whole sample	34.42 (1.33)	32.44 (1.05)	32.96 (1.35)	MAA status $F = 0.69 p = 0.503$	ns
Lymphocytes (%)	Women	37.12 (1.91)	35.56 (1.09)	34.80 (1.91)	Sex $F = 12.42 p = 0.001$	ns
,	Men	31.72 (1.85)	29.33 (1.80)	31.12 (1.91)	MAA*Sex $F = 0.28 p = 0.757$	ns
	Whole sample	5.99 (0.23)	6.05 (0.18)	6.27 (0.23)	MAA status F = $0.40 p = 0.669$	ns
Monocytes (%)	Women	5.62 (0.33)	5.61 (0.19)	6.29 (0.33)	Sex $F = 4.29 p = 0.040$	ns
()	Men	6.36 (0.32)	6.48 (0.31)	6.24 (0.33)	MAA*Sex $F = 1.30 p = 0.277$	ns
	Whole sample	2.83 (0.29)	2.75 (0.23)	3.27 (0.29)	MAA status F = $1.03 p = 0.359$	ns
Eosinophils (%)	Women	2.57 (0.41)	2.75 (0.23)	2.84 (0.41)	Sex $F = 2.27 p = 0.134$	ns
	Men	3.10 (0.40)	2.76 (0.39)	3.70 (0.41)	MAA*Sex F = $0.704 p = 0.496$	ns
	Whole sample	0.69 (0.05)	0.66 (0.04)	0.67 (0.05)	MAA status F = $0.10 p = 0.903$	ns
Basophils (%)	Women	0.68 (0.07)	0.69 (0.04)	0.74 (0.07)	Sex $F = 1.38 p = 0.242$	ns
(, -)	Men	0.69 (0.07)	0.63 (0.07)	0.60 (0.07)	MAA*Sex F = $0.597 p = 0.552$	ns
	Whole sample	2.31 (0.13)	2.08 (0.11)	2.47 (0.14)	MAA status F = 2.60 p = 0.078	ns
Large Unstained Cells	Women	2.19 (0.19)	2.18 (0.11)	2.44 (0.19)	Sex F = 0.05 p = 0.823	ns
(%)	Men	2.42 (0.19)	1.98 (0.18)	2.50 (0.19)	MAA*Sex F = $0.85 p = 0.428$	ns
	Whole sample	2.99 (0.16)	3.38 (0.13)	3.36 (0.16)	MAA status F = 2.20 p = 0.115	ns
Neutrophils (× 10³/μL)	Women	2.80 (0.23)	2.98 (0.13)	3.01 (0.23)	Sex F = 13.55 p < 0.001	ns
(^ 10 ⁻ /µL)	Men	3.17 (0.22)	3.79 (0.21)	3.71 (0.23)	MAA*Sex F = $0.63 p = 0.533$	Decel <normal <math="">p = 0.044</normal>
	Whole sample	1.85 (0.10)	1.93 (0.08)	1.97 (0.10)	MAA status $F = 0.40 p = 0.673$	ns
Lymphocytes (× $10^3/\mu$ L)	Women	1.93 (0.14)	1.98 (0.08)	1.90 (0.14)	Sex $F = 0.11 p = 0.74$	ns
	Men	1.77 (0.14)	1.89 (0.13)	2.04 (0.14)	MAA*Sex F = $0.60 p = 0.548$	ns
	Whole sample	0.33 (0.02)	0.35 (0.01)	0.39 (0.02)	MAA status $F = 3.69 p = 0.028$	Decel <accel <math="">p = 0.023</accel>
Monocytes	Women	0.29 (0.02)	0.31 (0.01)	0.36 (0.02)	Sex $F = 16.25 p$	ns
(× 10 ³ /μL)		, ,	, ,	,	0.0001 MAA*Sex	
	Men	0.36 (0.02)	0.39 (0.02)	0.42 (0.02)	F = 0.14 p = 0.868 MAA status	ns
Eosinophils	Whole sample	0.16 (0.02)	0.17 (0.01)	0.22 (0.02)	F = 2.89 p = 0.059 Sex	ns
εσεπισμιτίε (× 10³/μL)	Women	0.14 (0.03)	0.15 (0.02)	0.17 (0.03)	F = 6.41 p = 0.013	ns $Dand < A and n = 0.015$
	Men	0.17 (0.03)	0.18 (0.03)	0.26 (0.03)	MAA*Sex F = 1.01 p = 0.365	Decel <accel <math="">p = 0.015 Normal<accel <math="">p = 0.033</accel></accel>
	Whole sample	0.03 (0.02)	0.04 (0.01)	0.04 (0.02)	MAA status $F = 0.09 p = 0.913$	ns
Basophils (x10³/μL)	Women	0.03 (0.02)	0.06 (0.01)	0.04 (0.02)	Sex $F = 0.43 p = 0.515$	ns
	Men	0.04 (0.02)	0.02 (0.02)	0.03 (0.02)	MAA*Sex F = $0.527 p = 0.592$	ns
Larga Unstained	Whole sample	0.13 (0.01)	0.13 (0.01)	0.16 (0.01)	MAA status $F = 2.84 p = 0.062$	ns
Large Unstained Cells (× 10³/μL)	Women	0.11 (0.02)	0.12 (0.01)	0.14 (0.02)	Sex $F = 3.95 p = 0.049$	ns
(10 / MI)	Men	0.14 (0.01)	0.13 (0.01)	0.17 (0.02)	MAA*Sex $F = 0.15 p = 0.864$	ns
	Whole sample	94.75 (2.09)	95.17 (1.66)	89.44 (2.09)	MAA status $F = 2.57 p = 0.081$	ns
Glycemia (mg/dL)	Women	93.31 (2.96)	92.98 (1.69)	86.00 (2.96)	Sex $F = 4.33 p = 0.039$	ns
	Men	96.19 (2.96)	97.35 (2.87)	92.88 (2.96)	MAA*Sex $F = 0.23 p = 0.791$	ns
					•	
	Whole sample	35.32 (1.64)	37.62 (1.30)	37.53 (1.66)	MAA status $F = 0.69 p = 0.505$	ns
Urea (mg/dL)	Whole sample Women	35.32 (1.64) 33.94 (2.35)	37.62 (1.30) 35.51 (1.34)	37.53 (1.66) 35.75 (2.35)	MAA status F = 0.69 p = 0.505 Sex F = 3.89 p = 0.051	ns ns

