

The role of phenylalanine and tyrosine in longevity: a cohort and Mendelian randomization study

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ABSTRACT

Background: Protein restriction increases lifespan, however, the specific amino acids affecting lifespan are unclear. Tyrosine and its precursor, phenylalanine, may influence lifespan through their response to low-protein diet, with possible sex disparity.

Methods: We applied cohort study design and Mendelian randomization (MR) analysis. Specifically, we examined the overall and sex-specific relationships between circulating phenylalanine and tyrosine and all-cause mortality in the UK Biobank using Cox regression. To test causality, in two-sample MR analysis, we used genetic variants associated with phenylalanine and tyrosine in UK Biobank with genome-wide significance and uncorrelated ($r^2 < 0.001$) with each other, and applied them to large genome-wide association studies of lifespan, including parental, paternal, and maternal attained ages in the UK Biobank. We also conducted multivariable MR to examine the independent role of phenylalanine and tyrosine.

Results: Tyrosine was associated with shorter lifespan in both observational and MR study, with potential sex disparity. After controlling for phenylalanine using multivariable MR, tyrosine remained related to a shorter lifespan in men (−0.91 years of life, 95% confidence interval (CI) −1.60 to −0.21) but not in women (−0.36 years, 95% CI −0.96 to 0.23). Phenylalanine showed no association with lifespan in either men or women after controlling for tyrosine.

Conclusions: Reducing tyrosine in people with elevated concentrations may contribute to prolonging lifespan, with potential sex-specific differences. It is worthwhile to explore pathways underlying the sex-specific effects.

INTRODUCTION

Enhancing longevity and living a healthy life at older age are key objectives for healthcare systems worldwide. Dietary protein intake regulates longevity across various species [1]. Protein restriction has also been demonstrated to extend lifespan [2]. As proteins are composed of amino acids, we hypothesize that amino acids responding to the effects of protein restriction may affect lifespan. In an animal experiment,

tyrosine has been shown to be specifically involved in regulation of the physiological response to low-protein diet [1]. Another animal experiment further shows that restriction of tyrosine intake lowers internal tyrosine levels, modulates amino acid-sensing pathways, and prolongs lifespan [3]. Tyrosine plays a critical role in metabolic pathways as a precursor to important neurotransmitters like dopamine, norepinephrine, and epinephrine [4]. These neurotransmitters are crucial for regulating mood, cognition, and stress responses [5],

Table 1. The associations of phenylalanine and tyrosine with all-cause mortality in UK Biobank using Cox regression.

Exposure	Sex	HR ¹	95% CI ²	<i>p</i>
Phenylalanine	Overall	1.04	1.03, 1.05	1.1×10^{-9}
	Men	1.04	1.02, 1.05	7.1×10^{-7}
	Women	1.04	1.02, 1.07	1.2×10^{-3}
	Overall	1.02	1.00, 1.03	2.1×10^{-2}
Tyrosine	Men	1.03	1.01, 1.05	5.5×10^{-3}
	Women	1.00	0.98, 1.03	7.2×10^{-1}

¹HR: hazard ratio. In overall analysis we adjusted for age, body mass index (BMI), Townsend Deprivation Index, smoking status, alcohol consumption, physical activity, ethnicity, and education (in years). For combined-sex analyses, we additionally adjusted for sex. ²CI: confidence interval.

which are vital for metabolic health and potentially influencing lifespan [6]. Tyrosine deprivation may also lead to suppression of IIS and mTORC1 pathways in peripheral tissues, potentially suppress organismal aging [3]. Phenylalanine is the precursor of tyrosine; specifically, tyrosine is formed through the conversion of phenylalanine mediated by phenylalanine hydroxylase (PAH). Therefore, we also examined the role of phenylalanine. Elevated circulating phenylalanine has been associated with telomere loss [7], inflammatory disease [8], and type 2 diabetes [9]. Experimental evidence shows that phenylalanine can undergo oxidation to form toxic metabolite meta-tyrosine (m-tyrosine), which has been shown to shorten *C. elegans* lifespan [10, 11]. However, the role of phenylalanine and tyrosine in humans has been rarely examined.

Interestingly, lifespan differs by sex. In most regions worldwide, men have a consistently shorter life expectancy compared with women [12], and the disparity may have widened after the COVID-19 pandemic [13]. With US life expectancy declining from 78.8 years in 2019 to 77.0 in 2020 and 76.1 in 2021, the lifespan difference between men and women expanded to 5.8 years, marking the widest gap since 1996 [13]. Notably, tyrosine also differs by sex, with lower levels in young women than in young men [14]. Whether tyrosine explains or partly explains the sex difference in lifespan has not been clarified. In this study, we assessed the associations of tyrosine and its precursor phenylalanine with lifespan in overall people and in men and women separately, using UK Biobank, a large cohort in UK. Since conventional observational designs are inherently susceptible to residual confounding arising from variables such as socioeconomic factors and health status, we also used Mendelian randomization (MR) (Supplementary Figure 1). Using genetic variants as instruments, which are less affected by socioeconomic positions [15], MR has the

potential to mitigate confounding. Here we employed MR to assess the role of tyrosine and phenylalanine in lifespan overall and sex-specifically.

RESULTS

In the cohort study, 272,475 participants with death status information, measurement of amino acids, and information on confounders were included in the analysis. Among these, 125,359 were men. Of these 272,475 participants, 23,964 deaths were identified from death records, including 14,230 in men and 9,734 in women. After adjustment for multiple confounders (details shown in Methods), plasma phenylalanine was linked to elevated all-cause mortality overall (Hazard ratio (HR) 1.04 per SD increase in phenylalanine, 95% confidence interval (CI) 1.03–1.05), in men (HR 1.04, 95% CI 1.02–1.05) and in women (HR 1.04, 95% CI 1.02–1.07). The findings were similar for both men and women. Plasma tyrosine was associated with a higher risk of all-cause mortality overall and in men (HR 1.03, 95% CI 1.01–1.05), but not in women (HR 1.00, 95% CI 0.98–1.03) (Table 1), although the difference in the associations in men and women was not statistically significant ($p = 0.16$).

The associations of phenylalanine and tyrosine with lifespan, both overall and stratified by sex, remained after excluding deaths from accidents (Supplementary Table 1). The Pearson correlation coefficient between phenylalanine and tyrosine was 0.52 ($p < 0.01$). A greater tyrosine-to-phenylalanine ratio was linked to a lower overall risk of all-cause mortality in overall people (HR 0.98, 95% CI 0.97–1.00) and also in women (HR 0.96, 95% CI 0.94–0.99), whereas no association was observed in men (HR 1.00, 95% CI 0.98–1.02). Restricted cubic spline analysis suggested non-linearity, with the turning point at the standardized concentration of around 0 for both amino acids (p -value < 0.05 , Supplementary Figures 2, 3 and Supplementary Table

2). In disease-specific mortality, we found positive associations of phenylalanine with both cardiovascular disease (CVD) mortality (HR 1.03, 95% CI 1.00–1.06) and cancer mortality (HR 1.04, 95% CI 1.02–1.05), whereas tyrosine was not associated with either outcome (Supplementary Table 3). These observations imply that phenylalanine could participate in pathways relevant to cardiovascular health and carcinogenesis.

In the genome-wide association study (GWAS) of two amino acids, the heritability for phenylalanine and tyrosine was 0.04 and 0.09, respectively (Supplementary Table 4). The LDSC intercepts and attenuation ratio indicated no genomic inflation of test statistics due to confounding factors (Supplementary Table 4). The Manhattan plots were shown in Figures 1–4, Quantile-Quantile plots were presented in

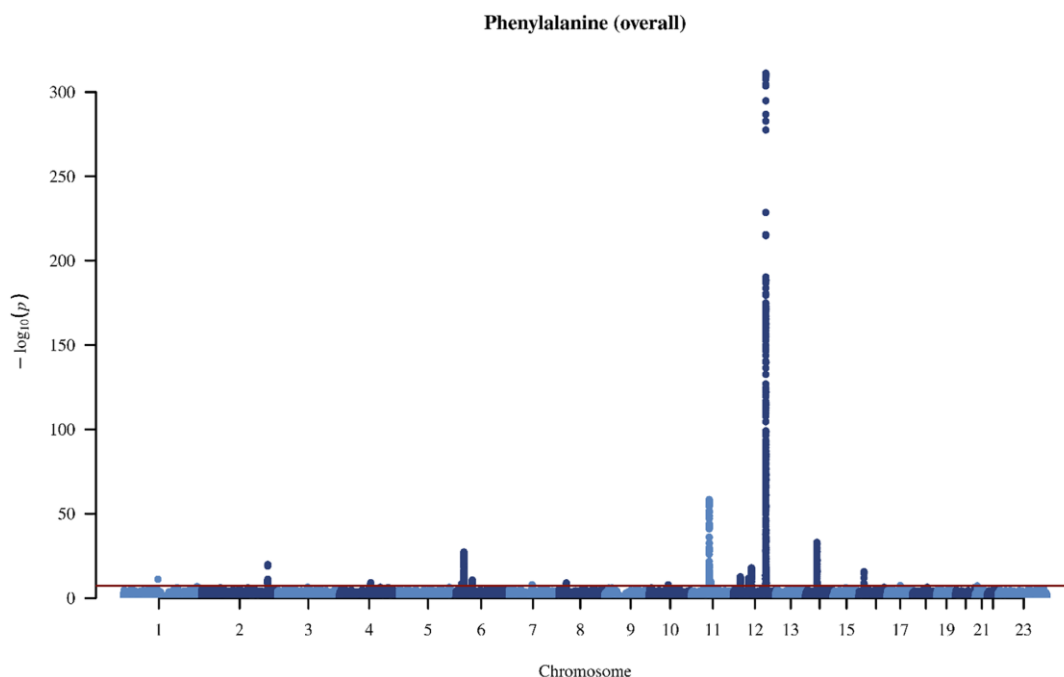


Figure 1. Manhattan plot on the genome-wide association study of phenylalanine in overall people.

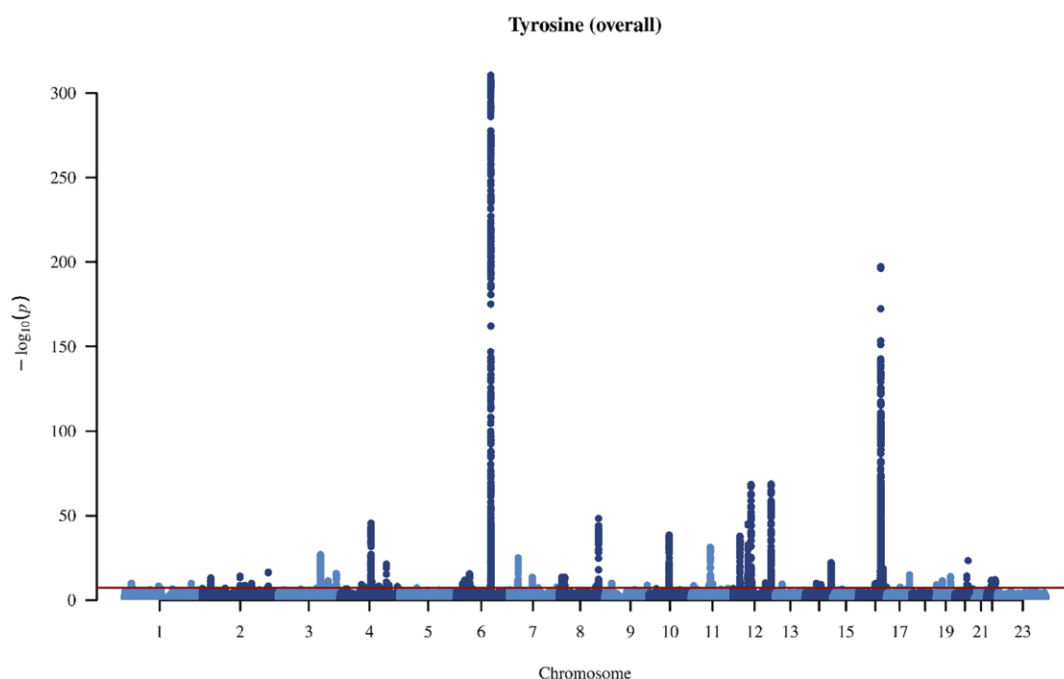


Figure 2. Manhattan plot on the genome-wide association study of tyrosine in overall people.

Supplementary Figure 4. In the overall analysis, we identified 2,422 genetic variants with genome-wide significance for phenylalanine, and 11,379 for tyrosine. In sex-specific GWAS, we identified 1,099 genetic variants for phenylalanine in men and 946 in women, while for tyrosine, 5,297 variants reached genome-wide significance in men and 4,840 in women.

After removing correlated genetic variants, we used 21 and 74 single nucleotide polymorphisms (SNP) as genetic instruments for phenylalanine and tyrosine, respectively in the overall analysis (Supplementary Tables 5 and 6). We used 12 SNPs in men and 10 SNPs in women for phenylalanine, and 45 SNPs in men and 29 SNPs in women for tyrosine (Supplementary Tables 7 and 8). The SNPs associated with phenylalanine and tyrosine are located within genes critical for amino

acid metabolism, transport, and regulation. For phenylalanine, essential genes include *PAH*, which catalyzes phenylalanine's conversion to tyrosine; members of the solute carrier (*SLC*) transporter family (*SLC17A1*, *SLC38A4*, and *SLC43A1*), which facilitate cellular uptake and distribution of amino acids; and carbamoyl-phosphate synthase 1 (*CPS1*), an essential enzyme in the urea cycle linking nitrogen metabolism with amino acid catabolism. Additionally, genes in the glutathione S-transferase (*GST*) family, including *GSTM1* and *GSTA2*, encode enzymes central to detoxification pathways by conjugating amino acid-derived metabolites. For tyrosine, SNPs further involve the previously highlighted genes *PAH*, *CPS1* and *GSTM1*, alongside *HPD* encoding 4-hydroxyphenylpyruvate dioxygenase, which participates in tyrosine breakdown through homogentisate formation.

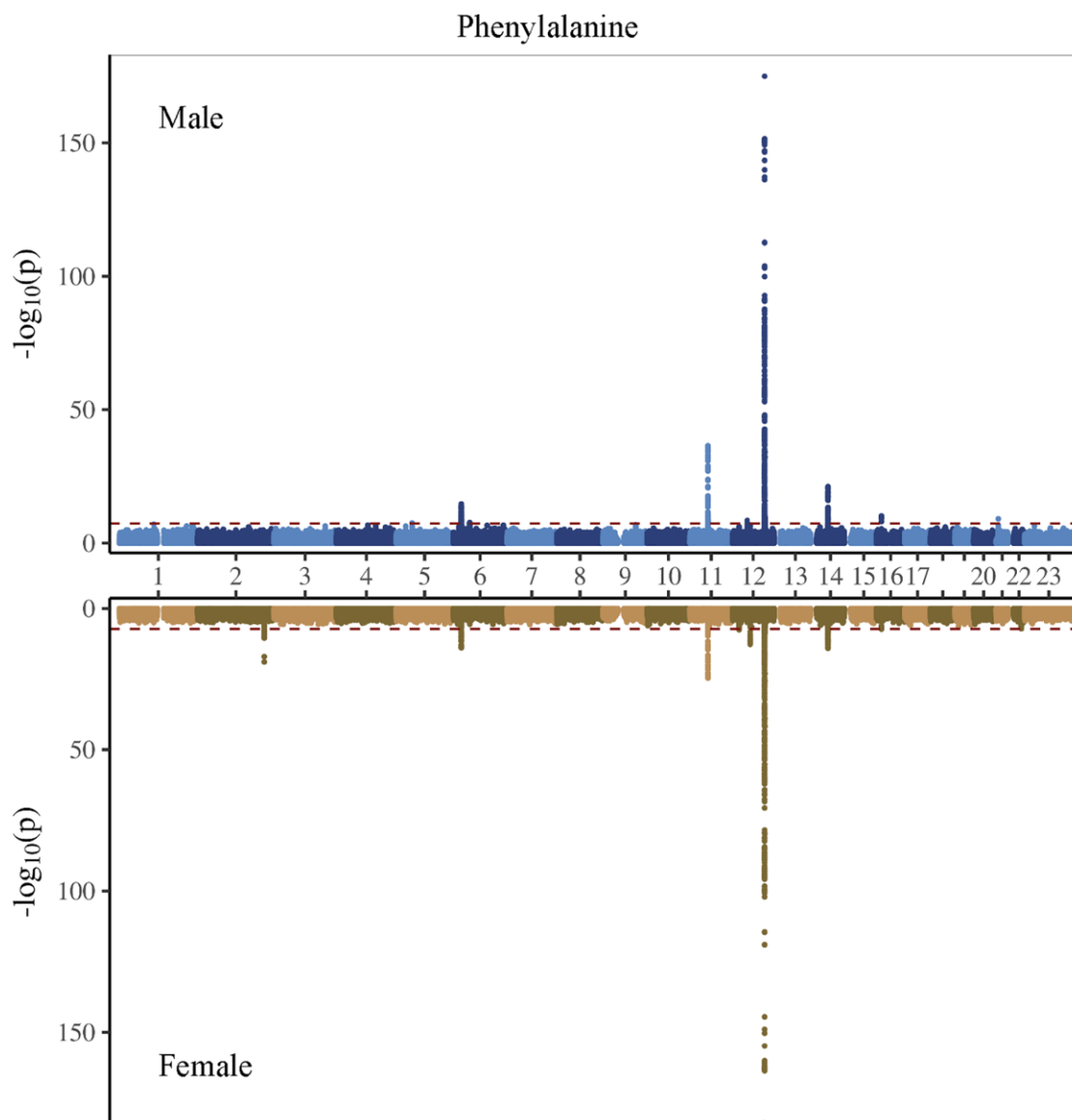


Figure 3. Manhattan plot on the genome-wide association study of phenylalanine in men and women.