	Whole sample	0.90 (0.02)	0.88 (0.02)	0.85 (0.03)	MAA status $F = 0.874 p = 0.422$	ns
Creatinine (mg/dL)	Women	0.77 (0.04)	0.77 (0.02)	0.80 (0.04)	Sex $F = 48.5 p < 0.0001$	ns
,	Men	1.03 (0.03)	0.99 (0.04)	0.91 (0.04)	MAA*Sex F = $2.60 p = 0.083$	Decel>Accel $p = 0.031$
	Whole sample	7.08 (0.07)	7.07 (0.06)	6.97 (0.07)	MAA status $F = 0.86 p = 0.426$	ns
Fotal protein (g/dL)	Women	7.06 (0.10)	7.11 (0.06)	6.93 (0.10)	Sex $F = 0.08 p = 0.782$	ns
	Men	7.11 (0.10)	7.03 (0.10)	7.01 (0.10)	MAA*Sex F = $0.45 p = 0.641$	ns
	Whole sample	4.26 (0.04)	4.27 (0.03)	4.18 (0.04)	MAA status $F = 1.73 p = 0.182$	ns
Albumin (g/dL)	Women	4.26 (0.05)	4.26 (0.03)	4.14 (0.05)	Sex $F = 0.66 p = 0.419$	ns
	Men	4.26 (0.05)	4.27 (0.05)	4.23 (0.05)	MAA*Sex F = $0.39 p = 0.677$	ns
Alkaline	Whole sample	75.65 (3.61)	80.34 (2.86)	73.25 (3.67)	MAA status $F = 1.27 p = 0.283$	ns
phosphatase (U/L)	Women	79.25 (5.18)	74.79 (2.96)	76.12 (5.18)	Sex $F = 0.02 p = 0.883$	ns
(0/L)	Men	72.06 (5.03)	85.94 (4.89)	70.38 (5.18)	MAA*Sex $F = 2.66 p = 0.074$	ns
	Whole sample	98.19 (4.81)	90.83 (3.89)	91.94 (4.89)	MAA status $F = 0.76 p = 0.472$	ns
Serum iron (μg/dL)	Women	91.19 (6.91)	89.59 (3.95)	87.44 (6.91)	Sex $F = 2.60 p = 0.109$	ns
	Men	105.18 (6.70)	92.06 (6.70)	96.44 (6.91)	MAA*Sex F = $0.45 p = 0.640$	ns
C-reactive	Whole sample	0.40 (0.03)	0.38 (0.02)	0.38 (0.03)	MAA status $F = 0.30 p = 0.738$	ns
protein	Women	0.42 (0.04)	0.35 (0.02)	0.40 (0.04)	Sex $F = 0.27 p = 0.604$	ns
(mg/dL)	Men	0.38 (0.04)	0.40 (0.03)	0.35 (0.04)	MAA*Sex $F = 1.50 p = 0.227$	ns
Interleukin 6 (ng/mL)	Whole sample	790.75 (46.06)	722.22 (36.45)	756.96 (46.76)	MAA status $F = 0.69 p = 0.502$	ns
	Women	715.26 (66.12)	689.17 (37.78)	694.72 (66.12)	Sex $F = 5.18 p = 0.025$	ns
	Men	866.23 (64.15)	755.28 (62.34)	819.21 (66.12)	MAA*Sex F = $0.29 p = 0.750$	ns
Insulin-like	Whole sample	0.76 (0.14)	0.81 (0.12)	0.74 (0.15)	MAA status $F = 0.07 p = 0.933$	ns
growth factor-1 (ng/ml)	Women	0.98 (0.21)	0.81 (0.12)	0.77 (0.21)	Sex $F = 1.20 p = 0.276$	ns
(lig/ilil)	Men	0.54 (0.19)	0.80 (0.21)	0.70 (0.22)	MAA*Sex F = $0.75 p = 0.476$	ns
	Whole sample	452.10 (60.03)	495.45 (47.50)	374.81 (60.94)	MAA status $F = 1.22 p = 0.299$	ns
Leptin (ng/mL)	Women	469.38 (86.18)	435.64 (49.24)	442.05 (86.18)	Sex $F = 0.06 p = 0.801$	ns
	Men	434.82 (83.60)	555.26 (81.25)	307.58 (86.18)	MAA*Sex $F = 1.44 p = 0.242$	ns
Plasma	Whole sample	5.09 (0.15)	4.63 (0.12)	4.54 (0.15)	MAA status $F = 4.00 p = 0.021$	Decel>Accel $p = 0.035$
protein-SH (micromol/g/	Women	5.01 (0.22)	4.58 (0.13)	4.44 (0.22)	Sex $F = 0.89 p = 0.346$	ns
proteins)	Men	5.17 (0.21)	4.67 (0.20)	4.64 (0.22)	MAA*Sex F = $0.04 p = 0.963$	ns
	Whole sample	2.73 (0.20)	2.73 (0.16)	2.40 (0.21)	MAA status $F = 0.93 \ p = 0.398$	ns
Malondialdehyde (micromol/L)	Women	2.35 (0.29)	2.32 (0.17)	1.94 (0.29)	Sex $F = 13.76 p < 0.001$	ns
	Men	3.11 (0.28)	3.13 (0.28)	2.85 (0.29)	MAA*Sex F = $0.03 p = 0.968$	ns
	Whole sample	159.09 (15.51)	168.53 (12.34)	129.39 (16.00)	MAA status $F = 1.92 p = 0.152$	ns
Paraoxonase (U/L)	Women	169.78 (22.26)	153.16 (12.99)	131.30 (22.99)	Sex $F = 0.01 p = 0.914$	ns
	Men	148.39 (21.59)	183.90 (20.99)	127.48 (22.26)	MAA*Sex F = $0.93 p = 0.394$	ns
	Whole sample	1.17 (0.08)	1.04 (0.06)	1.17 (0.08)	MAA status $F = 1.15 p = 0.318$	ns
Homocysteine (micromol/L)	Women	1.29 (0.12)	1.03 (0.07)	1.19 (0.11)	Sex $F = 1.03 p = 0.313$	ns
,	Men	1.05 (0.11)	1.05 (0.11)	1.15 (0.11)	MAA*Sex F = $0.85 p = 0.430$	ns

All data are scores, which are shown as mean \pm standard deviation. Abbreviations: MAA: Muscle Age Acceleration; μ L: microliters; g: grams; ns: no significance; mg/dl: milligrams/deciliters; μ m³: cubic micrometer; hb/RBC: hemoglobin/Red Blood Cell; L: liters; fL: femtolitreng/mL nanograms/milliliters; U: unit.

healthy ageing [12] and qualify as practically relevant endpoints of treatment to appraise the effectiveness of preventive/therapeutic approaches. TUG, which implies rising from a chair, accelerating to a target, turn around it and return to the starting position, embodies physical and executive functions that are critical for independence and safety but subjected to substantial age-related decline.

As a preliminary validation of the newly introduced clock of musculoskeletal ageing, we found it strongly associated with PhenoAge, as calculated by the ensemble of blood-based markers contributing to the Levine's epigenetic clock [8]. However, only the latter correlated in a significant manner to major health risks like coronary and cardiovascular diseases, atherosclerosis, and mortality. This would suggest that these musculoskeletal and biological proxies of ageing process address different health domains that could be

comprehensively captured their by concurrent evaluation. This seems particularly interesting from a pathophysiological standpoint as it suggests that the musculoskeletal system may start to decline even when signs of increased health risk are absent or not yet developed. In this context, sarcopenia would follow a distinct, anticipated trajectory, as is often the case with early-onset sarcopenia, which onsets subclinically as soon as fifth or even fourth decade of life. It also points toward what is possibly a closer relationship between lifestyle and musculoskeletal well-being, the latter being more strongly influenced by sedentariness and the inherent lack of mechanical loading than systemic health (as resampled by PhenoAge).

As such, our data point toward a complementary use of the tools here investigated, with MAA serving as a practical, low-cost independent predictor of sarcopenia, and PhenoAge as a blood-based estimate of major

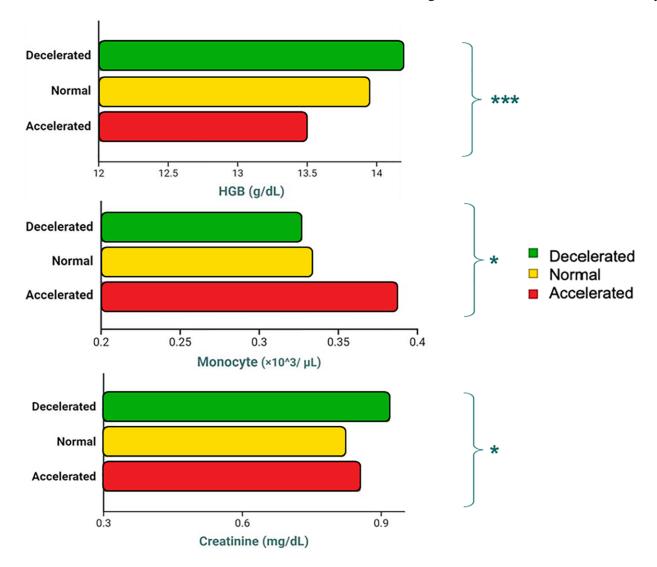


Figure 4. Differences in exemplificative blood-based markers depending on Muscle Age Acceleration (MAA) status (Decelerated vs. Normal vs. Accelerated).