Using two-sample MR, we estimated the effect on lifespan, i.e., years of life. Genetically predicted higher phenylalanine was related to longer lifespan in men but not related to lifespan in overall analysis or in women (Figure 5). The association in men showed consistent directions of associations applying various analytic methods (Figure 6). Genetically mimicked higher tyrosine levels were linked to a shorter lifespan in the overall population and in both sexes using inverse variance weighting (IVW) (Figure 5). The associations were also shown when we used Mendelian randomization pleiotropy residual sum and outlier (MR-PRESSO) (Figure 6), after excluding outliers (Supplementary Table 9 and Figure 6). The associations in weighted median and weighted mode showed aligned directions of association, but the CI included the null

(Figure 6). The associations persisted after the exclusion of SNPs with potential pleiotropy (Supplementary Table 10). Scatter plot and leave-one-out plot provided no indication that the relationships were affected by any individual SNP (Supplementary Figures 5 and 6). Sensitivity analysis using genetic instruments from another GWAS in overall people without UK Biobank participants showed consistent directions of associations (Supplementary Table 11). Genetically predicted phenylalanine was linked to longer lifespan in men, whereas no relationship was observed in the overall people or among women. Genetically predicted tyrosine had an inverse association with lifespan overall using MR-PRESSO and had the direction of inverse association in men and women especially using MR-PRESSO (Supplementary Table 11). Power calculation

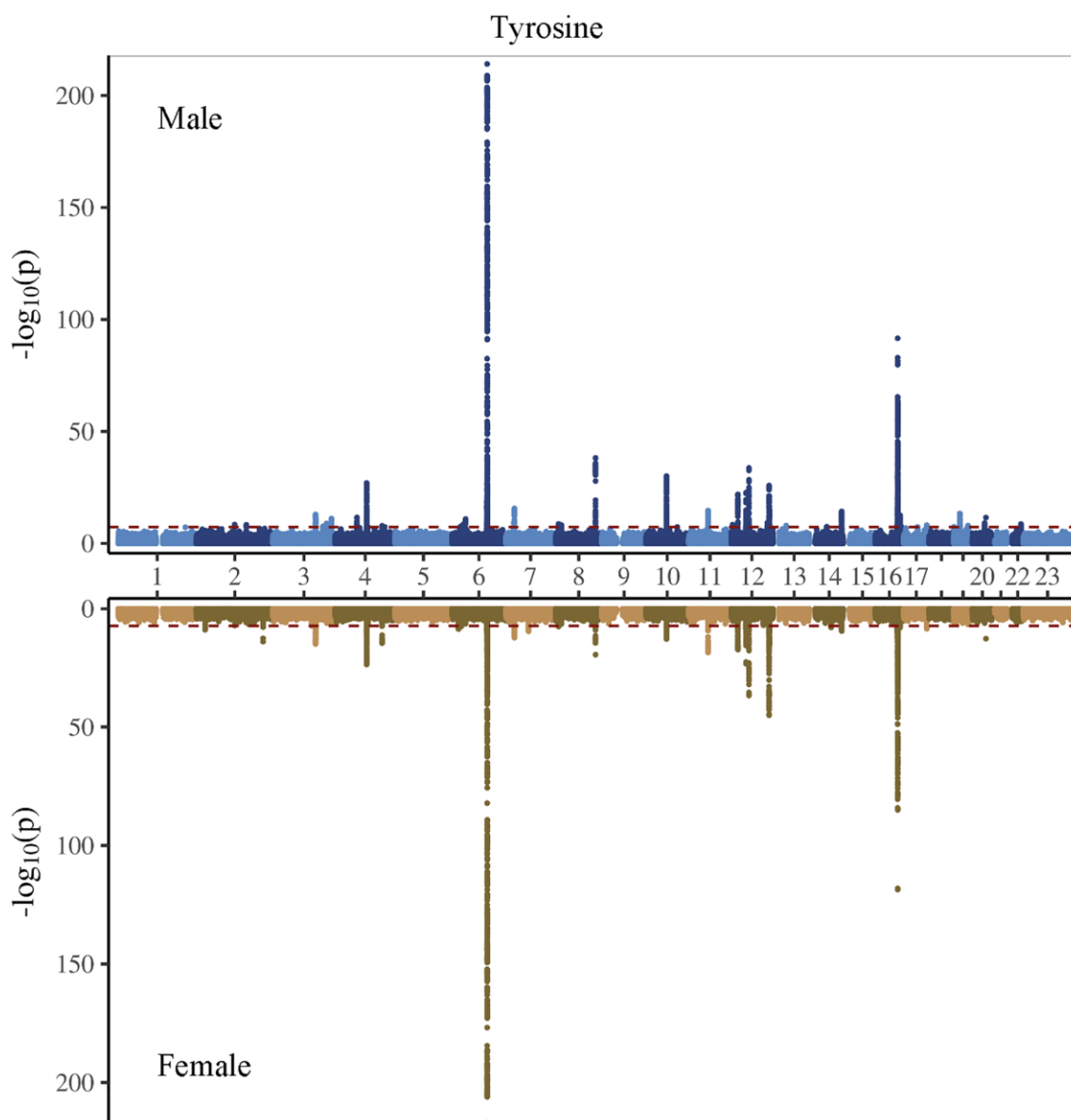


Figure 4. Manhattan plot on the genome-wide association study of tyrosine in men and women.

showed that at 80% statistical power, we can identify an effect size of ~1.6 life years for phenylalanine and 1.0 life years for tyrosine (Supplementary Table 12).

In multivariable MR study including both amino acids, we found that after controlling for tyrosine, phenylalanine was not related to lifespan. Interestingly, after controlling for phenylalanine, tyrosine was associated with shorter lifespan in men, while no clear relationship in women (Table 2). The positive association was shown in both IVW and MR Egger in men, but not shown in MR Egger in women (Table 2). MR Egger results provided no indication of directional pleiotropy (intercept $p > 0.05$).

DISCUSSION

Our novel finding contributes to the scarce epidemiological evidence regarding the role of tyrosine and phenylalanine in lifespan. Our study showed that tyrosine was associated with shorter lifespan in observational and MR studies. The association was

independent of phenylalanine, which remained in multivariable MR after controlling for phenylalanine. The role of tyrosine may be sex-specific, with a clearer effect in men than in women. Phenylalanine was not related to lifespan after controlling for tyrosine.

Based on our results, targeting tyrosine may be a potential strategy for improving lifespan. Partly consistent with our findings, animal experiment suggests that restricting dietary protein in rats extends lifespan while lowering tyrosine concentrations in liver and muscle [16]. The biological processes linking tyrosine to lifespan have not been thoroughly determined. Tyrosine was associated with insulin resistance [17]. According to evolutionary biology, more investment in growth and reproduction often comes at the expense of lifespan [18], while insulin acts as one of key regulators of growth and reproduction [19, 20]. Consistently, insulin resistance has been shown to be related to multiple diseases and decreased lifespan [21]. Insulin resistance may also have sex-specific effects [22, 23]. Restricting caloric intake, known to

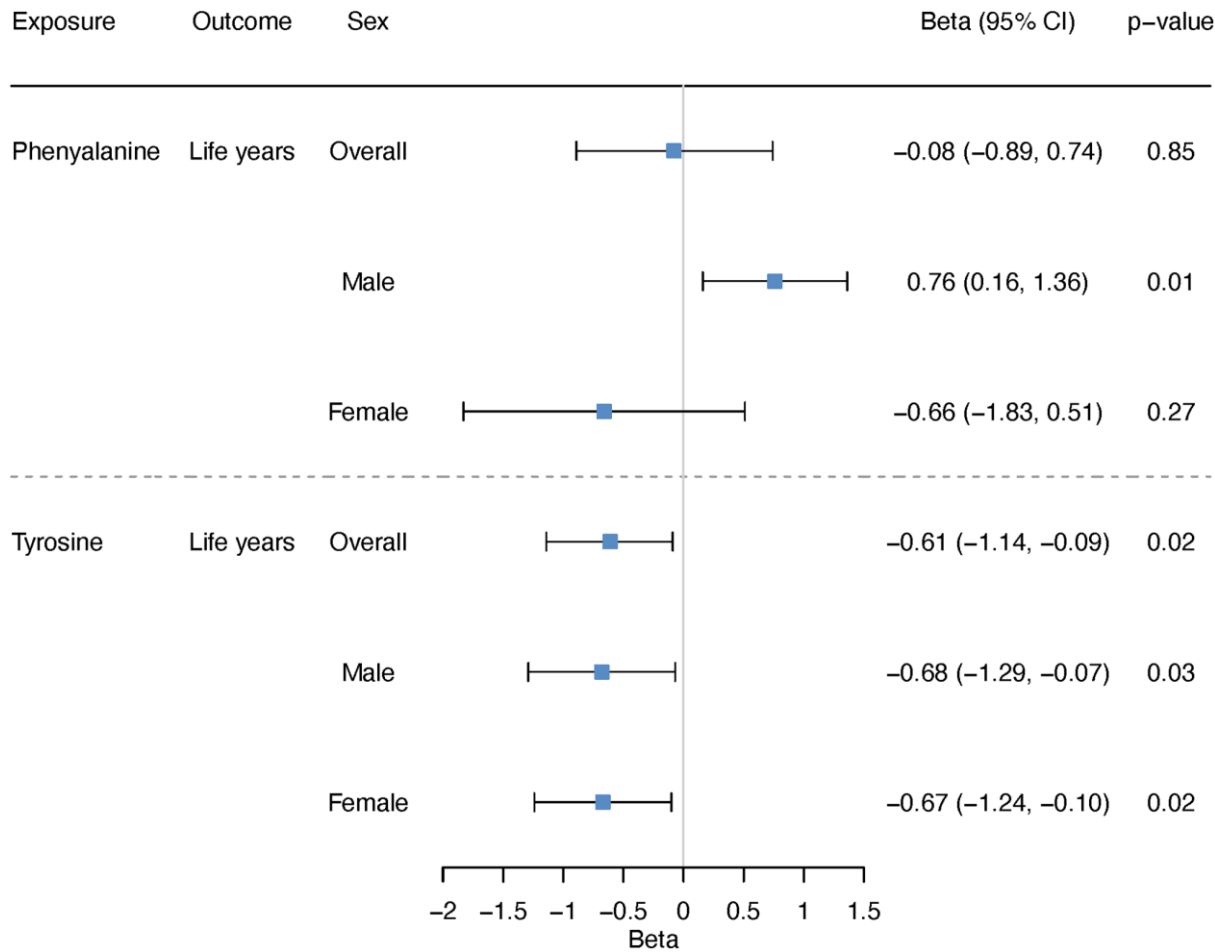


Figure 5. Overall and sex-specific associations of phenylalanine and tyrosine with lifespan using inverse variance weighting. We presented increased/decreased life years for ease of understanding; these estimates were calculated based on the log hazard ratios reported by the lifespan GWAS (detailed described in “Methods-Genetic associations with lifespan”).

reduce the risk of insulin resistance [24], also prolongs lifespan in a sex-specific way [25]. Insulin resistance may interact with sex hormones, and testosterone has been shown to be related to survival, with a more obvious effect in men than in women [26]. Meanwhile, tyrosine acts as a precursor for neurotransmitters such as dopamine, norepinephrine, and epinephrine [4], which are crucial for regulating mood, cognition, and stress responses [5] and potentially influencing lifespan [6]. Interestingly, these neurotransmitters are regulated by sex hormones [27, 28], which provides another explanation for the sex-specific associations.

Our study, for the first time, evaluated the role of tyrosine and phenylalanine in lifespan using both conventional observational study and MR. In this novel study, we also examined the sex difference in the associations, and suggested potential sex disparity in the role of tyrosine. Despite of the novelty, our study bears some limitations. First, traditional observational

study, including our study, is inevitably susceptible to residual confounding. Some confounders, such as socioeconomic position, is difficult to be accurately measured [29]. In contrast, MR study can minimize confounding by leveraging genetic variants that are randomly assigned at conception [30]. This may partly explain the inconsistent associations for phenylalanine in observational study and MR study. Second, MR required stringent assumptions: relevance, independence, and exclusion-restriction. Accordingly, we selected genetic instruments with strong associations with these amino acids. In addition, we tested the associations of these genetic instruments with potential confounders. Considering that phenylalanine and tyrosine have shared SNPs, including rs140584594, rs10750864 and rs1043011, we also conducted multi-variable MR to examine the independent role of phenylalanine and tyrosine. Third, genetic associations with the amino acids and with lifespan are both from UK Biobank, the sample overlap could introduce bias

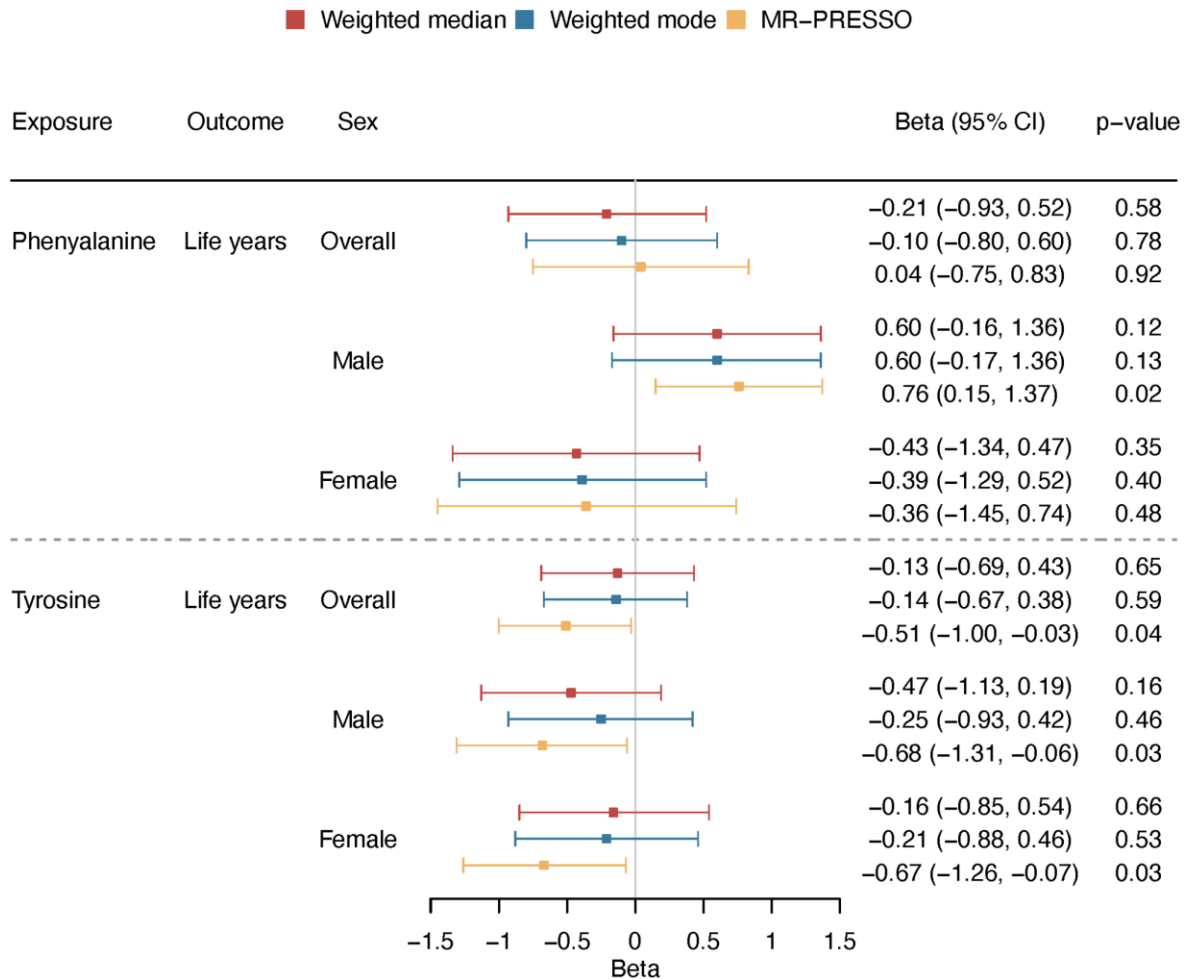


Figure 6. Overall and sex-specific associations of phenylalanine and tyrosine with lifespan using different analytic methods (weighted median, weighted mode and MR-PRESSO). We presented increased/decreased life years for ease of understanding; these estimates were calculated based on the log hazard ratios reported by the lifespan GWAS (detailed described in “Methods-Genetic associations with lifespan”).

Table 2. The sex-specific associations of genetically predicted phenylalanine and tyrosine with lifespan using multivariable MR, including inverse variance weighting (IVW) and MR-Egger.

Sex	Methods	Exposure	Life years	95% CI	<i>p</i>	MR-Egger intercept <i>p</i>
Men	IVW	phenylalanine	0.75	−0.07, 1.58	0.08	0.61
		tyrosine	−0.80	−1.40, −0.23	0.006	
	MR-Egger	phenylalanine	0.55	−0.59, 1.69	0.35	
		tyrosine	−0.91	−1.60, −0.21	0.01	
Women	IVW	phenylalanine	−0.72	−1.47, 0.05	0.07	0.15
		tyrosine	−0.59	−1.11, −0.05	0.03	
	MR-Egger	phenylalanine	−0.18	−1.24, 0.85	0.73	
		tyrosine	−0.36	−0.96, 0.23	0.24	

into the MR estimates. However, our sensitivity analysis leveraging genetic variants derived from GWAS conducted in combined-sex populations outside the UK Biobank showed consistent directions of associations. It would be ideal to replicate using sex-specific GWAS not conducted in UK Biobank, but such GWAS was not available yet. Moreover, a recent study suggested that MR analyses using overlapped samples in large cohorts like UK Biobank can still provide valid estimates [31]. Fourth, the study may lack adequate power to identify sex difference in the role of tyrosine, which may explain the marginal significance in the testing for sex disparity on the associations of tyrosine. Given the consistent trends observed in both observational and multivariable MR studies, it is more plausible that the marginal significance reflects limited statistical power, not an actual lack of effect. This is further supported by power calculations. Therefore, replicating the study in larger cohorts would be worthwhile. Fifth, MR study assessed the role of endogenous exposures, which is different from nutrient supplementation. While blood levels of amino acids respond to nutrient supplementation or diet rich in these amino acids [32–34], our findings on circulating tyrosine or phenylalanine may not directly reflect the role of dietary consumption of these amino acids. Sixth, these amino acids were only measured at a single time point. Future investigations with repeated measures would be valuable to further elucidate how circulating phenylalanine and tyrosine levels fluctuate over time and to clarify their influence on mortality outcomes. Seventh, our findings need to be interpreted with caution. Given the potential non-linearity, the positive associations with mortality are more applicable to people with higher levels of phenylalanine or tyrosine. Replicating these results in populations with different levels of amino acids would be worthwhile. Finally, MR study examined the lifelong effect of phenylalanine and tyrosine, which is not comparable to randomized controlled trials assessing short-term effects of supplementation.