Table 4. Correlation matrix of Muscle Age Acceleration and Phenotypic Age Acceleration with health risk scores.

		MAA	PhenoAA	FRS	АССАНА	STERN	MESA	CCI	Mortality score
MAA	Whole sample	-	0,014	-0,150	-0,032	-0,173	0,027	-0,061	-0,149*
	Women	-	0,135	-0,182	-0,123	-0,117	0,008	-0,122	-0,028
	Men	_	-0,120	-0,135	-0,075	-0,179	0,082	0,027	-0,252*
PhenoAA	Whole sample	0,014	-	0,280*	0,220	0,245**	0,314*	0,004	0,431**
	Women	0,135	-	0,067	0,149	0,406**	0,167	0,027	0,398**
	Men	-0,120	-	-0,120	-0,021	0,148	0,062	-0,034	0,463**

Data are described as Spearman's Rho. *Correlation is significant at the 0.05 level (1-tailed); **Correlation is significant at the 0.01 level (1-tailed). Abbreviations: MAA: Muscle Age Acceleration; PhenoAA: Phenotypic Age Acceleration; FRS: Framingham risk score coronary heart disease; ACCAHA: American College of Cardiology and American Heart Association cardiovascular diseases risk score; STERN: Stern Diabetes Risk; MESA: Multi-Ethnic Study of Atherosclerosis risk score; CCI: Charlson Comorbidity Index.

health issues, which, as such, are not suited to capture musculoskeletal deterioration. While our data contribute to a growing literature supporting the utility of biological clocks to estimate risk profiles, especially for cardiovascular health, further research is needed to determine the extent to which biological clocks can be used as clinical biomarkers [13].

As a further validation of MAA, once the cohort was classified according to MAA (decelerated/normal/ accelerated), several health-related and haematochemical scores polarized into a decelerated/healthier pattern as opposed to accelerated/unhealthier, with normal agers generally located among the two clusters vet generally closer to decelerated ones. It nevertheless has to be said that our cohort exhibited haematochemical values falling within normative ranges in all domains, even though the subgroup of accelerated agers displayed biological features (e.g., monocyte and eosinophil count, haemoglobin mass, creatinine, etc.) compatible with latent, low-grade inflammation and higher activation of the immune system that are signature elements of overt sarcopenia [14, 15]. Relatedly, given that systemic inflammation plays a key role in muscle aging and considering the bidirectional communication and influence between the gut microbiota (the community of microorganisms living in the intestines) and skeletal muscle, integrating gut microbiome alterations into the discussion could strengthen the argument that muscle aging is not only a localized process but influenced by systemic metabolic and inflammatory factors [16, 17]. Clearly, such factors concur and synergize in the frailty of the elderly, a multidimensional phenomenon involving musculoskeletal decline, systemic aging, and resilience loss [18]. In this framework, MAA may represent a potential predictor and early marker of frailty severity and progression as well as an endpoint of treatment if interventions to mitigate frailty are administered.

Longitudinal profiling of our cohort through follow-up evaluations of musculoskeletal and biological ageing will provide a unique opportunity to further validate MAA through a prevalence study to confirm or reject the hypothesis that accelerated agers have significantly higher likelihood of developing sarcopenia than normal and decelerated agers to evolve from subclinical features to a definite diagnosis of sarcopenia at threefive years from the current screening. Such follow-up re-evaluations with prevalence data will also allow to validate the regression-based predictive equations developed in our cohort, leading to the establishment of straightforward formulae that can be employed to quantify musculoskeletal ageing in research and clinical settings. So far, we have developed preliminary weighted-average equations for estimating individual Muscle Age based on the parsimonious model of best predictors here extracted (see: Supplementary Material 1).

Study limitations

The first limitation relates to the population under study, as the present findings were obtained from healthy, non-sarcopenic subjects, who were not compared in terms of musculoskeletal and biological ageing arcs to comparable yet overtly sarcopenic individuals. Upcoming controlled studies are needed to appraise the validity of MAA to differentiate between sarcopenic and non-sarcopenic subjects and capture different degrees of sarcopenia severity. Another weakness relates to the sample over which validation of MAA against biological, haemato-chemical markers could be completed, as approximately 60% of the original cohort underwent both musculoskeletal and biological testing.

Finally, the cross-sectional design chosen for this study cannot dynamically portray the curve of the individual ageing process over a defined timespan, preventing comparisons of predicted versus actual trajectories. To do so, a longitudinal design with repeated follow-up measurements is warranted, also taking potential confounders like (but not limited to) lifestyle, physical activity levels, comorbidities and socioeconomic profile into account. This effort has been initiated and is currently underway for the present cohort. Repeated measurements would also allow to establish the testretest consistency and responsiveness of MAA by calculating indexes such as intraclass correlation coefficient, standard error of measurement and minimal detectable difference, which would strengthen the reliability of upcoming studies.