From the perspective of etiology, our study suggests that tyrosine is involved in longevity. More mechanistic studies will be worthwhile to assess the possible pathways. The circulating level of tyrosine is modifiable. In terms of public health interventions, our findings indicate that nutrients or diets, such as protein-restriction diet, which lower tyrosine will be helpful for prolonging lifespan. Tyrosine is also a popular nutrient supplement, promoted as a neurotransmitter support for a positive mood and mental alertness. Our study is not directly related to tyrosine supplement, but given tyrosine supplement may increase blood tyrosine, our study did not support the benefit of long-term use of tyrosine on lifespan.

METHODS

Study design

To understand the role of phenylalanine and tyrosine in longevity, we used conventional observational study to examine their relationships with all-cause mortality in the UK Biobank. To minimize confounding, we applied univariable MR to assess the associations of genetically predicted phenylalanine and tyrosine with parental attained age. Given that phenylalanine and tyrosine are correlated, we further performed multivariable MR to examine their independent effects. To assess the sex-specific roles, we conducted sex-stratified analyses in both observational and MR studies. The study design was shown in the flow diagram in Supplementary Figure 1.

Cohort study

UK Biobank is a large-scale cohort study, with a current median follow up of 11.1 years [35]. Between 2006 and 2010, it enrolled 502,713 individuals aged 40–69 years, with a mean age of 56.5 years in England, Scotland and Wales. Among all participants, 45.6% are men and 94% were identified as of European ancestry by self-report.

Utilizing data from UK Biobank, we studied the associations of baseline plasma levels of phenylalanine and tyrosine with all-cause mortality using Cox regression, controlling for age, sex (in the overall analysis but not in sex-specific analysis), Townsend index, smoking habits, alcohol intake, physical activity, self-reported ethnicity (white, black, Asian, and other), education (years) and body mass index (BMI). Deaths were identified by death records. We also conducted sensitivity analysis excluding deaths from accidents (V00–Y99). To assess the potential nonlinear associations, we used restricted cubic splines [36]. We also examined the correlation between phenylalanine and tyrosine and the association of tyrosine-to-phenylalanine ratio with all-cause mortality. We set the censoring date to 19 Dec 2022, which is the latest date of death in the records. In addition to examining all-cause mortality, we also investigated disease-specific mortality outcomes based on the International Classification of Diseases (ICD-10) codes. Specifically, we assessed the associations with CVD mortality (I00–I99) and cancer mortality (C00–D48), the top two leading contributors to mortality in UK Biobank. Plasma levels of phenylalanine and tyrosine were quantified in absolute concentrations (mmol/L), measured in a high-throughput NMR-based metabolic biomarker profiling platform (Nightingale Health Ltd.). Procedures for sample preparation, spectrometer calibration, and quality-control protocols are detailed in previous publications [35, 37, 38]. All measures were standardized before analyses.

MR study

Overall and sex-specific GWAS of phenylalanine and tyrosine

We conducted a GWAS of the plasma levels of phenylalanine and tyrosine in the UK Biobank utilizing fastGWA tool (GCTA toolbox, version 1.94.1) [39]. In the mixed linear model association analyses, we utilized a sparse genetic relationship matrix with a cutoff value of 0.05, which was computed from linkage disequilibrium (LD)-pruned HapMap 3 SNP set. The LD-pruning parameters set in PLINK included a window of 1,000 variants, step size of 100, r^2 threshold of 0.9, and minor allele frequency exceeding 0.01 [40]. For our genome-wide association analyses, we excluded SNPs with an imputation score below 0.3, minor allele frequency under 0.1%, missing genotype rates exceeding 5% per individual, missing genotype rates over 5% per genetic variant, or p -value of Hardy-Weinberg equilibrium less than 1×10^{-8} . In the non-pseudoautosomal X chromosome region, males were coded as 0 or 2 copies of the effect allele. Participants of European ancestry were characterized in the Pan-ancestry genetic analysis of the UK Biobank (Pan-UK

Biobank) [41]. Additionally, participants were not included in the analysis if they had withdrawn consent, displayed discrepancies between self-reported and genetic sex, exhibited sex chromosome aneuploidy, were identified as heterogeneity outliers or missing genotype rate. After quality control, we performed both combined and sex-stratified GWAS of phenylalanine and tyrosine. In the sex-specific GWAS, age and 10 genetic principal components supplied by the Pan-UK Biobank were included as covariates, while sex was added as an additional covariate in the combined-sex GWAS. We applied the rank-based inverse normal transformation to phenylalanine and tyrosine measurements to enable interpretation per one standard deviation (SD) change [42]. We computed the SNP-based heritability and checked for inflation by LD score regression [43].

Genetic instruments for phenylalanine and tyrosine

Genetic proxies for circulating phenylalanine and tyrosine were obtained based on the GWAS we conducted in the UK Biobank. Specifically, we selected SNPs linked to circulating phenylalanine or tyrosine reaching genome-wide significance (5×10^{-8}) and meeting an LD cutoff of $r^2 < 0.001$. The instruments for overall analysis were based on GWAS in the overall sample, whilst the genetic instruments for sex-specific analyses were derived from the corresponding sex-specific GWAS. To ensure the validity of the genetic variants, we verified that the F-statistic exceeded 10 [44], with the F-statistic derived from a commonly used formula [45]. The selected genetic instruments were presented in Supplementary Tables 5–8. To understand the potential pleiotropy, we examined whether these selected SNPs were associated with potential confounders for the association between phenylalanine or tyrosine and all-cause mortality, such as Townsend index, education, smoking status, alcohol consumption and physical activity in the UK Biobank. SNPs showing genome-wide significant associations with any of these factors were excluded in sensitivity analysis, as shown in Supplementary Tables 13 and 14.

Genetic associations with lifespan

Lifespan was used as the outcome. We retrieved genetic associations for parental attained age (age at death or current age) from a large-scale GWAS involving 389,166 UK Biobank participants of European ancestry [46]. Utilizing parental lifespan is a common way in GWAS of longevity [46], as longevity is heritable [47], so parental lifespan can provide a proxy measure for offsprings' lifespan, and it can be used even when participants are still alive. The combined parental attained age was calculated by adding the z-standardized maternal and paternal

attained ages [46]. The GWAS controlled for age, sex, and the first five principal components [46]. Genetic associations for paternal and maternal attained age were obtained from sex-specific GWAS of parental longevity in participants of European descent from the UK Biobank (fathers: $n = 415,311$; mothers: $n = 412,937$) [46]. Employing sex-stratified Cox proportional hazards model, the GWAS estimated the effect of offspring genetic variant on parental survival, adjusting for age as well as 10 principal components of ancestry. To enhance interpretability, GWAS summary statistics (log hazard ratio) were transformed into years of life through sign inversion and multiplication by 10 [46, 48]. Considering the effect sizes derived from offspring genetic data represent half the true parental variant effect, we doubled the log hazard ratios in the overall analyses [46], and multiplied by 2.2869 for fathers and 2.5863 for mothers, respectively, in the sex-specific analyses [46, 49], as previously described [50].

Statistical analysis

In the univariable MR, SNP-specific estimates were derived from Wald ratios, which were calculated as the genetic association with parental attained age divided by the association with phenylalanine or tyrosine. These ratios were integrated via IVW with multiplicative random effects [51]. For the sex-stratified analysis, we utilized the genetic associations from sex-specific GWAS of lifespan and phenylalanine or tyrosine. The MR estimates were presented as life years per SD increase in phenylalanine or tyrosine. Multiple comparisons were accounted for using a false discovery rate (FDR) threshold of less than 0.05. Associations showing nominal significance ($p < 0.05$) that failed multiple testing correction were defined as suggestive associations. To assess whether the sex difference has statistical significance, we performed the heterogeneity test with the “meta” package in R.

To address possible pleiotropy, as previously [52–55], we applied multiple analytic approaches robust to pleiotropy, such as the weighted median, weighted mode, MR-PRESSO and MR-Egger methods. The weighted median approach offers a reliable estimate of the causal effect even if as much as half of the information comes from SNPs that invalid instruments [56]. Weighted mode assumes that the largest group of are valid, that is, no larger group of invalid instruments providing the same causal estimate exists [57]. MR-PRESSO detects and removes outlier SNP(s) that disproportionately influenced associations [58], and gives the corrected estimates after the removal of the outliers. MR-Egger can determine if genetic variants exhibit directional pleiotropy, that is, whether their

pleiotropic effects on the outcome deviate from zero on average, as indicated by a non-zero intercept, and it also provides a corrected estimate [59]. However, this approach usually gives wider confidence intervals compared to other methods [60]. Considering the overlapping in samples of GWAS for exposure and outcomes, we additionally performed a sensitivity analysis utilizing SNPs for phenylalanine and tyrosine derived from a GWAS which does not include participants from UK Biobank [61], but only overall GWAS is available.

Multivariable MR

In addition to univariable MR analyses, we conducted multivariable MR, which leverages pleiotropic SNPs associated with more than one exposure, to assess the causal effects of individual exposure adjusting for other exposure(s) [62]. We included genetic instruments for both phenylalanine and tyrosine, to examine the independent effect of phenylalanine and tyrosine. The genetic instruments for each amino acid were as used in univariable MR. After integrating the SNPs for both amino acids, we removed overlapping and correlated ($r^2 > 0.05$) SNPs, and the remaining genetic variants were utilized for the multivariable MR analysis. We employed multivariable MR-Egger analysis to detect directional pleiotropy, and when it was identified, we adopted the multivariable MR-Egger estimates as the main analysis results [63].

Power calculation

In power calculation, the required sample size for MR studies is roughly the conventional observational study sample size divided by the proportion of variance in the exposure explained by the genetic instruments [64]. Variance explained by individual SNP was computed via $\beta^2 \times 2 \times (\text{EAF}) \times (1 - \text{EAF})$, with β as the effect allele's standardized beta coefficient, EAF as its frequency [65]. For lifespan, we first calculated the detected effect size at current sample size (i.e., log odds ratio) based on case/non-case ratio, total sample size, and the variance explained by SNPs [66], and then converted to life years using the same way as we did in the statistical analysis.

All statistical analyses were performed using the R packages “TwoSampleMR”, “MendelianRandomization”, “MRPRESSO” and “meta” (R version 4.0.1, R Foundation for Statistical Computing, Vienna, Austria).

Availability of data and materials

The dataset supporting the conclusions of this article is available upon request and approval by the UK Biobank (<https://www.ukbiobank.ac.uk/enable-your-research/apply-for-access>).

AUTHOR CONTRIBUTIONS

JVZ conceived the idea. JVZ designed the study with the help of KY. YS conducted the genome-wide association studies and Cox regression with the help of KY, JVZ and JZ conducted the Mendelian randomization study. JVZ interpreted the results with the help of KY. JVZ drafted the manuscript, YS, KY and JZ provided important input on improvement. All authors read the final version and approved the submission.

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CONFLICTS OF INTEREST

The authors declare no conflicts of interest related to this study.

ETHICAL STATEMENT AND CONSENT

This research has been conducted using the UK Biobank Resource under Application Number 48818. This work uses data provided by patients and collected by the NHS as part of their care and support. These data are copyrighted, 2022, NHS England. Reused with the permission of the NHS England and UK Biobank. All rights reserved. This research used data assets made available by National Safe Haven as part of the Data and Connectivity National Core Study, led by Health Data Research UK in partnership with the Office for National Statistics and funded by UK Research and Innovation (grant ref MC_PC_20058). Informed consent was obtained from all subjects involved in the study.

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REFERENCES

1. Kosakamoto H, Okamoto N, Aikawa H, Sugiura Y, Suematsu M, Niwa R, Miura M, Obata F. Sensing of the non-essential amino acid tyrosine governs the response to protein restriction in *Drosophila*. *Nat Metab*. 2022; 4:944–59.
<https://doi.org/10.1038/s42255-022-00608-7>
PMID:35879463
2. Kitada M, Ogura Y, Monno I, Koya D. The impact of dietary protein intake on longevity and metabolic health. *EBioMedicine*. 2019; 43:632–40.
<https://doi.org/10.1016/j.ebiom.2019.04.005>
PMID:30975545
3. Kosakamoto H, Sakuma C, Okada R, Miura M, Obata F. Context-dependent impact of the dietary non-essential amino acid tyrosine on *Drosophila* physiology and longevity. *Sci Adv*. 2024; 10:eadn7167.
<https://doi.org/10.1126/sciadv.adn7167>
PMID:39213345
4. Fernstrom JD, Fernstrom MH. Tyrosine, phenylalanine, and catecholamine synthesis and function in the brain. *J Nutr*. 2007 (Suppl 1); 137:1539S–47.
<https://doi.org/10.1093/jn/137.6.1539S>
PMID:17513421
5. Carroll AJ, Bogucki OE. Chapter 24 - Depression and biomarkers of cardiovascular disease. *The Neuroscience of Depression*. Academic Press. 2021; 239–49.
<https://doi.org/10.1016/B978-0-12-817933-8.00018-9>
6. Gilman SE, Sucha E, Kingsbury M, Horton NJ, Murphy JM, Colman I. Depression and mortality in a longitudinal study: 1952–2011. *CMAJ*. 2017; 189:E1304–10.
<https://doi.org/10.1503/cmaj.170125>
PMID:29061855
7. Eriksson JG, Guzzardi MA, Iozzo P, Kajantie E, Kautiainen H, Salonen MK. Higher serum phenylalanine concentration is associated with more rapid telomere shortening in men. *Am J Clin Nutr*. 2017; 105:144–50.
<https://doi.org/10.3945/ajcn.116.130468>
PMID:27881392
8. Neurauter G, Schröcksnadel K, Scholl-Bürgi S, Sperner-Unterwieser B, Schubert C, Ledochowski M, Fuchs D. Chronic immune stimulation correlates with reduced phenylalanine turnover. *Curr Drug Metab*. 2008; 9:622–7.
<https://doi.org/10.2174/138920008785821738>
PMID:18781914
9. Guasch-Ferré M, Hruby A, Toledo E, Clish CB, Martínez-González MA, Salas-Salvadó J, Hu FB. Metabolomics in Prediabetes and Diabetes: A

Systematic Review and Meta-analysis. *Diabetes Care*. 2016; 39:833–46.

<https://doi.org/10.2337/dc15-2251>

PMID:27208380

10. Dato S, Hoxha E, Crocco P, Iannone F, Passarino G, Rose G. Amino acids and amino acid sensing: implication for aging and diseases. *Biogerontology*. 2019; 20:17–31.
<https://doi.org/10.1007/s10522-018-9770-8>
PMID:30255223
11. Edwards C, Canfield J, Copes N, Brito A, Rehan M, Lipps D, Brunquell J, Westerheide SD, Bradshaw PC. Mechanisms of amino acid-mediated lifespan extension in *Caenorhabditis elegans*. *BMC Genet*. 2015; 16:8.
<https://doi.org/10.1186/s12863-015-0167-2>
PMID:25643626
12. Hossin MZ. The male disadvantage in life expectancy: can we close the gender gap? *Int Health*. 2021; 13:482–4.
<https://doi.org/10.1093/inthealth/ihaa106>
PMID:33533409
13. Yan BW, Arias E, Geller AC, Miller DR, Kochanek KD, Koh HK. Widening Gender Gap in Life Expectancy in the US, 2010–2021. *JAMA Intern Med*. 2024; 184:108–10.
<https://doi.org/10.1001/jamainternmed.2023.6041>
PMID:37955927
14. Guevara-Cruz M, Vargas-Morales JM, Méndez-García AL, López-Barradas AM, Granados-Portillo O, Ordaz-Nava G, Rocha-Viggiano AK, Gutierrez-Leyte CA, Medina-Cerda E, Rosado JL, Morales JC, Torres N, Tovar AR, Noriega LG. Amino acid profiles of young adults differ by sex, body mass index and insulin resistance. *Nutr Metab Cardiovasc Dis*. 2018; 28:393–401.
<https://doi.org/10.1016/j.numecd.2018.01.001>
PMID:29422298
15. Lawlor DA, Harbord RM, Sterne JA, Timpson N, Davey Smith G. Mendelian randomization: using genes as instruments for making causal inferences in epidemiology. *Stat Med*. 2008; 27:1133–63.
<https://doi.org/10.1002/sim.3034>
PMID:17886233
16. Kalhan SC, Uppal SO, Moorman JL, Bennett C, Gruca LL, Parimi PS, Dasarathy S, Serre D, Hanson RW. Metabolic and genomic response to dietary isocaloric protein restriction in the rat. *J Biol Chem*. 2011; 286:5266–77.
<https://doi.org/10.1074/jbc.M110.185991>
PMID:21147771
17. Chen S, Miki T, Fukunaga A, Eguchi M, Kochi T, Nanri A, Kabe I, Mizoue T. Associations of serum amino acids with insulin resistance among people with and without overweight or obesity: A prospective study in Japan. *Clin Nutr*. 2022; 41:1827–33.
<https://doi.org/10.1016/j.clnu.2022.06.039>
PMID:35839544
18. Wells JCK, Nesse RM, Sear R, Johnstone RA, Stearns SC. Evolutionary public health: introducing the concept. *Lancet*. 2017; 390:500–9.
[https://doi.org/10.1016/S0140-6736\(17\)30572-X](https://doi.org/10.1016/S0140-6736(17)30572-X)
PMID:28792412
19. Lin X, Yao Y, Wang B, Emlen DJ, Lavine LC. Ecological Trade-offs between Migration and Reproduction Are Mediated by the Nutrition-Sensitive Insulin-Signaling Pathway. *Int J Biol Sci*. 2016; 12:607–16.
<https://doi.org/10.7150/ijbs.14802>
PMID:27143957
20. Douros A, Dell'Aniello S, Yu OHY, Filion KB, Azoulay L, Suissa S. Sulfonyleureas as second line drugs in type 2 diabetes and the risk of cardiovascular and hypoglycaemic events: population based cohort study. *BMJ*. 2018; 362:k2693.
<https://doi.org/10.1136/bmj.k2693>
PMID:30021781
21. Kolb H, Kempf K, Röhling M, Martin S. Insulin: too much of a good thing is bad. *BMC Med*. 2020; 18:224.
<https://doi.org/10.1186/s12916-020-01688-6>
PMID:32819363
22. Zhao JV, Schooling CM. Sex-specific associations of insulin resistance with chronic kidney disease and kidney function: a bi-directional Mendelian randomisation study. *Diabetologia*. 2020; 63:1554–63.
<https://doi.org/10.1007/s00125-020-05163-y>
PMID:32409868
23. Zhao JV, Luo S, Schooling CM. Sex-specific Mendelian randomization study of genetically predicted insulin and cardiovascular events in the UK Biobank. *Commun Biol*. 2019; 2:332.
<https://doi.org/10.1038/s42003-019-0579-z>
PMID:31508506
24. Prasannarong M, Vichaiwong K, Saengsirisuwan V. Calorie restriction prevents the development of insulin resistance and impaired insulin signaling in skeletal muscle of ovariectomized rats. *Biochim Biophys Acta*. 2012; 1822:1051–61.
<https://doi.org/10.1016/j.bbadis.2012.02.018>
PMID:22406051
25. Suchacki KJ, Thomas BJ, Ikushima YM, Chen KC, Fyfe C, Tavares AAS, Sulston RJ, Lovdel A, Woodward HJ, Han X, Mattiucci D, Brain EJ, Alcaide-Corral CJ, et al. The effects of caloric restriction on adipose tissue and metabolic health are sex-and age-dependent. *Elife*. 2023; 12:e88080.