CONCLUSIONS

Non-sarcopenic, community-dwelling middle-aged and older adults could be validly classified in terms of their individual musculoskeletal ageing trajectories with a novel muscular clock, MAA. This comprehensive estimate, developed from the EWGSOP2 framework, could identify signs of accelerated musculoskeletal ageing when subclinical, slight alterations presented concurrently. Participants with such signs were also those displaying slightly altered haemato-chemical features.

If validated through a properly planned, populationbased, controlled, longitudinal study, this novel estimate may represent an objective tool for a comprehensive and cost-effective evaluation of the musculoskeletal wellbeing of the elderly. In this perspective, MAA may complement and strengthen other quantitative and qualitative tools for sarcopenia screening including, but not limited to, the Ishii test for sarcopenia [9] and the SARC-F questionnaire [19]. The parsimonious models consisting of three very simple and common motor tests could be easily administered both in clinical and community settings. To this end, the simple formula here provided (see: Results) will allow to estimate MAA on a vast scale, at least for screening purposes, until a more robust validation of this tool is attained, potentially green flagging more diagnostic applications.

MATERIALS AND METHODS

This cross-sectional study was conducted at the Department of Biomedical Sciences, University of Sassari, Italy, from September 2020 to December 2023. Potential candidates were searched through phone interviews and public engagement initiatives.

Participants were required to be aged 50-90 years with no medical, physical, or cognitive conditions that might interfere with functional assessments. After

eligibility screening, a clinical examination assessed exclusion criteria like severe respiratory, neurological, or cardiovascular diseases, recent injuries, and metabolic conditions. The cognitive status was also monitored by Montreal Cognitive Assessment (MOCA), as well as functioning and independence during activities of daily living (ADL) [20] and instrumental ADL (IADL) [21] through questionnaires.

Sarcopenia assessment

Participants were evaluated according to the EWGSOP2 guidelines using the Find-Assess-Confirm-Severity approach [1]. Sarcopenia was screened with the Ishii test, which calculates risk using an equation combining age, grip strength, and calf circumference. A score >105 for men and >120 for women indicates high probability of sarcopenia [9].

Muscle strength

Handgrip strength (HGS) was measured using a handgrip dynamometer (G200, Biometrics Ltd., UK) over three trials, with one-minute rests. The 5-times sitto-stand test (5-TSTS) was performed to estimate lower body strength [22].

Body composition

Skeletal muscle mass (SMM) and appendicular (ASMM) were assessed via bioelectrical impedance analysis (BIA 101 BIVA; Akern, Italy) [23].

Physical performance

Gait speed was measured with the 10-Meter Walk Test (10MWT) at a comfortable walking pace [24]. Physical performance was evaluated by Short Physical Performance Battery (SPPB), assessing gait speed, balance, and strength [24]. The Timed Up and Go Test (TUG) was used to assess fall risk, balance and functional mobility [25]. The distance covered during the Six-minute Walk Test (Six-MWT) was recorded to evaluate cardiovascular fitness [26].

Blood analyses and biomarkers

Blood samples were analysed for several key parameters, including white blood cells (WBC), red blood cells (RBC), haemoglobin (HGB), haematocrit (HCT), creatinine, urea, alkaline phosphatase, serum iron, and C-reactive protein (CRP). Concentrations of interleukin-6 (IL-6), leptin, tumour necrosis factoralpha (TNF-α), and insulin-like growth factor 1 (IGF-1) were measured by ELISA.

Oxidative stress markers were also assessed, with paraoxonase-1 (PON-1) activity measured using paraoxon as a substrate, protein-SH (PSH) levels, and malondialdehyde (MDA) levels.

Selected blood-measured biomarkers were combined to calculate the PhenoAge, aka biological age, according to Levine [8].

Risk scores for selected major health issues were calculated using the Framingham Risk Score (FRS) for coronary heart disease [27], CVD risk from the American College of Cardiology (ACC) and American Heart Association (AHA) [28], and 10-year risk of atherosclerosis based on the Multi-Ethnic Study of Atherosclerosis (MESA) study [29]. The risk of diabetes 2 [30], 10-year mortality risk score [8], and Charlson Comorbidity Index (CCI) [31] were also calculated.

Statistical analysis

Muscle age and muscle age acceleration modelling

Muscle Age and its derivative measure of acceleration MAA were modelled separately for men and women to account for sex differences in ageing trajectories, using a two-phase approach. First, all tests from EWGSOP2 consensus were used as predictors. Muscle Age was modelled with Elastic Net regression (ElasticNetCV, Scikit-learn v.1.5.1), with 100 permutations and 80/20 train-test splits [32]. Five-fold cross-validation was used to optimize regularisation strength (alpha) and the balance between L1 and L2 penalties (11 ratio). Average root mean square error (RMSE) of the differences between predicted and observed values and R² of the regression across the 100 permutations were reported as metrics of robustness and stability of the predictive models. Predictor coefficients representing the relative importance of each feature were averaged across permutations for robustness. Secondly, for parsimonious modelling, only those predictors with beta coefficients ≥1 standard deviation were included in refined, sex-specific models, using the same Elastic Net method. Sex-specific models were developed to reflect the established sexual differences in aging that are evident in the early and later phases of adulthood and senescence.

For both phases, MAA was calculated as residuals from the linear regression of predicted versus chronological age. Subjects were then categorised into three groups (MAA_status) based on MAA scores: decelerated (0–25th percentile), normal (26th-75th percentile), and accelerated (above 76th percentile).

Shapiro-Wilk's, Mauchly's and Levene's tests were performed to appraise data normality, sphericity and

homogeneity. A general linear model analysis of variance (ANOVA) was conducted to examine the main effects of MAA_status, sex and the interaction MAA_status*sex on functional and health-related outcomes. Pairwise comparisons were adjusted (Bonferroni) to reduce type-I error rate.

Correlation analysis was performed to assess the association between MAA and other continuous variables, including the relationship with PhenoAge acceleration (PhenoAA) which, likewise MAA, was calculated as the residuals from the linear regression of predicted versus chronological age.

New and noteworthy

This study introduces an evidence-based phenotypic clock, the Muscle Age Acceleration (MAA), which could identify signs of accelerated musculoskeletal ageing when subclinical, slight alterations presented concurrently. Participants with such signs were also those displaying slightly altered haemato-chemical features. If validated through a properly planned, population-based, controlled longitudinal study, this novel predictor of musculoskeletal decline may represent an objective tool for a comprehensive and cost-effective evaluation of sarcopenia for individuals transitioning from middle-age to senescence.

Data availability

All datasets are stored at the Department of Biomedical Sciences, University of Sassari, Sassari, Italy, under the responsibility of Professor Franca Deriu. Data will be made available upon reasonable request.

AUTHOR CONTRIBUTIONS

LV, FD and AM conceived and designed research, AC, MM, GM, AB, MGC, BM and NL performed experiments, OY, GF, GS analyzed data, AM and FD interpreted results of experiments, LV, SC and DC prepared figures, LV and AM drafted manuscript, MM, AZ, CC, FD and AM edited and revised manuscript, all authors approved final version of manuscript.

ACKNOWLEDGMENTS

We are indebted to all the volunteers who made this work possible.

CONFLICTS OF INTEREST

The authors declare no conflicts of interest related to this study.

ETHICAL STATEMENT AND CONSENT

The study was approved by the Institutional Review Board (PNRR-MAD-2022-12376667) and conducted in accordance with the Declaration of Helsinki. Written informed consent was obtained from all participants.

FUNDING

This work was funded by European Union – Next Generation EU – PNRR M6C2 – Investimento 2.1 Rafforzamento, valorizzazione e potenziamento della ricerca biomedica del SSN - finanziato dall'Unione europea NextGenerationEU PNRR-MAD-2022-12376667 (recipient: Franca Deriu).

REFERENCES

PMID:22108098

 Cruz-Jentoft AJ, Bahat G, Bauer J, Boirie Y, Bruyère O, Cederholm T, Cooper C, Landi F, Rolland Y, Sayer AA, Schneider SM, Sieber CC, Topinkova E, et al, and Writing Group for the European Working Group on Sarcopenia in Older People 2 (EWGSOP2), and the Extended Group for EWGSOP2. Sarcopenia: revised European consensus on definition and diagnosis. Age Ageing. 2019; 48:16–31.

https://doi.org/10.1093/ageing/afy169 PMID:30312372

- Lynch GS. Sarcopenia. In: Mooren FC, editor. Encyclopedia of Exercise Medicine in Health and Disease. Berlin, Heidelberg: Springer Berlin Heidelberg. 2012; 775–9. https://doi.org/10.1007/978-3-540-29807-6
- Beyer I, Mets T, Bautmans I. Chronic low-grade inflammation and age-related sarcopenia. Curr Opin Clin Nutr Metab Care. 2012; 15:12–22. https://doi.org/10.1097/MCO.0b013e32834dd297
- Santilli V, Bernetti A, Mangone M, Paoloni M. Clinical definition of sarcopenia. Clin Cases Miner Bone Metab. 2014; 11:177–80. PMID:25568649
- Curcio F, Ferro G, Basile C, Liguori I, Parrella P, Pirozzi F, Della-Morte D, Gargiulo G, Testa G, Tocchetti CG, Bonaduce D, Abete P. Biomarkers in sarcopenia: A multifactorial approach. Exp Gerontol. 2016; 85:1–8. https://doi.org/10.1016/j.exger.2016.09.007
 PMID:27633530
- Cawthon PM, Lui LY, Taylor BC, McCulloch CE, Cauley JA, Lapidus J, Orwoll E, Ensrud KE. Clinical Definitions of Sarcopenia and Risk of Hospitalization in Community-Dwelling Older Men: The Osteoporotic Fractures in Men Study. J Gerontol A Biol Sci Med Sci. 2017; 72:1383–9.