<https://doi.org/10.7554/eLife.88080>

PMID:[37096321](https://pubmed.ncbi.nlm.nih.gov/37096321/)

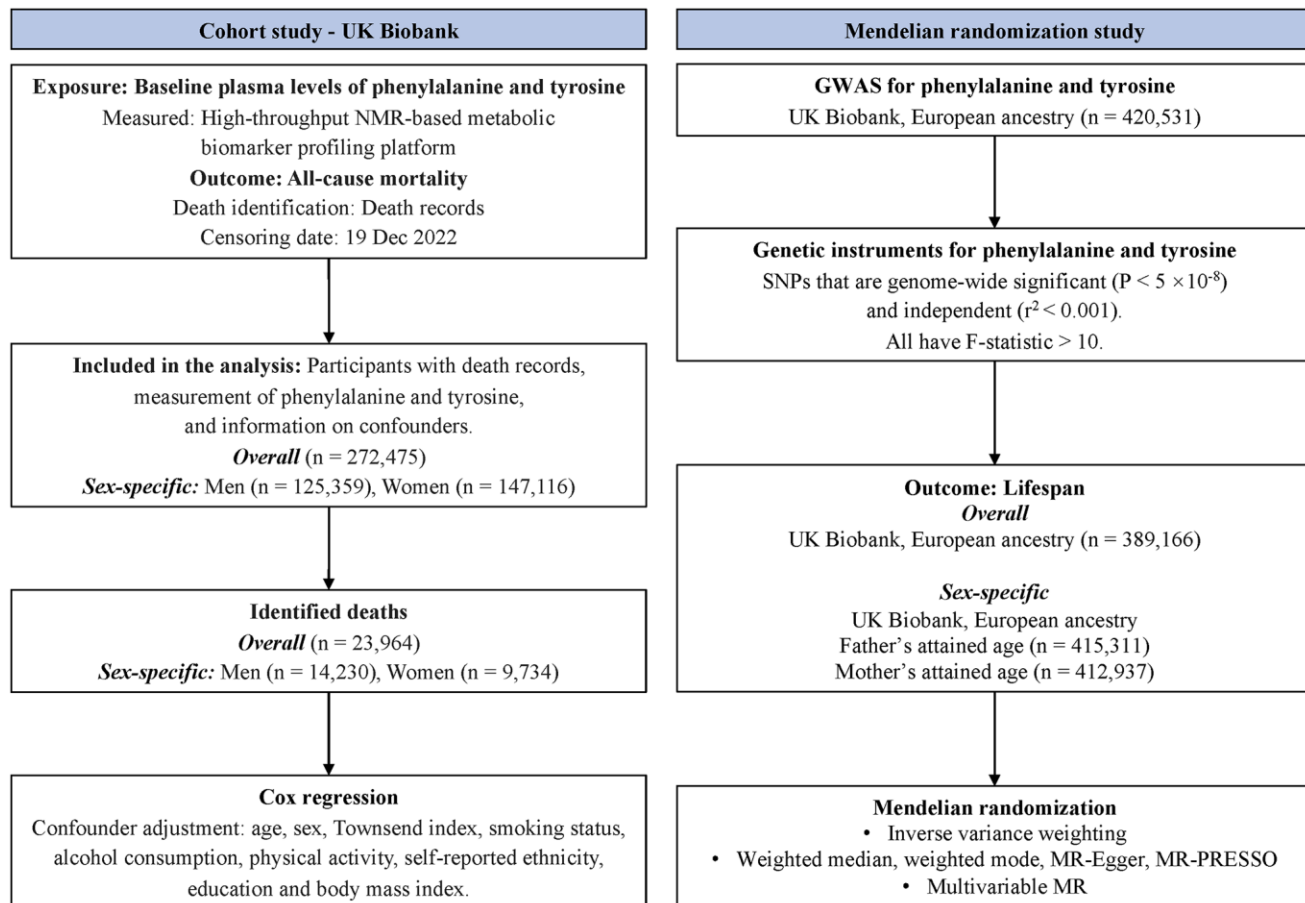
26. Schooling CM, Zhao JV. Investigating the association of testosterone with survival in men and women using a Mendelian randomization study in the UK Biobank. *Sci Rep*. 2021; 11:14039.
<https://doi.org/10.1038/s41598-021-93360-z>
PMID:[34234209](https://pubmed.ncbi.nlm.nih.gov/34234209/)
27. Purves-Tyson TD, Owens SJ, Double KL, Desai R, Handelsman DJ, Weickert CS. Testosterone induces molecular changes in dopamine signaling pathway molecules in the adolescent male rat nigrostriatal pathway. *PLoS One*. 2014; 9:e91151.
<https://doi.org/10.1371/journal.pone.0091151>
PMID:[24618531](https://pubmed.ncbi.nlm.nih.gov/24618531/)
28. McEwen BS, Milner TA. Understanding the broad influence of sex hormones and sex differences in the brain. *J Neurosci Res*. 2017; 95:24–39.
<https://doi.org/10.1002/jnr.23809>
PMID:[27870427](https://pubmed.ncbi.nlm.nih.gov/27870427/)
29. Kelly S. Individual socioeconomic status is important but hard to measure. *BMJ*. 2021; 372:n174.
<https://doi.org/10.1136/bmj.n174>
PMID:[33483303](https://pubmed.ncbi.nlm.nih.gov/33483303/)
30. Davey Smith G, Hemani G. Mendelian randomization: genetic anchors for causal inference in epidemiological studies. *Hum Mol Genet*. 2014; 23:R89–98.
<https://doi.org/10.1093/hmg/ddu328>
PMID:[25064373](https://pubmed.ncbi.nlm.nih.gov/25064373/)
31. Minelli C, Del Greco M F, van der Plaat DA, Bowden J, Sheehan NA, Thompson J. The use of two-sample methods for Mendelian randomization analyses on single large datasets. *Int J Epidemiol*. 2021; 50:1651–9.
<https://doi.org/10.1093/ije/dyab084>
PMID:[33899104](https://pubmed.ncbi.nlm.nih.gov/33899104/)
32. van de Rest O, Bloemendaal M, de Heus R, Aarts E. Dose-Dependent Effects of Oral Tyrosine Administration on Plasma Tyrosine Levels and Cognition in Aging. *Nutrients*. 2017; 9:1279.
<https://doi.org/10.3390/nu9121279>
PMID:[29168741](https://pubmed.ncbi.nlm.nih.gov/29168741/)
33. Melamed E, Glaeser B, Growdon JH, Wurtman RJ. Plasma tyrosine in normal humans: effects of oral tyrosine and protein-containing meals. *J Neural Transm*. 1980; 47:299–306.
<https://doi.org/10.1007/BF01247323>
PMID:[7190187](https://pubmed.ncbi.nlm.nih.gov/7190187/)
34. Davis BA, O'Reilly RL, Placatka CL, Paterson IA, Yu PH, Durden DA. Effect of dietary phenylalanine on the plasma concentrations of phenylalanine, phenylethylamine and phenylacetic acid in healthy volunteers. *Prog Neuropsychopharmacol Biol Psychiatry*. 1991; 15:611–23.
[https://doi.org/10.1016/0278-5846\(91\)90051-2](https://doi.org/10.1016/0278-5846(91)90051-2)
PMID:[1956990](https://pubmed.ncbi.nlm.nih.gov/1956990/)
35. Sudlow C, Gallacher J, Allen N, Beral V, Burton P, Danesh J, Downey P, Elliott P, Green J, Landray M, Liu B, Matthews P, Ong G, et al. UK biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. *PLoS Med*. 2015; 12:e1001779.
<https://doi.org/10.1371/journal.pmed.1001779>
PMID:[25826379](https://pubmed.ncbi.nlm.nih.gov/25826379/)
36. Harrell FE Jr, Lee KL, Pollock BG. Regression models in clinical studies: determining relationships between predictors and response. *J Natl Cancer Inst*. 1988; 80:1198–202.
<https://doi.org/10.1093/jnci/80.15.1198>
PMID:[3047407](https://pubmed.ncbi.nlm.nih.gov/3047407/)
37. Julkunen H, Cichońska A, Tiainen M, Koskela H, Nybo K, Mäkelä V, Nokso-Koivisto J, Kristiansson K, Perola M, Salomaa V, Jousilahti P, Lundqvist A, Kangas AJ, et al. Atlas of plasma NMR biomarkers for health and disease in 118,461 individuals from the UK Biobank. *Nat Commun*. 2023; 14:604.
<https://doi.org/10.1038/s41467-023-36231-7>
PMID:[36737450](https://pubmed.ncbi.nlm.nih.gov/36737450/)
38. Würtz P, Kangas AJ, Soininen P, Lawlor DA, Davey Smith G, Ala-Korpela M. Quantitative Serum Nuclear Magnetic Resonance Metabolomics in Large-Scale Epidemiology: A Primer on -Omic Technologies. *Am J Epidemiol*. 2017; 186:1084–96.
<https://doi.org/10.1093/aje/kwx016>
PMID:[29106475](https://pubmed.ncbi.nlm.nih.gov/29106475/)
39. Jiang L, Zheng Z, Qi T, Kemper KE, Wray NR, Visscher PM, Yang J. A resource-efficient tool for mixed model association analysis of large-scale data. *Nat Genet*. 2019; 51:1749–55.
<https://doi.org/10.1038/s41588-019-0530-8>
PMID:[31768069](https://pubmed.ncbi.nlm.nih.gov/31768069/)
40. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet*. 2011; 88:76–82.
<https://doi.org/10.1016/j.ajhg.2010.11.011>
PMID:[21167468](https://pubmed.ncbi.nlm.nih.gov/21167468/)
41. Pan-UK Biobank. <https://pan.ukbb.broadinstitute.org/>.
42. McCaw ZR, Lane JM, Saxena R, Redline S, Lin X. Operating characteristics of the rank-based inverse normal transformation for quantitative trait analysis in genome-wide association studies. *Biometrics*. 2020; 76:1262–72.
<https://doi.org/10.1111/biom.13214>
PMID:[31883270](https://pubmed.ncbi.nlm.nih.gov/31883270/)

43. Bulik-Sullivan BK, Loh PR, Finucane HK, Ripke S, Yang J, Patterson N, Daly MJ, Price AL, Neale BM, and Schizophrenia Working Group of the Psychiatric Genomics Consortium. LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet.* 2015; 47:291–5.
<https://doi.org/10.1038/ng.3211>
PMID:25642630
44. Staiger D, Stock JH. Instrumental variables regression with weak instruments. *Econometrica.* 1997; 65:557–86.
<https://doi.org/10.2307/2171753>
45. Bowden J, Del Greco M F, Minelli C, Davey Smith G, Sheehan NA, Thompson JR. Assessing the suitability of summary data for two-sample Mendelian randomization analyses using MR-Egger regression: the role of the I² statistic. *Int J Epidemiol.* 2016; 45:1961–74.
<https://doi.org/10.1093/ije/dyw220>
PMID:27616674
46. Pilling LC, Kuo CL, Sicinski K, Tamosauskaite J, Kuchel GA, Harries LW, Herd P, Wallace R, Ferrucci L, Melzer D. Human longevity: 25 genetic loci associated in 389,166 UK biobank participants. *Aging (Albany NY).* 2017; 9:2504–20.
<https://doi.org/10.18632/aging.101334>
PMID:29227965
47. Dutta A, Henley W, Robine JM, Langa KM, Wallace RB, Melzer D. Longer lived parents: protective associations with cancer incidence and overall mortality. *J Gerontol A Biol Sci Med Sci.* 2013; 68:1409–18.
<https://doi.org/10.1093/gerona/glt061>
PMID:23685624
48. Zenin A, Tsepilov Y, Sharapov S, Getmantsev E, Menshikov LI, Fedichev PO, Aulchenko Y. Identification of 12 genetic loci associated with human healthspan. *Commun Biol.* 2019; 2:41.
<https://doi.org/10.1038/s42003-019-0290-0>
PMID:30729179
49. Timmers PR, Mounier N, Lall K, Fischer K, Ning Z, Feng X, Bretherick AD, Clark DW, Shen X, Esko T, Kutalik Z, Wilson JF, Joshi PK, and eQTLGen Consortium. Genomics of 1 million parent lifespans implicates novel pathways and common diseases and distinguishes survival chances. *Elife.* 2019; 8:e39856.
<https://doi.org/10.7554/eLife.39856>
PMID:30642433
50. Schooling CM, Kwok MK, Zhao JV. The relationship of fatty acids to ischaemic heart disease and lifespan in men and women using Mendelian randomization. *Int J Epidemiol.* 2023; 52:1845–52.
<https://doi.org/10.1093/ije/dyad108>
PMID:37536998
51. Burgess S, Scott RA, Timpson NJ, Davey Smith G, Thompson SG, and EPIC- InterAct Consortium. Using published data in Mendelian randomization: a blueprint for efficient identification of causal risk factors. *Eur J Epidemiol.* 2015; 30:543–52.
<https://doi.org/10.1007/s10654-015-0011-z>
PMID:25773750
52. Zhao JV, Fan B, Burgess S. Using genetics to examine the overall and sex-specific associations of branch-chain amino acids and the valine metabolite, 3-hydroxyisobutyrate, with ischemic heart disease and diabetes: A two-sample Mendelian randomization study. *Atherosclerosis.* 2023; 381:117246.
<https://doi.org/10.1016/j.atherosclerosis.2023.117246>
PMID:37660674
53. Zhao JV, Burgess S, Fan B, Schooling CM. L-carnitine, a friend or foe for cardiovascular disease? A Mendelian randomization study. *BMC Med.* 2022; 20:272.
<https://doi.org/10.1186/s12916-022-02477-z>
PMID:36045366
54. Zhao JV, Yao M, Liu Z. Using genetics and proteomics data to identify proteins causally related to COVID-19, healthspan and lifespan: a Mendelian randomization study. *Aging (Albany NY).* 2024; 16:6384–416.
<https://doi.org/10.18632/aging.205711>
PMID:38575325
55. Zhao JV, Schooling CM. The role of testosterone in chronic kidney disease and kidney function in men and women: a bi-directional Mendelian randomization study in the UK Biobank. *BMC Med.* 2020; 18:122.
<https://doi.org/10.1186/s12916-020-01594-x>
PMID:32493397
56. Burgess S, Bowden J, Fall T, Ingelsson E, Thompson SG. Sensitivity Analyses for Robust Causal Inference from Mendelian Randomization Analyses with Multiple Genetic Variants. *Epidemiology.* 2017; 28:30–42.
<https://doi.org/10.1097/EDE.0000000000000559>
PMID:27749700
57. Hartwig FP, Davey Smith G, Bowden J. Robust inference in summary data Mendelian randomization via the zero modal pleiotropy assumption. *Int J Epidemiol.* 2017; 46:1985–98.
<https://doi.org/10.1093/ije/dyx102>
PMID:29040600
58. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization

- between complex traits and diseases. *Nat Genet.* 2018; 50:693–8.
<https://doi.org/10.1038/s41588-018-0099-7>
 PMID:[29686387](https://pubmed.ncbi.nlm.nih.gov/29686387/)
59. Burgess S, Thompson SG. Interpreting findings from Mendelian randomization using the MR-Egger method. *Eur J Epidemiol.* 2017; 32:377–89.
<https://doi.org/10.1007/s10654-017-0255-x>
 PMID:[28527048](https://pubmed.ncbi.nlm.nih.gov/28527048/)
 60. Slob EAW, Burgess S. A comparison of robust Mendelian randomization methods using summary data. *Genet Epidemiol.* 2020; 44:313–29.
<https://doi.org/10.1002/gepi.22295>
 PMID:[32249995](https://pubmed.ncbi.nlm.nih.gov/32249995/)
 61. Karjalainen MK, Karthikeyan S, Oliver-Williams C, Sliz E, Allara E, Fung WT, Surendran P, Zhang W, Jousilahti P, Kristiansson K, Salomaa V, Goodwin M, Hughes DA, et al, and China Kadoorie Biobank Collaborative Group, and Estonian Biobank Research Team, and FinnGen. Genome-wide characterization of circulating metabolic biomarkers. *Nature.* 2024; 628:130–8.
<https://doi.org/10.1038/s41586-024-07148-y>
 PMID:[38448586](https://pubmed.ncbi.nlm.nih.gov/38448586/)
 62. Burgess S, Thompson SG. Multivariable Mendelian randomization: the use of pleiotropic genetic variants to estimate causal effects. *Am J Epidemiol.* 2015; 181:251–60.
<https://doi.org/10.1093/aje/kwu283>
 PMID:[25632051](https://pubmed.ncbi.nlm.nih.gov/25632051/)
 63. Rees JMB, Wood AM, Burgess S. Extending the MR-Egger method for multivariable Mendelian randomization to correct for both measured and unmeasured pleiotropy. *Stat Med.* 2017; 36:4705–18.
<https://doi.org/10.1002/sim.7492>
 PMID:[28960498](https://pubmed.ncbi.nlm.nih.gov/28960498/)
 64. Freeman G, Cowling BJ, Schooling CM. Power and sample size calculations for Mendelian randomization studies using one genetic instrument. *Int J Epidemiol.* 2013; 42:1157–63.
<https://doi.org/10.1093/ije/dyt110>
 PMID:[23934314](https://pubmed.ncbi.nlm.nih.gov/23934314/)
 65. Guan W, Steffen BT, Lemaitre RN, Wu JHY, Tanaka T, Manichaikul A, Foy M, Rich SS, Wang L, Nettleton JA, Tang W, Gu X, Bandinelli S, et al. Genome-wide association study of plasma N6 polyunsaturated fatty acids within the cohorts for heart and aging research in genomic epidemiology consortium. *Circ Cardiovasc Genet.* 2014; 7:321–31.
<https://doi.org/10.1161/CIRCGENETICS.113.000208>
 PMID:[24823311](https://pubmed.ncbi.nlm.nih.gov/24823311/)
 66. Burgess S. Sample size and power calculations in Mendelian randomization with a single instrumental variable and a binary outcome. *Int J Epidemiol.* 2014; 43:922–9.
<https://doi.org/10.1093/ije/dyu005>
 PMID:[24608958](https://pubmed.ncbi.nlm.nih.gov/24608958/)

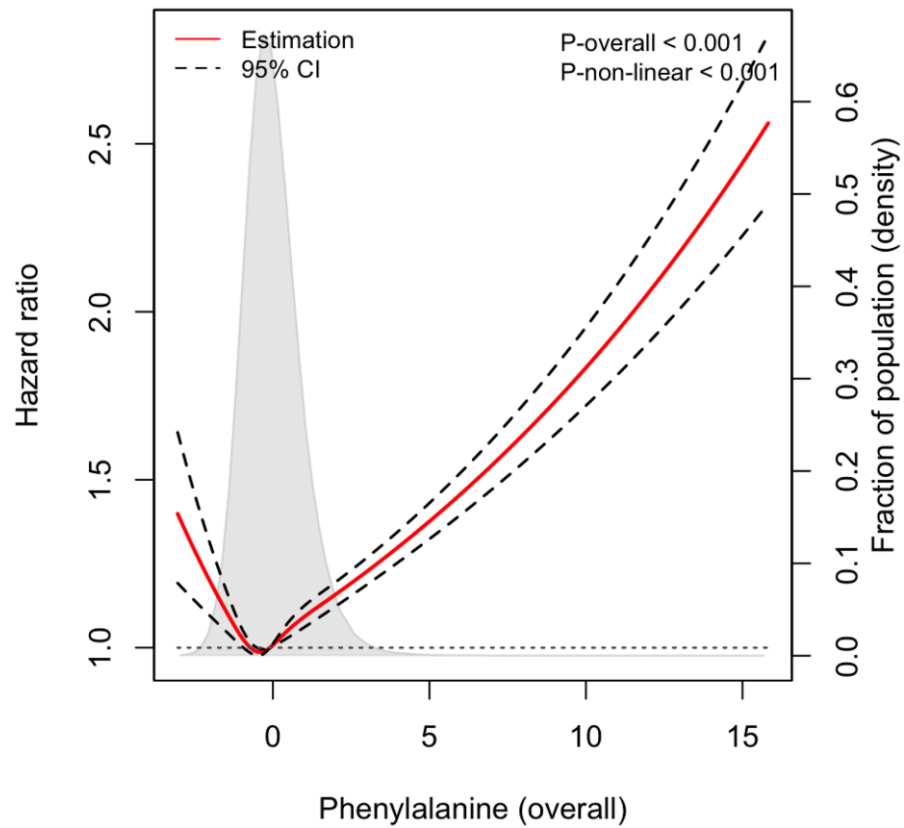
SUPPLEMENTARY MATERIALS

Supplementary Figures



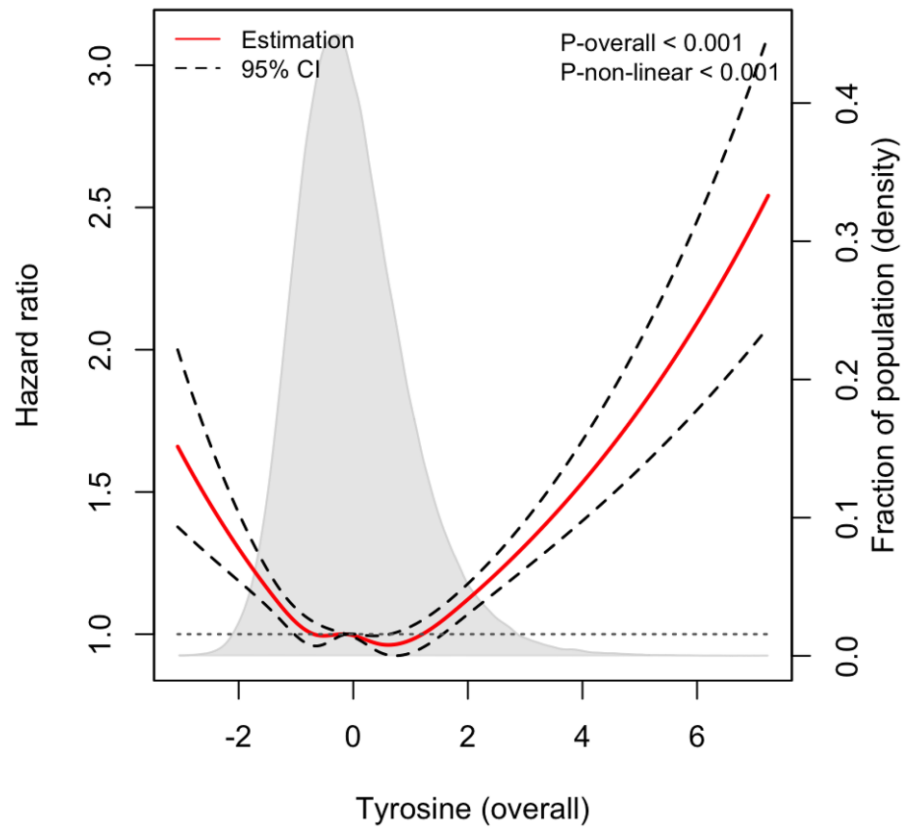
Supplementary Figure 1. Flow chart of study design.

All-cause Mortality

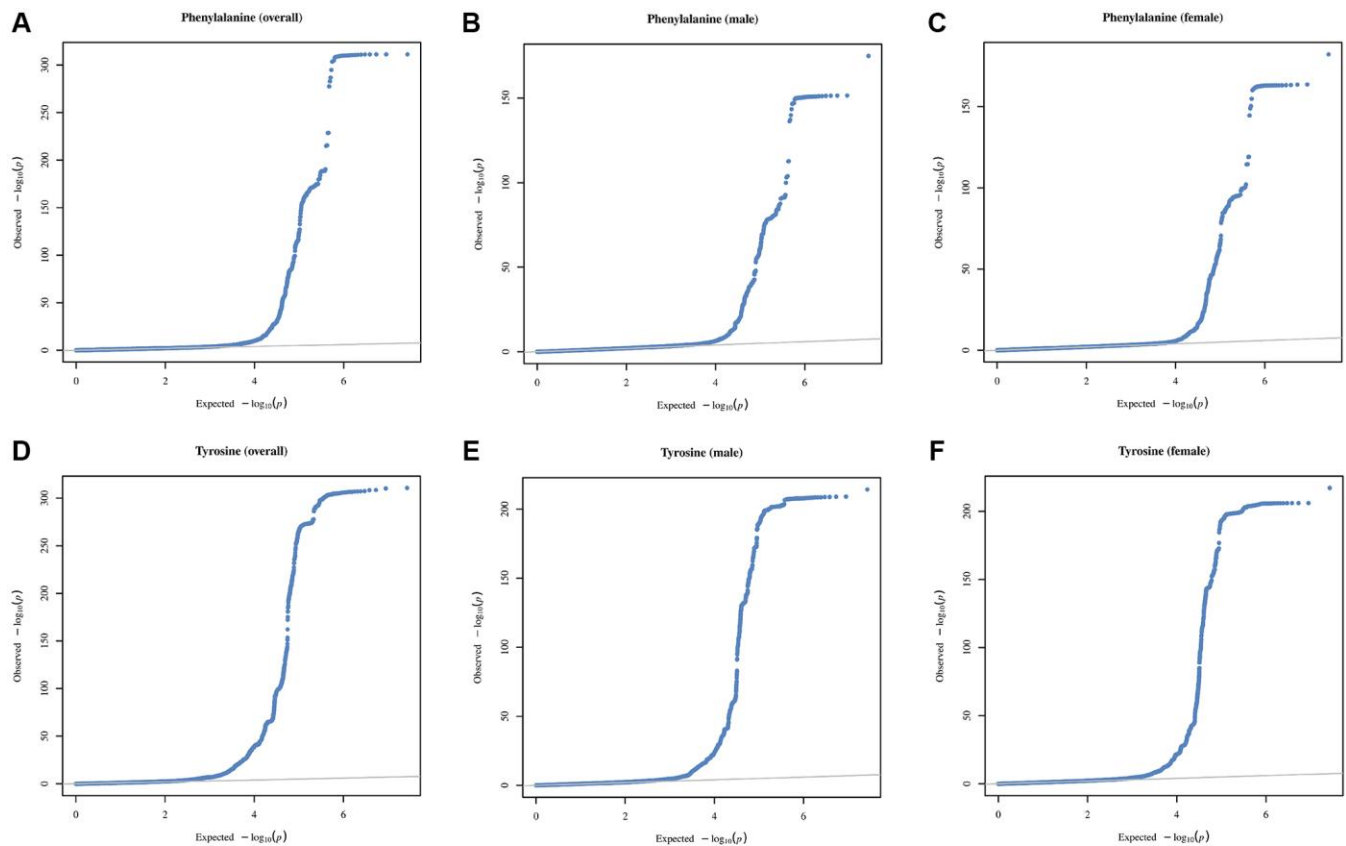


Supplementary Figure 2. Association of phenylalanine with all-cause mortality using restricted cubic splines.

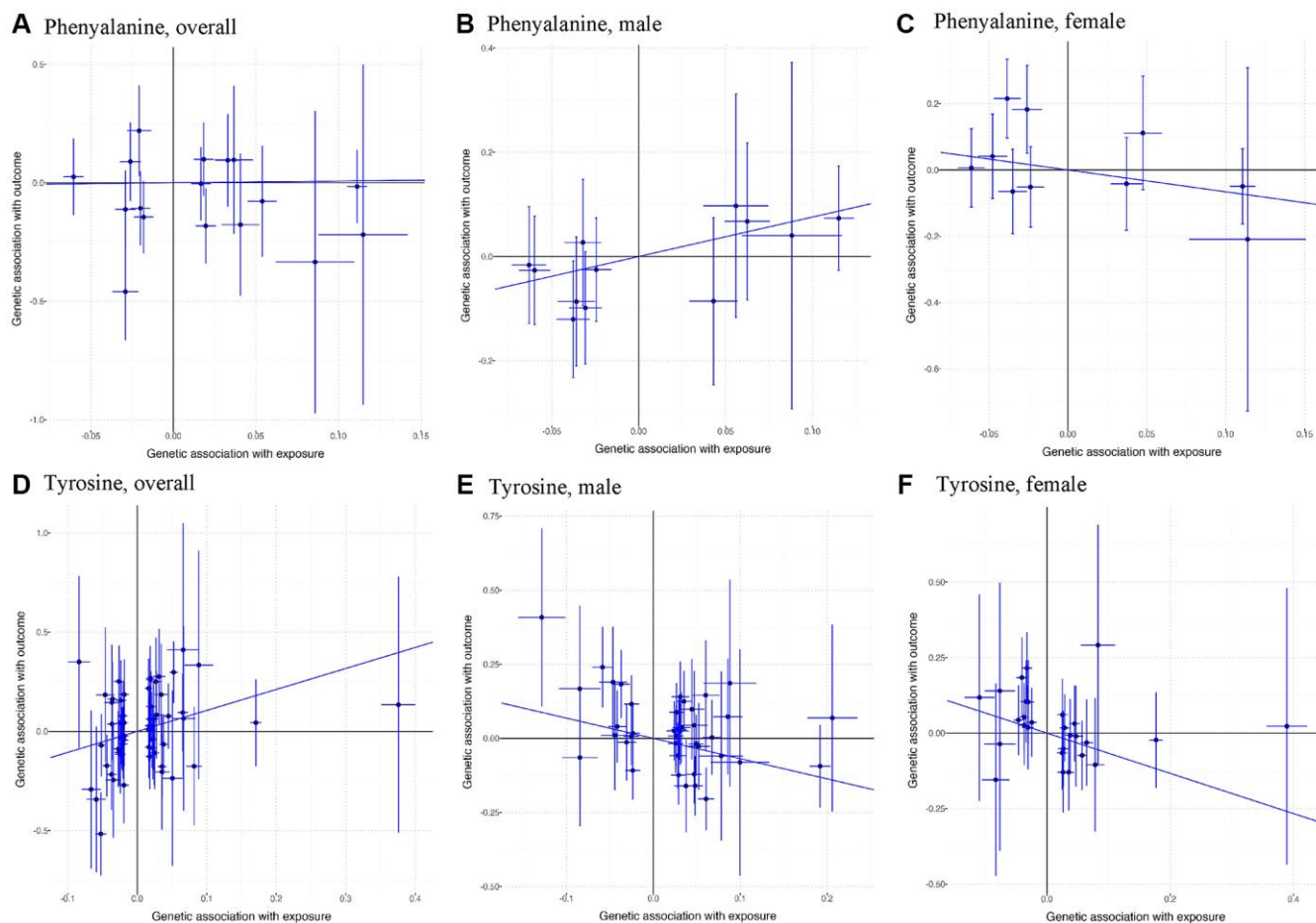
All-cause Mortality



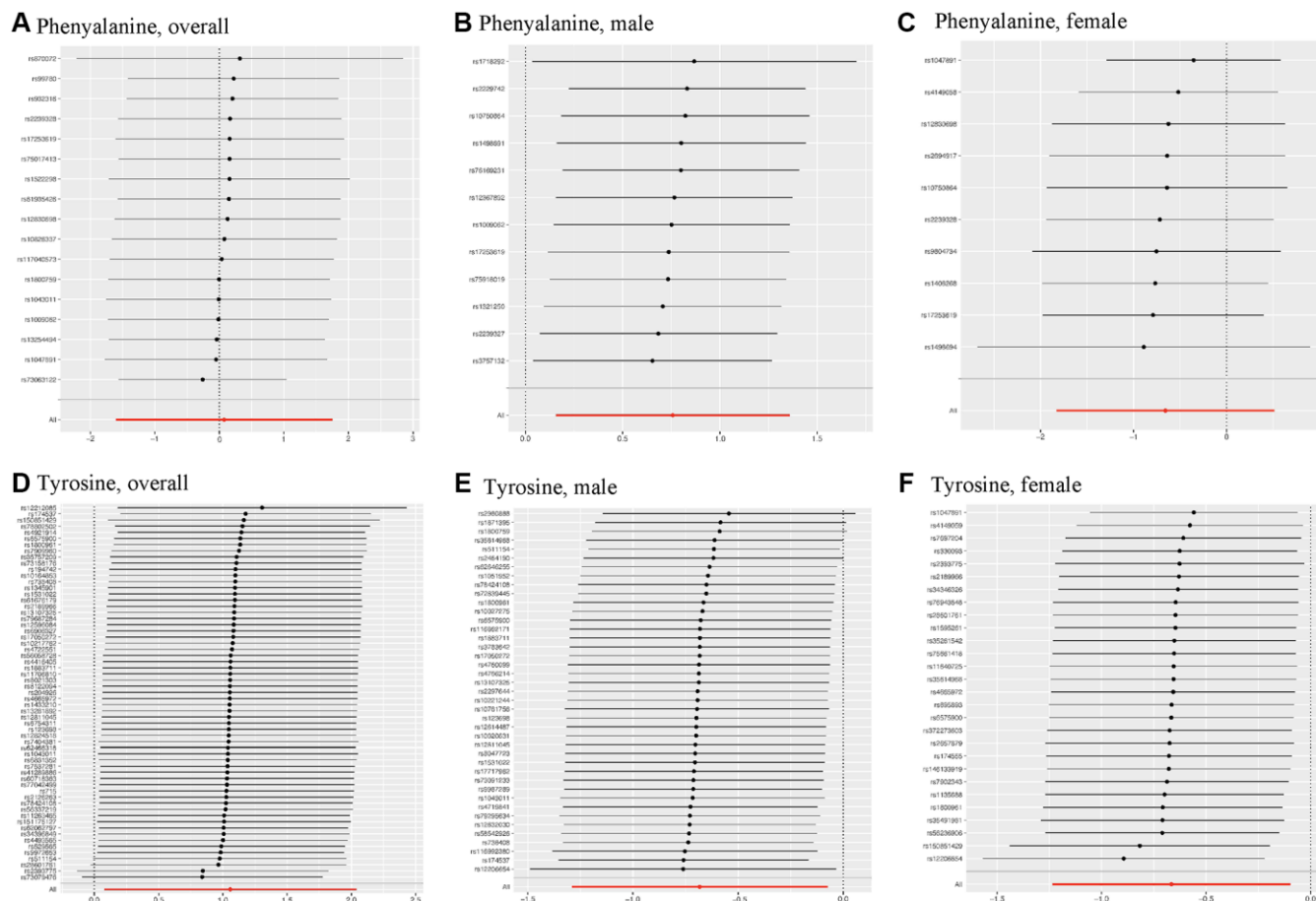
Supplementary Figure 3. Association of tyrosine with all-cause mortality using restricted cubic splines.