https://doi.org/10.1093/gerona/glw327 PMID:28329087

7. Cruz-Jentoft AJ, Baeyens JP, Bauer JM, Boirie Y, Cederholm T, Landi F, Martin FC, Michel JP, Rolland Y, Schneider SM, Topinková E, Vandewoude M, Zamboni M, and European Working Group on Sarcopenia in Older People. Sarcopenia: European consensus on definition and diagnosis: Report of the European Working Group on Sarcopenia in Older People. Age Ageing. 2010; 39:412–23.

https://doi.org/10.1093/ageing/afq034 PMID:20392703

8. Levine ME, Lu AT, Quach A, Chen BH, Assimes TL, Bandinelli S, Hou L, Baccarelli AA, Stewart JD, Li Y, Whitsel EA, Wilson JG, Reiner AP, et al. An epigenetic biomarker of aging for lifespan and healthspan. Aging (Albany NY). 2018; 10:573–91.

https://doi.org/10.18632/aging.101414 PMID:29676998

 Ishii S, Tanaka T, Shibasaki K, Ouchi Y, Kikutani T, Higashiguchi T, Obuchi SP, Ishikawa-Takata K, Hirano H, Kawai H, Tsuji T, Iijima K. Development of a simple screening test for sarcopenia in older adults. Geriatr Gerontol Int. 2014 (Suppl 1); 14:93–101.

https://doi.org/10.1111/ggi.12197 PMID:24450566

 Chua KY, Lim WS, Lin X, Yuan JM, Koh WP. Handgrip Strength and Timed Up-and-Go (TUG) Test are Predictors of Short-Term Mortality among Elderly in a Population-Based Cohort in Singapore. J Nutr Health Aging. 2020; 24:371–8.

https://doi.org/10.1007/s12603-020-1337-0 PMID:32242204

 Manca A, Fiorito G, Morrone M, Boi A, Mercante B, Martinez G, Ventura L, Delitala AP, Cano A, Catte MG, Solinas G, Melis F, Ginatempo F, Deriu F. A novel estimate of biological aging by multiple fitness tests is associated with risk scores for age-related diseases. Front Physiol. 2023; 14:1164943.

https://doi.org/10.3389/fphys.2023.1164943 PMID:37228822

 Lara J, Cooper R, Nissan J, Ginty AT, Khaw KT, Deary IJ, Lord JM, Kuh D, Mathers JC. A proposed panel of biomarkers of healthy ageing. BMC Med. 2015; 13:222.

https://doi.org/10.1186/s12916-015-0470-9 PMID:26373927

13. Fransquet PD, Ryan J. The current status of blood epigenetic biomarkers for dementia. Crit Rev Clin Lab Sci. 2019; 56:435–57.

https://doi.org/10.1080/10408363.2019.1639129 PMID:31328605 14. Cooper C, Dere W, Evans W, Kanis JA, Rizzoli R, Sayer AA, Sieber CC, Kaufman JM, Abellan van Kan G, Boonen S, Adachi J, Mitlak B, Tsouderos Y, et al. Frailty and sarcopenia: definitions and outcome parameters. Osteoporos Int. 2012; 23:1839–48. https://doi.org/10.1007/s00198-012-1913-1 PMID:22290243

 Liang Z, Zhang T, Liu H, Li Z, Peng L, Wang C, Wang T. Inflammaging: The ground for sarcopenia? Exp Gerontol. 2022; 168:111931. https://doi.org/10.1016/j.exger.2022.111931

PMID:35985553

16. Meléndez-Oliva E, Martínez-Pozas O, Sinatti P, Martín Carreras-Presas C, Cuenca-Zaldívar JN, Turroni S, Sánchez Romero EA. Relationship Between the Gut

Microbiome, Tryptophan-Derived Metabolites, and Osteoarthritis-Related Pain: A Systematic Review with Meta-Analysis. Nutrients. 2025; 17:264.

https://doi.org/10.3390/nu17020264 PMID:39861394

 Gonzalez-Alvarez ME, Sanchez-Romero EA, Turroni S, Fernandez-Carnero J, Villafañe JH. Correlation between the Altered Gut Microbiome and Lifestyle Interventions in Chronic Widespread Pain Patients: A Systematic Review. Medicina (Kaunas). 2023; 59:256. https://doi.org/10.3390/medicina59020256 PMID:36837458

 Fernández-Carnero S, Martínez-Pozas O, Cuenca-Zaldívar JN, Sánchez-Romero EA. Addressing frailty in older adults: an integrated challenge for health, science, and society. Aging (Albany NY). 2024; 16:13176–80.

https://doi.org/10.18632/aging.206162 PMID:39611815

 Malmstrom TK, Miller DK, Simonsick EM, Ferrucci L, Morley JE. SARC-F: a symptom score to predict persons with sarcopenia at risk for poor functional outcomes. J Cachexia Sarcopenia Muscle. 2016; 7:28– 36

https://doi.org/10.1002/jcsm.12048 PMID:<u>27066316</u>

Katz S. Assessing self-maintenance: activities of daily living, mobility, and instrumental activities of daily living. J Am Geriatr Soc. 1983; 31:721–7.
 https://doi.org/10.1111/j.1532-5415.1983.tb03391.x

 PMID:6418786

21. Lawton MP, Brody EM. Assessment of older people: self-maintaining and instrumental activities of daily living. Gerontologist. 1969; 9:179–86. PMID:5349366

22. Khuna L, Soison T, Plukwongchuen T, Tangadulrat N. Reliability and concurrent validity of 30-s and 5-time

sit-to-stand tests in older adults with knee osteoarthritis. Clin Rheumatol. 2024; 43:2035–45. https://doi.org/10.1007/s10067-024-06969-6 PMID:38634965

 Janssen I, Heymsfield SB, Baumgartner RN, Ross R. Estimation of skeletal muscle mass by bioelectrical impedance analysis. J Appl Physiol (1985). 2000; 89:465–71.

https://doi.org/10.1152/jappl.2000.89.2.465 PMID:10926627

Perera S, Mody SH, Woodman RC, Studenski SA. Meaningful change and responsiveness in common physical performance measures in older adults. J Am Geriatr Soc. 2006; 54:743–9.
 https://doi.org/10.1111/j.1532-5415.2006.00701.x
 PMID:16696738

25. Podsiadlo D, Richardson S. The timed "Up & Go": a test of basic functional mobility for frail elderly persons. J Am Geriatr Soc. 1991; 39:142–8. https://doi.org/10.1111/j.1532-5415.1991.tb01616.x PMID:1991946

26. ATS Committee on Proficiency Standards for Clinical Pulmonary Function Laboratories. ATS statement: guidelines for the six-minute walk test. Am J Respir Crit Care Med. 2002; 166:111–7. https://doi.org/10.1164/ajrccm.166.1.at1102 PMID:12091180

D'Agostino RB Sr, Vasan RS, Pencina MJ, Wolf PA, Cobain M, Massaro JM, Kannel WB. General cardiovascular risk profile for use in primary care: the Framingham Heart Study. Circulation. 2008; 117:743–53.
 https://doi.org/10.1161/CIRCULATIONAHA.107.6995
79

PMID: 18212285

28. Goff DC Jr, Lloyd-Jones DM, Bennett G, Coady S, D'Agostino RB, Gibbons R, Greenland P, Lackland DT, Levy D, O'Donnell CJ, Robinson JG, Schwartz JS, Shero ST, et al, and American College of Cardiology/American Heart Association Task Force on Practice Guidelines. 2013 ACC/AHA guideline on the assessment of cardiovascular risk: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines. Circulation. 2014; 129:S49–73.

https://doi.org/10.1161/01.cir.0000437741.48606.98 PMID:24222018

29. Budoff MJ, Young R, Burke G, Jeffrey Carr J, Detrano RC, Folsom AR, Kronmal R, Lima JAC, Liu KJ, McClelland RL, Michos E, Post WS, Shea S, et al. Tenyear association of coronary artery calcium with atherosclerotic cardiovascular disease (ASCVD) events: the multi-ethnic study of atherosclerosis (MESA). Eur Heart J. 2018; 39:2401–8.

- https://doi.org/10.1093/eurheartj/ehy217 PMID:29688297
- 30. Stern MP, Williams K, Haffner SM. Identification of persons at high risk for type 2 diabetes mellitus: do we need the oral glucose tolerance test? Ann Intern Med. 2002; 136:575–81.

https://doi.org/10.7326/0003-4819-136-8-

200204160-00006

PMID:<u>11955025</u>

- 31. Roffman CE, Buchanan J, Allison GT. Charlson Comorbidities Index. J Physiother. 2016; 62:171. https://doi.org/10.1016/j.jphys.2016.05.008 PMID:27298055
- 32. Pedregosa F, Varoquaux G, Gramfort A, Michel V, Thirion B, Grisel O, et al. Scikit-learn: Machine learning in Python. The Journal of machine Learning research. 2011; 12:2825–30.

SUPPLEMENTARY MATERIALS

Supplementary Material 1

Formulae to estimate the individual muscle age*

Men

 $MuscleAge = 65.65 + 2.92 \cdot Scaled_TUG - 1.13 \cdot Scaled_Handgrip - 2.26 \cdot Scaled_ASMM$

where:

Scaled measure = (individual measurement – mean of the study sample)/standard deviation of the study sample Scaled $TUG^{\#}$ = (TUG time (s) - 6.04)/1.08 Scaled Handgrip = (Handgrip (kg) - 36.45)/7.15 Scaled ASMM= (ASMM (kg) - 22.6/2.76

Women

MuscleAge = 65.72 - 2.54 · Scaled_TUG - 1.25 · Scaled_Handgrip - 1.21 · Scaled_6MWT

where:

Scaled TUG = (TUG time (s) - 6.69)/1.00 Scaled Handgrip = (Handgrip (kg) - 20.69)/4.17 Scaled 6MWT = (6MWT (m) - 508.18)/74.00

*based on data from 215 moderately active healthy participants (118 women, 97 men; mean age; 66.0 ± 7.3 years) [#] for men's TUG, which data were significantly skewed to the left of the distribution curve (skewness = 2.209), median and interquartile range of the study sample by Ventura et al. were used instead of mean and standard deviation.

Abbreviations

TUG: Timed-Up-and-Go test; ASMM: Appendicular Skeletal Muscle Mass; 6MWT: 6-Minute Walking test.