Supplementary Figure 4. Q-Q plot on the genome-wide association study of phenylalanine and tyrosine in overall people, men and women. (A) Q-Q plot for phenylalanine in overall people; (B) Q-Q plot for phenylalanine in men; (C) Q-Q plot for phenylalanine in women; (D) Q-Q plot for tyrosine in overall people; (E) Q-Q plot for tyrosine in men; (F) Q-Q plot for tyrosine in women.



Supplementary Figure 5. Scatter plots on the associations of each SNP with amino acids and with lifespan in overall people, men and women. (A) Scatter plot for phenylalanine in overall people; (B) Scatter plot for phenylalanine in men; (C) Scatter plot for phenylalanine in women; (D) Scatter plot for tyrosine in overall people; (E) Scatter plot for tyrosine in men; (F) Scatter plot for tyrosine in women.



Supplementary Figure 6. Leave-one-out analysis on the associations of phenylalanine and tyrosine with lifespan. (A) Leave-one-out analysis for phenylalanine in overall people; (B) leave-one-out analysis for phenylalanine in men; (C) leave-one-out analysis for phenylalanine in women; (D) leave-one-out analysis for tyrosine in overall people; (E) leave-one-out analysis for tyrosine in men; (F) leave-one-out analysis for tyrosine in women.

Supplementary Tables

Supplementary Table 1. The associations of phenylalanine and tyrosine with all-cause mortality excluding deaths from accidents in UK Biobank using Cox regression.

Exposure	Sex	HR ¹	95% CI ²	<i>p</i>
Phenylalanine	Overall	1.04	1.03, 1.05	3.3×10^{-9}
	Men	1.04	1.02, 1.05	3.2×10^{-7}
	Women	1.04	1.02, 1.07	8.7×10^{-4}
Tyrosine	Overall	1.02	1.00, 1.03	1.9×10^{-2}
	Men	1.03	1.01, 1.05	3.4×10^{-3}
	Women	1.00	0.98, 1.03	7.8×10^{-1}

¹HR: hazard ratio. In overall analysis we adjusted for age, body mass index (BMI), Townsend Deprivation Index, smoking status, alcohol consumption, physical activity, ethnicity, and education (in years). For combined-sex analyses, we additionally adjusted for sex. ²CI: confidence interval.

Supplementary Table 2. The associations of phenylalanine and tyrosine with all-cause mortality stratified by amino acid level in UK Biobank using Cox regression.

Exposure	Stratified by amino acids levels ¹	HR ²	95% CI ³	<i>p</i>
Phenylalanine	Lower level	0.95	0.91, 0.99	2.5×10^{-2}
	Higher level	1.05	1.04, 1.06	8.4×10^{-17}
Tyrosine	Lower level	0.89	0.85, 0.93	2.7×10^{-13}
	Higher level	1.10	1.07, 1.13	4.5×10^{-7}

¹Higher or lower level means above or below standardized concentration of 0 for phenylalanine and tyrosine. ²HR: hazard ratio. We adjusted for age, sex, BMI, Townsend Deprivation Index, smoking status, alcohol consumption, physical activity, ethnicity, and education (in years). ³CI: confidence interval.

Supplementary Table 3. The associations of phenylalanine and tyrosine with cardiovascular disease (CVD) and cancer mortality in UK Biobank using Cox regression.

Exposure	Outcome	Sex	HR ¹	95% CI ²	<i>p</i>
Phenylalanine	CVD mortality	Overall	1.03	1.00, 1.06	3.4×10^{-2}
		Men	1.03	0.99, 1.07	1.7×10^{-1}
		Women	1.05	0.99, 1.12	9.3×10^{-2}
Tyrosine	CVD mortality	Overall	1.01	0.98, 1.04	6.3×10^{-1}
		Men	1.01	0.97, 1.05	7.0×10^{-1}
		Women	1.00	0.94, 1.07	9.8×10^{-1}
Phenylalanine	Cancer mortality	Overall	1.04	1.02, 1.05	6.3×10^{-5}
		Men	1.04	1.01, 1.06	2.7×10^{-3}
		Women	1.04	1.01, 1.07	2.2×10^{-2}
Tyrosine	Cancer mortality	Overall	1.02	0.99, 1.04	1.6×10^{-1}
		Men	1.01	0.99, 1.04	3.3×10^{-1}
		Women	1.02	0.99, 1.06	1.9×10^{-1}

¹HR: hazard ratio. In overall analysis we adjusted for age, BMI, Townsend Deprivation Index, smoking status, alcohol consumption, physical activity, ethnicity, and education (in years). For combined-sex analyses, we additionally adjusted for sex. ²CI: confidence interval.

Supplementary Table 4. Linkage disequilibrium score regression.

Phenotype	Population	Sex	h ²	h ² _se	lambda	intercept	intercept_se	ratio	ratio_se
Phenylalanine	EUR	Overall	0.036	0.009	1.121	1.028	0.009	0.143	0.043
Phenylalanine	EUR	Female	0.031	0.008	1.068	1.014	0.008	0.145	0.080
Phenylalanine	EUR	Male	0.042	0.010	1.059	1.011	0.009	0.110	0.082
Tyrosine	EUR	Overall	0.087	0.018	1.250	1.062	0.011	0.120	0.022
Tyrosine	EUR	Female	0.080	0.023	1.127	1.040	0.009	0.153	0.035
Tyrosine	EUR	Male	0.114	0.026	1.152	1.035	0.010	0.114	0.032

Supplementary Table 5. Genetic instruments for phenylalanine in the overall analysis.

SNP	effect_allele	other_allele	gene	beta	se	p-value	F-statistics
rs1009062	G	T	<i>GSTA2</i>	−0.02	0.003	1.76E-11	45.2
rs1043011	T	G	<i>GLS2, SPRYD4</i>	0.033	0.004	1.40E-18	77.4
rs1047891	A	C	<i>CPS1</i>	−0.029	0.003	1.25E-20	86.7
rs10750864	T	A	<i>SLC43A1</i>	−0.055	0.003	3.37E-59	263.2
rs10826337	A	G	<i>SLC16A9, MRPL50P4</i>	0.017	0.003	1.30E-08	32.3
rs117040573	A	G	<i>C12orf42</i>	0.037	0.006	3.92E-10	39.2
rs12830698	G	T	<i>STAB2</i>	0.115	0.014	6.48E-17	69.8
rs13254494	C	T	<i>SLC25A37</i>	−0.018	0.003	1.02E-09	37.3
rs140584594	A	G	<i>GSTM1</i>	0.022	0.003	6.40E-12	47.2
rs1522298	C	G	<i>PAH</i>	−0.06	0.003	7.19E-85	381.1
rs17253619	C	T	<i>WDHD1</i>	0.054	0.004	1.45E-33	145.8
rs1800759	T	G	<i>ADH4, LOC100507053</i>	0.018	0.003	7.58E-10	37.9
rs2239328	T	C	<i>ABCC6</i>	−0.026	0.003	2.29E-16	67.3
rs34121855	G	T	<i>MLXIPL</i>	−0.021	0.004	1.04E-08	32.8
rs3757132	T	C	<i>SLC17A1</i>	−0.036	0.003	1.83E-27	117.9
rs61935426	A	C	<i>LINC02456</i>	0.086	0.012	9.04E-13	51
rs73063122	C	A	<i>SLCO1B1</i>	−0.029	0.004	2.36E-13	53.7
rs75017413	A	T	<i>SLC38A4</i>	0.041	0.006	2.41E-12	49.1
rs870072	C	T	<i>PAH</i>	0.111	0.003	2.66E-303	1408.7
rs932316	C	T	<i>SCGN, CARMIL1</i>	−0.021	0.004	1.44E-08	32.1
rs99780	T	C	<i>FADS2</i>	0.02	0.003	9.88E-11	41.8

Supplementary Table 6. Genetic instruments for tyrosine in the overall analysis.

SNP	effect_allele	other_allele	gene	beta	se	p-value	F-statistics
rs10027275	G	C	<i>ARHGAP10</i>	0.032	0.003	4.72E-22	93.2
rs10164853	G	A	<i>ACVR1C</i>	0.036	0.006	1.27E-10	41.3
rs10217762	C	T	<i>CDKN2B-AS1</i>	−0.019	0.003	9.61E-11	41.9
rs1043011	T	G	<i>GLS2, SPRYD4</i>	0.065	0.004	3.94E-69	308.8
rs10750864	T	A	<i>SLC43A1</i>	−0.02	0.003	2.06E-09	35.9
rs11263465	G	A	<i>LOC105369370</i>	0.027	0.004	1.86E-10	40.6
rs114232169	T	G	<i>HRG, HRG-AS1</i>	0.025	0.003	1.88E-16	67.7

rs11614623	T	C	<i>HPD</i>	0.049	0.004	4.14E-29	125.4
rs11643623	T	C	<i>ZNF276</i>	0.018	0.003	1.86E-08	31.6
rs11706810	C	T	<i>TRIM59, TRIM59-IFT80</i>	-0.02	0.003	4.93E-12	47.7
rs12212085	C	A	<i>SLC16A10</i>	0.171	0.004	0	1699.8
rs123698	G	C	<i>PTBP1</i>	0.018	0.003	9.64E-10	37.4
rs12596084	C	A	<i>RNA5SP427, MPHOSPH10P1</i>	-0.02	0.003	4.99E-10	38.7
rs12811045	G	A	<i>LOC102723639</i>	0.023	0.004	6.24E-11	42.7
rs12824518	T	C	/	0.028	0.004	4.60E-11	43.3
rs13107325	T	C	<i>SLC39A8</i>	-0.037	0.005	2.36E-11	44.6
rs13142887	T	A	/	-0.018	0.003	7.40E-09	33.4
rs13281892	G	A	<i>SLC7A2</i>	-0.021	0.003	2.55E-11	44.5
rs1345901	C	T	<i>LOC105371356</i>	0.022	0.003	1.15E-13	55.1
rs140584594	A	G	<i>GSTM1</i>	0.019	0.003	4.71E-09	34.3
rs1433210	C	A	<i>LINC01091</i>	-0.02	0.003	5.78E-09	33.9
rs150851429	C	G	<i>LOC105371334</i>	0.376	0.013	5.51E-198	901.1
rs151175127	T	C	/	-0.059	0.007	4.22E-17	70.7
rs1531022	A	G	<i>UGT2B15</i>	0.018	0.003	6.05E-10	38.3
rs17050272	A	G	<i>LOC105373585</i>	0.023	0.003	7.05E-15	60.6
rs174537	T	G	<i>MYRF</i>	0.036	0.003	5.38E-32	138.6
rs1800961	T	C	<i>HNF4A</i>	-0.084	0.008	4.67E-24	102.3
rs183657985	T	C	<i>EXOC3L2</i>	0.021	0.003	4.65E-10	38.8
rs1883711	C	G	<i>LINC01370, MAFB</i>	0.067	0.009	6.17E-15	60.8
rs194742	T	C	<i>MAGOH3P, ZFP36L1</i>	-0.024	0.004	7.99E-10	37.8
rs204926	G	A	<i>LMO1</i>	0.017	0.003	3.09E-09	35.1
rs2126263	G	A	<i>LOC157273</i>	0.034	0.005	1.13E-12	50.6
rs2189966	C	T	<i>JAZF1</i>	-0.037	0.004	3.74E-25	107.3
rs2393775	G	A	<i>HNF1A</i>	0.052	0.003	2.32E-69	309.9
rs28601761	G	C	<i>TRIB1AL</i>	-0.044	0.003	4.79E-49	216.7
rs34396849	C	A	<i>PGBD1</i>	0.031	0.004	1.80E-12	49.7
rs35048664	G	T	<i>PAH</i>	0.02	0.003	5.65E-10	38.4
rs35757209	T	C	<i>UNK</i>	0.024	0.003	1.42E-15	63.7
rs41289886	A	G	<i>RPF2</i>	0.066	0.012	1.88E-08	31.6
rs4416405	A	G	/	0.016	0.003	3.95E-08	30.2
rs4493565	A	C	<i>SHROOM3</i>	0.016	0.003	1.40E-08	32.2
rs4665972	T	C	<i>SNX17</i>	-0.022	0.003	7.13E-14	56
rs4722551	C	T	<i>LOC105375199</i>	-0.023	0.004	3.18E-09	35.1
rs4921914	C	T	<i>PSD3, NAT2</i>	-0.026	0.003	2.30E-14	58.3
rs511154	A	G	<i>RPL31P23, PCCB</i>	-0.037	0.003	1.24E-27	118.7
rs529565	C	T	<i>ABO</i>	0.019	0.003	1.50E-09	36.5
rs56058728	A	G	<i>INSR</i>	0.021	0.004	1.03E-08	32.8
rs56337219	T	C	<i>SLC16A10</i>	-0.066	0.007	3.57E-21	89.2
rs56401710	C	A	<i>SLC22A7,</i>	0.024	0.003	3.35E-16	66.6

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rs60718363	C	T	WWC2	0.02	0.003	8.86E-09	33.1
rs61676179	A	C	DEPDC5	-0.046	0.007	1.88E-12	49.6
rs62062797	G	T	MAPT	-0.019	0.003	2.44E-08	31.1
rs62466318	T	C	MLXIPL	-0.027	0.004	3.27E-14	57.6
rs6575900	C	G	WDR20, LOC105370677	-0.036	0.004	6.02E-23	97.3
rs6754311	C	T	DARS1	-0.021	0.003	2.01E-09	36
rs6831352	T	C	ADH4, LOC100507053	0.045	0.003	4.21E-46	203.2
rs6906327	A	G	CDKAL1	0.017	0.003	2.17E-08	31.3
rs715	C	T	CPS1	-0.026	0.003	3.28E-17	71.2
rs73079476	C	A	SLCO1B1	-0.053	0.004	1.67E-38	168.4
rs73158176	G	C	PRKAG2	-0.019	0.003	2.24E-08	31.3
rs738408	T	C	PNPLA3	0.025	0.003	8.18E-13	51.2
rs7404381	A	G	AP1G1	-0.052	0.003	4.54E-67	299.4
rs7537281	T	A	PPIAP34, ZBTB40	-0.024	0.004	1.12E-10	41.6
rs75891099	A	C	REV3L	0.154	0.02	3.78E-15	61.8
rs77042499	C	T	TRAF3IP2-AS1	0.089	0.01	2.69E-17	71.6
rs78424108	G	C	WBP4, MIR3168	-0.034	0.006	3.89E-10	39.2
rs78802502	A	G	SLC38A4	0.082	0.006	1.07E-45	201.3
rs7909960	A	T	JMJD1C	0.038	0.003	5.61E-39	170.6
rs79687284	C	G	PROX1-AS1	0.051	0.008	1.38E-10	41.2
rs8021303	A	G	WDHD1	-0.019	0.003	1.42E-10	41.1
rs8100204	A	G	SUGP1	0.029	0.004	4.64E-12	47.8
rs8122094	G	C	TOP1	0.019	0.003	4.37E-08	30
rs814573	T	A	APOC1P1, APOC1	-0.029	0.004	1.41E-14	59.2
rs9972653	T	G	FTO	0.019	0.003	1.81E-10	40.7

Supplementary Table 7. Genetic instruments for phenylalanine in the sex-specific analysis.

SNP	effect_allele	other_allele	gene	beta	se	p-value	F-statistics
<i>Male</i>							
rs1321250	C	T	/	-0.036	0.005	3.46E-11	43.9
rs3757132	T	C	SLC17A1	-0.038	0.005	9.59E-15	60.0
rs1009062	G	T	GSTA2	-0.025	0.004	2.08E-08	31.4
rs10750864	T	A	SLC43A1	-0.063	0.005	3.54E-37	162.3
rs75918019	G	A	SLC38A4	0.056	0.009	3.89E-09	34.7
rs76169231	C	T	LINC02456	-0.032	0.005	1.44E-09	36.6
rs1718292	G	A	PAH	0.115	0.004	3.15E-152	690.7
rs1498691	G	A	PAH	-0.060	0.005	1.28E-39	173.5
rs12367892	C	G	C12orf42	0.088	0.015	1.76E-09	36.2
rs17253619	C	T	WDHD1	0.062	0.007	1.72E-21	90.6
rs2239327	A	C	ABCC6	-0.031	0.005	7.45E-11	42.4
rs2229742	C	G	NR1P1	0.043	0.007	8.81E-10	37.6

<i>Female</i>							
rs1047891	A	C	<i>CPS1</i>	−0.039	0.004	1.13E-19	82.4
rs1408268	A	T	<i>SLC17A1</i>	−0.035	0.005	1.08E-14	59.7
rs10750864	T	A	<i>SLC43A1</i>	−0.048	0.005	2.24E-25	108.4
rs4149058	G	A	<i>SLCO1B1</i>	−0.026	0.005	4.82E-08	29.8
rs2694917	C	T	<i>RBMS2</i>	0.037	0.005	1.69E-13	54.3
rs1498694	A	G	<i>PAH, LOC124902999</i>	0.111	0.004	2.08E-164	746.7
rs9804734	T	C	<i>PAH</i>	−0.061	0.004	1.97E-47	209.3
rs12830698	G	T	<i>STAB2</i>	0.114	0.019	1.36E-09	36.7
rs17253619	C	T	<i>WDHD1</i>	0.047	0.006	7.39E-15	60.5
rs2239328	T	C	<i>ABCC6</i>	−0.024	0.004	4.38E-08	30.0

Supplementary Table 8. Genetic instruments for tyrosine in the sex-specific analysis.

SNP	effect_allele	other_allele	gene	beta	se	p-value	F-statistics
<i>Male</i>							
rs17050272	A	G	<i>LOC105373585</i>	0.025	0.004	4.28E-09	34.5
rs12614487	T	C	<i>ACVR1C</i>	0.047	0.008	5.43E-09	34.0
rs511154	A	G	<i>RPL31P23, PCCB</i>	−0.037	0.005	1.35E-13	54.8
rs12632030	C	T	<i>SMC4, TRIM59-IFT80</i>	−0.024	0.004	2.28E-08	31.2
rs5402	A	T	<i>SLC2A2</i>	0.040	0.007	1.27E-09	36.9
rs114232169	T	G	<i>HRG, HRG-AS1</i>	0.031	0.005	8.50E-12	46.6
rs10020631	A	G	<i>TMPRSS11E</i>	0.031	0.005	4.99E-10	38.7
rs1531022	A	G	<i>UGT2B15</i>	0.030	0.004	2.93E-12	48.7
rs1800759	T	G	<i>ADH4, LOC100507053</i>	0.048	0.004	8.85E-28	119.3
rs13107325	T	C	<i>SLC39A8</i>	−0.044	0.008	3.65E-08	30.3
rs10027275	G	C	<i>ARHGAP10</i>	0.028	0.005	1.53E-08	32.0
rs72839445	A	G	<i>CDCA7P1, POM121L2</i>	0.038	0.007	3.03E-08	30.7
rs1051952	C	A	<i>TEAD3, TULP1, LOC124901309</i>	−0.025	0.004	3.79E-09	34.7
rs62646255	C	T	<i>SLC22A7</i>	0.029	0.004	4.19E-11	43.5
rs12206654	C	T	<i>MFSD4B-DT</i>	0.192	0.006	5.81E-215	979.3
rs17717962	C	T	<i>LOC105377944</i>	0.088	0.015	1.08E-08	32.7
rs35614968	A	G	<i>FYN</i>	−0.129	0.014	3.85E-21	89.0
rs116862171	G	A	<i>FYN</i>	0.100	0.017	6.55E-09	33.7
rs4719841	G	A	<i>LOC105375199</i>	0.027	0.004	7.84E-10	37.8
rs73091233	C	T	<i>JAZF1</i>	−0.085	0.010	1.83E-16	67.8
rs9987289	A	G	<i>LOC157273</i>	0.044	0.007	2.22E-09	35.8
rs2980888	T	C	<i>TRIB1AL</i>	0.061	0.005	6.00E-39	170.4
rs10761756	T	C	<i>JMJD1C</i>	0.049	0.004	1.00E-30	132.8
rs2297644	C	T	<i>HOGA1</i>	−0.031	0.006	4.63E-08	29.9
rs174537	T	G	<i>MYRF</i>	0.035	0.004	3.80E-15	61.8
rs4766214	G	A	<i>LOC105369612</i>	−0.025	0.004	4.88E-09	34.2
rs1871395	G	A	<i>SLCO1B1</i>	−0.059	0.006	1.07E-22	96.1
rs79295634	G	A	<i>SLC38A4</i>	0.086	0.009	1.82E-23	99.6

rs1043011	T	G	<i>GLS2, SPRYD4</i>	0.067	0.006	1.70E-34	150.0
rs12811045	G	A	<i>LOC102723639</i>	0.032	0.005	7.46E-10	37.9
rs2464190	C	T	<i>HNFI1A</i>	0.047	0.004	1.27E-26	114.0
rs4760099	A	G	<i>HPD</i>	0.052	0.006	9.16E-16	64.6
rs78424108	G	C	<i>WBP4, MIR3168</i>	-0.047	0.008	1.19E-08	32.5
rs3783642	C	T	<i>GCHI</i>	-0.024	0.004	3.71E-08	30.3
rs6575900	C	G	<i>WDR20, LOC105370677</i>	-0.042	0.005	5.89E-15	60.9
rs9746832	A	G	<i>LOC105371334</i>	0.087	0.004	2.37E-92	415.5
rs116992380	C	T	<i>PKDIL3</i>	0.206	0.015	3.23E-45	199.1
rs8047723	T	G	/	0.029	0.004	7.98E-11	42.3
rs10221244	G	A	<i>UNK</i>	0.026	0.004	8.30E-09	33.2
rs123698	G	C	<i>PTBP1</i>	0.024	0.004	4.04E-08	30.1
rs58542926	T	C	<i>TM6SF2</i>	0.060	0.008	8.09E-14	55.8
rs814573	T	A	<i>APOC1P1, APOC1</i>	-0.032	0.006	1.18E-08	32.5
rs1883711	C	G	<i>LINC01370, MAFB</i>	0.078	0.013	9.02E-10	37.5
rs1800961	T	C	<i>HNFI4A</i>	-0.085	0.012	3.17E-12	48.6
rs738408	T	C	<i>PNPLA3</i>	0.031	0.005	2.30E-09	35.7

Female

rs4665972	T	C	<i>SNXI7</i>	-0.025	0.004	1.11E-09	37.1
rs1047891	A	C	<i>CPS1</i>	-0.032	0.004	1.15E-14	59.6
rs895893	C	T	<i>RPL31P23, PCCB</i>	-0.037	0.005	1.12E-15	64.2
rs35491981	C	T	<i>LOC100507053</i>	0.045	0.004	2.94E-24	103.3
rs7697204	C	T	<i>ARHGAP10</i>	0.035	0.004	2.53E-15	62.6
rs35261542	A	C	<i>CDKAL1</i>	0.027	0.004	2.24E-09	35.8
rs11961853	T	C	<i>UBQLN1P1, MICC</i>	0.028	0.005	4.84E-08	29.8
rs75661418	G	A	<i>SLC16A10</i>	-0.076	0.012	9.66E-10	37.4
rs12206654	C	T	<i>MFSD4B-DT</i>	0.177	0.006	7.63E-218	992.5
rs56236906	A	G	<i>FYN</i>	0.083	0.014	5.28E-09	34.1
rs35614968	A	G	<i>FYN</i>	-0.109	0.012	1.11E-18	77.9
rs146133919	C	T	<i>FYN</i>	-0.076	0.013	4.92E-09	34.2
rs2189966	C	T	<i>JAZF1</i>	-0.034	0.005	1.79E-12	49.7
rs34346326	C	T	<i>MLXIPL</i>	-0.030	0.005	4.09E-10	39.1
rs330093	G	C	<i>PPP1R3B-DT</i>	0.026	0.005	2.61E-08	31.0
rs28601761	G	C	<i>TRIB1AL</i>	-0.037	0.004	3.83E-20	84.5
rs7902343	T	C	<i>JMJD1C</i>	0.029	0.004	1.83E-13	54.2
rs174555	C	T	<i>FADS1</i>	0.038	0.004	3.29E-19	80.3
rs4149059	T	C	<i>SLCO1B1</i>	-0.041	0.005	5.37E-18	74.7
rs76943648	C	T	<i>SLC38A4</i>	0.078	0.008	6.15E-24	101.8
rs2657879	G	A	<i>GLS2, SPRYD4</i>	0.064	0.005	2.31E-37	163.2
rs2393775	G	A	<i>HNFI1A</i>	0.057	0.004	1.22E-45	201.1
rs372273603	G	A	<i>HPD</i>	0.047	0.006	5.04E-16	65.8
rs1595261	C	A	/	0.025	0.004	1.12E-08	32.6
rs6575900	C	G	<i>WDR20, LOC105370677</i>	-0.031	0.005	3.16E-10	39.6
rs150851429	C	G	<i>LOC105371334</i>	0.389	0.017	3.12E-119	539.0
rs11640725	A	G	<i>APIG1, LOC124903714</i>	-0.046	0.004	9.96E-30	128.2

rs1135688	C	T	<i>UNC13D</i>	0.025	0.004	2.82E-09	35.3
rs1800961	T	C	<i>HNF4A</i>	−0.083	0.011	1.96E-13	54.0

Supplementary Table 9. The outliers detected in MR-PRESSO.

Phenotype	Sex	Outlier
Phenylalanine	Overall	rs73063122
	Female	rs1047891
Tyrosine	Overall	rs183657985, rs4493565

Supplementary Table 10. MR results after exclusion of SNPs significantly associated with potential confounders.

Phenotype	Sex	Method	beta	se	p-value
Phenylalanine	Overall	Inverse variance weighted	−0.05	0.43	0.90
		Weighted median	−0.20	0.37	0.58
		Weighted mode	−0.09	0.35	0.80
		MR-PRESSO	0.17	0.33	0.61
Phenylalanine	Male	Inverse variance weighted	0.76	0.31	0.01
		Weighted median	0.60	0.39	0.12
		Weighted mode	0.60	0.39	0.13
		MR-PRESSO	0.76	0.28	0.02
Phenylalanine	Female	Inverse variance weighted	−0.66	0.60	0.27
		Weighted median	−0.43	0.46	0.35
		Weighted mode	−0.39	0.46	0.40
		MR-PRESSO	−0.36	0.48	0.48
Tyrosine	Overall	Inverse variance weighted	−0.60	0.28	0.03
		Weighted median	−0.14	0.29	0.62
		Weighted mode	−0.15	0.28	0.59
		MR-PRESSO	−0.41	0.25	0.10
Tyrosine	Male	Inverse variance weighted	−0.69	0.32	0.03
		Weighted median	−0.47	0.34	0.16
		Weighted mode	−0.26	0.35	0.47
		MR-PRESSO	−0.69	0.32	0.04
Tyrosine	Female	Inverse variance weighted	−0.68	0.30	0.03
		Weighted median	−0.16	0.36	0.66
		Weighted mode	−0.21	0.33	0.53
		MR-PRESSO	−0.68	0.30	0.03

For phenylalanine, rs34121855 was excluded in the overall analysis, due to its potential pleiotropic effect concerning alcohol intake frequency. For tyrosine, the following SNPs were excluded due to their potential pleiotropic effects: in the overall analysis, rs6754311 and rs7909960 were excluded due to their associations with age completed full time education. Additionally, rs13107325, rs4665972, rs62062797, rs62466318, and rs9972653 were excluded because of their potential effects on alcohol intake frequency. Within the female subgroup, rs7902343 was excluded for its pleiotropic effect with the age of completing full-time education, and rs4665972 was also excluded due to its impact on alcohol intake frequency. In the male subgroup, rs13107325 was removed from the analysis for its pleiotropic effect on alcohol intake frequency.

Supplementary Table 11. Sensitivity analysis on the association of genetically predicted phenylalanine and tyrosine with life years using genetic instruments from a GWAS without UK Biobank participants.

Exposure	Sex	Method	#SNPs	Beta (95% CI)	p-value	outlier
Phenylalanine	Overall	Inverse variance weighted	9	0.35 (−0.74, 1.44)	0.53	
		Weighted median	9	−0.10 (−1.04, 0.85)	0.84	
		Weighted mode	9	−0.12 (−1.14, 0.90)	0.82	
		MR-PRESSO	8	−0.02 (−1.23, 1.20)	0.97	rs28365897
	Male	Inverse variance weighted	9	1.05 (0.03, 2.08)	0.04	
		Weighted median	9	1.00 (0.05, 1.95)	0.04	
		Weighted mode	9	0.94 (−0.06, 1.93)	0.07	
		MR-PRESSO	9	1.05 (−0.15, 2.26)	0.08	
	Female	Inverse variance weighted	9	−0.12 (−1.55, 1.32)	0.88	
		Weighted median	9	−0.19 (−1.31, 0.94)	0.74	
		Weighted mode	9	−0.39 (−1.60, 0.82)	0.53	
		MR-PRESSO	7	−0.19 (−1.09, 0.71)	0.63	rs1047891, rs28365897
Tyrosine	Overall	Inverse variance weighted	19	−0.21 (−2.16, 1.73)	0.83	
		Weighted median	19	−0.31 (−0.90, 0.28)	0.30	
		Weighted mode	19	−0.36 (−0.89, 0.17)	0.19	
		MR-PRESSO	17	−0.67 (−1.24, −0.09)	0.03	rs4149083, rs429358
	Male	Inverse variance weighted	19	−0.52 (−2.03, 0.99)	0.50	
		Weighted median	19	−0.47 (−1.13, 0.19)	0.16	
		Weighted mode	19	−0.47 (−1.09, 0.15)	0.13	
		MR-PRESSO	17	−0.82 (−1.67, 0.03)	0.06	rs1021956, rs117866491
	Female	Inverse variance weighted	19	0.09 (−2.20, 2.38)	0.94	
		Weighted median	19	−0.14 (−0.88, 0.59)	0.70	
		Weighted mode	19	−0.18 (−0.86, 0.50)	0.61	
		MR-PRESSO	17	−0.58 (−1.18, 0.02)	0.06	rs1021956, rs117866491

Supplementary Table 12. Power calculation in the MR analysis on lifespan.

Exposure	Sex	r ²	Case	Control	Sample size	Effect size detected (odds ratio)	Effect size detected (life years)
Phenylalanine	Overall		208,118	181,048	389,166	1.09	−1.72
	Men	0.01	317,652	97,659	415,311	1.10	−2.18
	Women		246,941	165,996	412,937	1.09	−2.23
Tyrosine	Overall		208,118	181,048	389,166	1.06	−1.17
	Men	0.03	317,652	97,659	415,311	1.06	−1.33
	Women		246,941	165,996	412,937	1.06	−1.51

The calculation of r^2 was based on $\beta^2 \times 2 \times (\text{EAF}) \times (1 - \text{EAF})$. The number of cases and controls are from the GWAS of parental lifespan we used in the MR analysis (Pilling LC et al., Aging (Albany NY). 2017; 9:2504–20). The power calculations for the MR analysis were performed using the online tool available at <https://sb452.shinyapps.io/power/>.

Supplementary Table 13. Associations of genetic instruments for phenylalanine with potential confounders in UK Biobank.

SNP	<i>p</i> -value with potential confounders					
	Townsend deprivation index at recruitment	Age completed full time education	Current tobacco smoking	Alcohol intake frequency	Time spent doing moderate physical activity	Time spent doing vigorous physical activity
Overall						
rs1009062	4.08E-01	1.48E-01	7.22E-01	3.80E-01	9.17E-01	6.08E-01
rs1043011	2.76E-01	2.26E-01	3.28E-01	3.60E-02	1.99E-01	3.57E-01
rs1047891	7.47E-01	5.65E-01	7.13E-01	6.55E-01	6.43E-01	5.96E-01
rs10750864	8.60E-01	7.38E-01	1.92E-01	5.31E-01	2.82E-01	8.53E-01
rs10826337	8.67E-01	2.28E-01	3.65E-01	8.92E-01	9.91E-01	6.33E-01
rs117040573	6.06E-01	3.96E-01	3.23E-01	7.11E-01	9.31E-01	8.96E-01
rs12830698	1.07E-01	7.87E-01	5.99E-01	2.05E-01	3.95E-01	9.10E-01
rs13254494	3.63E-02	9.43E-01	2.52E-01	2.56E-01	3.23E-01	5.61E-01
rs1522298	9.87E-01	2.09E-01	9.15E-01	5.10E-01	2.26E-01	5.16E-01
rs17253619	2.55E-01	7.67E-01	7.82E-01	3.43E-01	8.67E-01	7.11E-02
rs1800759	9.50E-02	1.58E-01	1.07E-02	6.18E-02	9.67E-01	4.28E-01
rs2239328	1.78E-01	4.65E-02	4.32E-01	2.89E-01	3.50E-01	5.83E-01
rs34121855	9.48E-01	1.34E-01	4.78E-01	1.45E-09	3.96E-01	7.49E-01
rs3757132	6.20E-02	1.97E-01	9.95E-01	2.89E-01	8.80E-01	1.35E-01
rs61935426	2.01E-01	6.92E-01	1.86E-01	5.99E-01	9.95E-01	9.35E-01
rs73063122	9.59E-01	3.35E-01	7.93E-01	6.59E-03	3.12E-01	3.17E-01
rs75017413	6.30E-01	4.46E-01	2.50E-01	4.75E-01	7.10E-01	6.04E-02
rs870072	6.17E-01	5.12E-01	6.58E-02	7.36E-01	3.44E-01	1.80E-01
rs932316	4.21E-02	6.04E-01	7.37E-01	4.04E-02	5.17E-01	8.98E-01
rs99780	5.22E-03	2.17E-01	1.33E-01	2.42E-01	2.80E-01	3.63E-01
Male						
rs1009062	6.84E-01	3.99E-02	2.48E-01	2.78E-01	7.57E-01	5.05E-01
rs10750864	9.38E-01	2.97E-01	7.82E-01	9.46E-01	2.74E-01	2.26E-01
rs12367892	7.20E-01	7.03E-01	7.30E-01	8.41E-01	6.71E-03	6.59E-01
rs1321250	1.42E-01	9.64E-01	3.54E-01	3.01E-01	6.76E-01	1.55E-01
rs1498691	6.95E-01	9.02E-01	2.29E-01	8.08E-01	6.10E-01	7.84E-01
rs1718292	1.53E-01	7.76E-01	6.69E-01	3.65E-01	9.39E-01	3.96E-02
rs17253619	7.90E-02	6.80E-01	7.19E-01	2.55E-01	2.90E-01	6.11E-02
rs2229742	8.04E-01	7.03E-01	3.07E-01	3.47E-02	4.54E-02	7.23E-01
rs2239327	1.82E-01	3.12E-02	8.02E-01	6.36E-03	1.03E-01	9.26E-01
rs3757132	2.21E-01	7.37E-01	5.54E-02	4.99E-01	5.85E-01	4.30E-01
rs75918019	5.99E-01	8.97E-01	1.45E-01	5.11E-01	3.82E-01	1.69E-02
rs76169231	6.03E-01	2.98E-01	6.38E-01	6.11E-01	4.32E-01	2.08E-01
Female						
rs1047891	5.59E-01	7.24E-01	6.97E-01	8.12E-01	4.00E-01	4.00E-01
rs10750864	9.58E-01	3.27E-01	4.78E-01	3.22E-01	1.93E-01	1.93E-01
rs12830698	7.16E-02	5.36E-01	8.82E-01	2.82E-01	7.36E-02	7.36E-02
rs1408268	8.21E-01	3.74E-01	1.10E-01	6.44E-02	9.39E-02	9.39E-02
rs1498694	8.27E-01	6.17E-01	3.80E-02	2.02E-01	7.24E-01	7.24E-01

rs17253619	4.36E-01	6.12E-01	3.71E-01	2.35E-01	5.94E-01	5.94E-01
rs2239328	9.82E-01	8.96E-01	9.93E-01	4.51E-01	9.77E-01	9.77E-01
rs2694917	3.31E-01	4.91E-01	3.02E-01	1.50E-01	6.98E-01	6.98E-01
rs4149058	8.92E-01	2.97E-01	4.81E-01	5.96E-01	8.76E-01	8.76E-01
rs9804734	6.82E-01	9.49E-02	8.91E-02	6.57E-01	2.65E-01	2.65E-01

A significant association was observed between rs34121855 and alcohol intake frequency in the overall analysis.

Supplementary Table 14. Associations of genetic instruments for tyrosine with potential confounders in UK Biobank.

SNP	<i>p</i> -value with potential confounders					
	Townsend deprivation index at recruitment	Age completed full time education	Current tobacco smoking	Alcohol intake frequency	Time spent doing moderate physical activity	Time spent doing vigorous physical activity
<i>Overall</i>						
rs10027275	4.22E-01	2.30E-02	5.63E-01	8.65E-01	7.62E-01	8.64E-01
rs10164853	5.43E-03	1.33E-01	4.75E-01	7.63E-01	3.17E-01	3.54E-01
rs10217762	1.98E-02	3.76E-03	2.82E-01	1.34E-01	6.48E-01	3.39E-01
rs1043011	2.76E-01	2.26E-01	3.28E-01	3.60E-02	1.99E-01	3.57E-01
rs10750864	8.60E-01	7.38E-01	1.92E-01	5.31E-01	2.82E-01	8.53E-01
rs11263465	9.19E-01	2.89E-01	6.34E-01	1.77E-01	6.03E-01	9.25E-02
rs11614623	7.33E-01	9.79E-01	1.47E-01	1.55E-01	5.68E-01	1.91E-01
rs11643623	7.83E-01	5.02E-01	3.52E-01	1.53E-01	2.93E-01	9.28E-01
rs11706810	2.01E-02	2.29E-02	3.01E-01	1.15E-04	3.83E-01	2.37E-01
rs12212085	6.01E-02	2.20E-02	1.25E-02	6.70E-01	7.28E-01	3.05E-01
rs123698	4.82E-02	8.98E-01	6.81E-01	7.04E-01	4.14E-01	4.17E-01
rs12596084	8.09E-01	8.16E-01	6.93E-01	3.84E-01	4.11E-01	3.00E-01
rs12811045	5.26E-01	6.75E-02	4.21E-01	6.74E-01	4.82E-01	6.15E-01
rs12824518	2.92E-01	8.89E-02	7.39E-01	8.43E-01	4.06E-02	6.61E-02
rs13107325	1.13E-01	6.86E-04	5.26E-05	6.90E-15	7.56E-02	6.82E-02
rs13142887	3.37E-01	1.24E-01	1.41E-01	8.19E-01	3.70E-02	9.90E-01
rs13281892	3.58E-01	9.48E-01	2.48E-01	8.42E-01	3.79E-01	4.60E-01
rs1345901	5.22E-01	5.33E-01	4.90E-01	1.19E-01	9.58E-01	5.72E-01
rs1433210	1.73E-01	5.45E-01	1.10E-01	4.10E-02	7.25E-01	7.86E-01
rs150851429	1.31E-01	5.11E-04	4.15E-01	3.92E-01	8.74E-01	8.21E-01
rs151175127	7.05E-01	4.91E-02	4.26E-01	2.07E-01	1.45E-01	1.39E-01
rs1531022	7.35E-01	9.55E-01	3.23E-01	1.98E-01	6.48E-01	6.56E-01
rs17050272	3.54E-01	9.43E-01	9.45E-02	8.25E-01	3.47E-02	2.13E-01
rs174537	1.53E-02	1.87E-01	1.74E-01	1.45E-01	4.00E-01	4.30E-01
rs1800961	2.23E-01	5.28E-01	3.89E-01	7.53E-01	8.20E-01	4.26E-01
rs183657985	5.15E-03	1.46E-04	4.42E-01	4.37E-04	7.74E-01	9.44E-01
rs1883711	5.45E-02	1.29E-01	3.08E-01	2.49E-02	9.61E-01	4.84E-01
rs194742	6.59E-01	5.53E-01	6.49E-01	6.10E-01	5.21E-01	7.71E-01
rs204926	5.16E-01	8.86E-01	7.45E-02	3.28E-01	8.67E-01	8.46E-01
rs2126263	3.92E-01	8.37E-01	9.10E-03	4.80E-03	7.82E-01	9.72E-01
rs2189966	5.23E-01	9.57E-01	2.26E-01	6.22E-02	1.06E-01	8.13E-01
rs2393775	4.72E-01	5.63E-01	5.15E-01	2.70E-02	7.19E-01	1.17E-01
rs28601761	1.47E-01	5.07E-02	9.22E-01	3.64E-05	3.42E-01	5.10E-01

rs34396849	5.75E-05	7.95E-02	6.57E-04	3.44E-01	9.10E-01	1.08E-01
rs35048664	1.00E+00	2.07E-01	9.25E-01	4.98E-01	2.25E-01	5.17E-01
rs35757209	9.56E-01	3.71E-01	8.82E-01	6.63E-01	8.48E-01	3.00E-02
rs41289886	1.80E-01	1.57E-01	5.29E-01	6.71E-01	2.04E-01	6.73E-01
rs4493565	1.80E-02	1.37E-01	4.81E-02	3.12E-02	7.66E-01	7.26E-01
rs4665972	3.83E-01	3.05E-01	1.74E-01	3.10E-55	3.41E-01	9.50E-01
rs4722551	7.84E-01	7.26E-01	3.27E-01	1.79E-02	8.01E-01	9.23E-02
rs4921914	9.65E-01	1.65E-04	5.17E-01	2.00E-01	9.49E-01	5.61E-01
rs511154	9.79E-01	2.48E-01	7.19E-01	2.46E-01	4.72E-01	7.69E-01
rs56058728	1.33E-01	4.31E-01	2.80E-01	2.33E-01	9.39E-01	6.63E-01
rs56337219	1.69E-01	3.95E-01	2.02E-01	3.05E-02	2.93E-01	1.53E-01
rs56401710	7.95E-01	7.13E-01	1.00E-01	1.26E-04	2.44E-01	9.62E-01
rs61676179	2.82E-01	6.06E-01	5.49E-01	6.93E-01	1.15E-01	5.39E-01
rs62062797	9.63E-01	2.27E-01	1.94E-02	3.09E-11	1.54E-01	3.53E-03
rs62466318	9.36E-01	2.10E-02	2.30E-01	1.44E-11	7.15E-01	8.15E-01
rs6575900	1.56E-01	4.02E-01	3.67E-01	4.12E-01	1.03E-03	3.14E-01
rs6754311	2.02E-01	2.11E-10	5.81E-01	7.68E-02	2.42E-01	1.30E-01
rs6831352	5.83E-02	9.62E-02	7.61E-04	8.25E-04	7.86E-01	5.56E-01
rs6906327	2.03E-01	8.49E-01	7.99E-01	7.48E-01	3.71E-01	9.83E-01
rs715	6.32E-01	6.41E-01	6.19E-01	6.64E-01	7.13E-01	7.73E-01
rs73079476	9.36E-01	3.13E-01	6.90E-01	1.80E-03	3.06E-01	2.31E-01
rs73158176	7.99E-03	8.07E-01	9.87E-01	4.71E-01	7.11E-01	7.25E-01
rs738408	7.65E-03	3.17E-01	6.52E-01	1.63E-02	4.29E-01	9.96E-01
rs7404381	4.49E-03	2.11E-02	5.01E-04	3.45E-01	1.56E-01	6.48E-01
rs7537281	2.43E-01	7.68E-02	3.64E-01	6.54E-03	2.69E-01	2.60E-01
rs77042499	1.92E-02	7.94E-02	5.66E-03	6.52E-01	3.56E-01	9.88E-01
rs78424108	9.02E-02	6.01E-01	4.25E-01	4.56E-01	1.57E-01	6.24E-01
rs78802502	6.19E-01	4.46E-01	2.39E-01	4.61E-01	7.62E-01	6.27E-02
rs7909960	2.69E-01	3.71E-11	1.34E-05	1.80E-01	5.50E-01	2.47E-01
rs79687284	4.32E-02	5.59E-01	1.74E-01	4.12E-01	2.23E-01	7.56E-02
rs8021303	1.93E-01	8.29E-01	1.59E-01	8.30E-01	2.03E-01	1.09E-01
rs8122094	4.36E-01	9.08E-01	9.73E-01	1.64E-04	9.24E-01	7.71E-01
rs9972653	5.78E-01	1.33E-01	5.42E-01	1.83E-09	9.07E-01	7.24E-01

Male

rs10020631	3.37E-01	6.33E-01	2.86E-01	9.27E-01	9.18E-01	5.90E-01
rs10027275	6.33E-01	7.72E-01	2.10E-01	5.41E-02	7.83E-01	5.70E-01
rs10221244	8.65E-01	6.50E-01	2.53E-01	7.39E-01	3.85E-01	1.86E-01
rs1043011	2.09E-01	5.84E-01	5.85E-01	3.60E-01	1.53E-01	4.58E-01
rs1051952	4.67E-01	9.91E-01	1.05E-02	8.31E-01	1.08E-01	4.11E-01
rs10761756	1.37E-01	5.50E-03	9.17E-02	2.71E-01	3.41E-01	2.41E-01
rs116862171	9.54E-01	1.86E-01	2.23E-01	1.72E-02	3.13E-01	7.77E-02
rs116992380	7.85E-01	2.00E-01	6.55E-01	6.78E-01	5.73E-01	9.88E-01
rs12206654	1.08E-02	6.04E-01	1.46E-04	6.69E-01	8.37E-01	6.78E-01
rs123698	6.77E-01	7.31E-01	7.08E-01	2.12E-01	7.47E-01	7.23E-01
rs12614487	1.39E-01	4.70E-01	9.49E-01	4.17E-01	3.60E-01	1.29E-01
rs12632030	1.39E-01	5.51E-02	5.16E-02	1.23E-03	7.97E-01	2.88E-01
rs12811045	7.47E-01	6.10E-02	4.41E-01	2.17E-01	2.18E-01	7.27E-01
rs13107325	1.68E-01	3.25E-03	4.93E-02	5.49E-12	7.76E-01	4.58E-02

rs1531022	6.14E-01	9.52E-01	9.80E-01	2.29E-01	6.26E-01	7.28E-01
rs17050272	9.63E-01	1.29E-01	2.29E-01	5.13E-01	4.13E-01	7.76E-01
rs174537	3.09E-02	6.32E-02	2.29E-01	1.62E-01	3.31E-01	4.46E-01
rs17717962	8.91E-01	1.69E-01	3.42E-01	7.57E-01	7.63E-02	8.59E-01
rs1800759	2.33E-01	2.35E-02	3.49E-03	2.71E-01	2.18E-01	3.56E-01
rs1800961	1.49E-01	6.07E-01	4.53E-01	2.29E-01	3.94E-01	9.91E-01
rs1871395	3.06E-01	3.74E-01	3.22E-01	1.24E-01	6.88E-01	6.84E-01
rs1883711	4.79E-01	9.51E-01	8.20E-01	5.26E-03	9.49E-01	7.16E-01
rs2297644	3.71E-01	1.88E-01	4.44E-01	5.80E-01	1.45E-01	9.89E-01
rs2464190	7.12E-02	5.88E-01	8.99E-01	3.72E-01	8.19E-01	7.08E-01
rs2980888	3.53E-01	4.77E-01	1.13E-01	2.46E-02	1.79E-01	8.17E-01
rs35614968	9.48E-01	9.16E-01	8.74E-02	6.75E-01	8.01E-01	7.76E-01
rs3783642	7.39E-01	8.06E-01	1.69E-02	1.91E-01	1.90E-01	7.80E-02
rs4719841	7.44E-01	9.17E-01	7.96E-01	1.51E-01	9.87E-01	7.10E-02
rs4760099	4.16E-01	1.29E-01	8.34E-01	6.45E-01	4.76E-01	7.44E-02
rs4766214	8.23E-02	4.26E-01	6.95E-01	1.99E-01	6.21E-01	5.22E-02
rs511154	1.02E-01	2.72E-01	3.15E-01	2.24E-01	6.74E-01	3.41E-01
rs58542926	5.20E-01	1.85E-03	5.12E-01	3.45E-01	2.19E-01	8.95E-01
rs62646255	3.53E-01	7.26E-01	5.09E-01	6.42E-04	6.09E-01	2.12E-01
rs6575900	4.22E-01	2.10E-01	3.69E-01	3.43E-01	2.32E-04	1.68E-01
rs72839445	3.77E-03	5.10E-01	8.81E-05	9.95E-01	6.33E-01	8.53E-01
rs73091233	4.76E-01	1.37E-01	4.59E-01	2.39E-01	6.13E-01	4.65E-01
rs738408	4.06E-02	6.35E-01	1.10E-01	3.69E-01	8.91E-02	1.40E-01
rs78424108	2.31E-03	9.21E-01	3.82E-01	2.43E-01	6.69E-01	5.03E-01
rs79295634	4.18E-01	7.96E-01	1.29E-01	9.06E-01	7.53E-02	1.96E-03
rs8047723	7.49E-01	1.66E-01	1.35E-01	8.25E-01	8.57E-01	2.34E-01
rs9987289	9.13E-01	6.10E-02	2.62E-02	7.12E-01	1.41E-01	8.95E-01

Female

rs1047891	5.59E-01	7.24E-01	6.97E-01	8.12E-01	4.00E-01	4.00E-01
rs1135688	4.84E-01	4.03E-01	6.07E-01	3.80E-01	9.52E-01	9.52E-01
rs11640725	2.40E-02	1.90E-03	1.66E-01	4.00E-01	8.47E-01	8.47E-01
rs12206654	7.67E-01	3.85E-01	4.97E-01	3.03E-01	4.35E-01	4.35E-01
rs146133919	9.49E-01	3.07E-01	6.54E-01	2.73E-01	2.37E-01	2.37E-01
rs150851429	9.94E-01	2.08E-03	8.76E-01	1.73E-01	6.60E-01	6.60E-01
rs1595261	4.71E-01	7.72E-01	9.28E-01	6.24E-01	9.97E-01	9.97E-01
rs174555	5.08E-03	7.43E-01	5.62E-01	5.13E-01	2.75E-01	2.75E-01
rs1800961	8.75E-01	9.62E-01	3.47E-01	7.43E-01	4.93E-01	4.93E-01
rs2189966	1.11E-01	6.17E-01	8.64E-01	6.47E-01	7.46E-01	7.46E-01
rs2393775	1.24E-01	8.55E-01	5.90E-02	2.07E-01	1.59E-02	1.59E-02
rs2657879	2.39E-01	1.53E-01	5.06E-01	1.94E-01	3.68E-01	3.68E-01
rs28601761	1.90E-01	5.31E-02	1.72E-01	2.04E-03	3.65E-01	3.65E-01
rs330093	7.26E-01	3.04E-03	9.60E-01	2.46E-01	1.45E-01	1.45E-01
rs34346326	3.56E-01	8.05E-01	3.10E-01	1.33E-05	9.21E-01	9.21E-01
rs35261542	3.39E-01	4.68E-01	3.70E-01	1.09E-01	1.19E-01	1.19E-01
rs35491981	4.99E-01	8.94E-01	2.43E-01	5.44E-01	5.20E-01	5.20E-01
rs35614968	4.62E-01	1.11E-01	4.70E-01	9.63E-01	2.42E-01	2.42E-01
rs372273603	6.15E-01	1.19E-01	2.54E-01	3.27E-01	7.76E-01	7.76E-01
rs4149059	9.03E-01	3.09E-01	5.11E-01	6.02E-01	8.72E-01	8.72E-01

rs4665972	7.50E-01	6.57E-01	3.71E-02	8.95E-21	7.45E-01	7.45E-01
rs56236906	5.05E-02	3.78E-01	1.28E-01	6.92E-01	8.54E-01	8.54E-01
rs6575900	7.37E-01	8.57E-01	7.02E-01	1.89E-01	4.91E-01	4.91E-01
rs75661418	9.41E-01	6.46E-01	2.00E-01	2.40E-01	4.78E-01	4.78E-01
rs76943648	9.59E-01	6.52E-01	4.48E-01	4.42E-01	7.13E-01	7.13E-01
rs7697204	3.62E-01	2.76E-01	3.39E-01	5.81E-01	4.35E-01	4.35E-01
rs7902343	2.70E-02	3.23E-09	2.27E-02	1.50E-01	9.07E-01	9.07E-01
rs895893	3.90E-01	9.92E-01	5.84E-01	3.69E-01	3.92E-01	3.92E-01
